Chemical Carcinogens: A Review of the Science and Its Associated Principles

By the U.S. Interagency Staff Group on Carcinogens*

In order to articulate a view of chemical carcinogenesis that scientists generally hold in common today and to draw upon this understanding to compose guiding principles that can be used as a bases for the efforts of the regulatory agencies to establish guidelines for assessing carcinogenic risk to meet the specific requirements of the legislative acts they are charged to implement, the Office of Science and Technology Policy, Executive Office, the White House drew on the expertise of a number of regulatory agencies to elucidate present scientific views in critical areas of the major disciplines important to the process of risk assessment.

The document is composed of two major sections, Principles and the State-of-the-Science. The latter consists of subsections on the mechanisms of carcinogenesis, short-term and long-term testing, and epidemiology, which are important components in the risk assessment step of hazard identification. These subsections are followed by one on exposure assessment, and a final section which includes analyses of dose-response (hazard) assessment and risk characterization.

The principles are derived from considerations in each of the subsections. Because of present gaps in understanding, the principles contain judgmental (science policy) decisions on major unresolved issues as well as statements of what is generally accepted as fact. These judgments are basically assumptions which are responsible for much of the uncertainty in the process of risk assessment. There was an attempt to clearly distinguish policy and fact. The subsections of the State-of-the-Science portion provide the underlying support to the principles articulated, and to read the "Principles" section without a full appreciation of the State-of-the-Science section is to invite oversimplification and misinterpretation.

Finally, suggestions are made for future research efforts which will improve the process of risk assessment.

Introduction

Background

Over the years, agencies and programs have been established to deal with hazardous substances, with recent focus on the deleterious long-term effects, such as cancer. A detailed analysis of legislative acts concerning toxic substances (Table 1) reveals an evolution of thought, in that each reflects the scientific views that existed at the time of enactment. Consequently, these acts are not uniform in their view of disease, the role chemical substances might play in its incidence, and what ought to be done about potential toxic substances and potential carcinogens.

Purpose of This Report

The purpose of this document is to articulate a view of chemical carcinogenesis that scientists generally hold in common today and to draw upon this understanding to compose, as was done here by senior scientists from a number of federal agencies, a series of general principles that can be used to establish guidelines for assessing carcinogenic risk. Because of present gaps in

understanding, the principles contain judgmental (science policy) decisions on unresolved issues as well as statements of what is generally accepted as fact. There

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Table 1. Federal laws related to exposures to toxic substances.

Legislation	Agency ^a	Area of concern	
Food, Drug and Cosmetics Act (1906, 1938, amended 1958, 1960, 1962, 1968, 1976)	FDA	Food, drugs, cosmetics, food additives, color additives, new drugs, animal and feed additives, and medical devices	
Federal Insecticide, Fungicide and Rodenticide Act (1948, amended 1972, 1975, 1978)	EPA	Pesticides	
Dangerous Cargo Act (1952)	DOT, USCG	Water shipment of toxic materials	
Atomic Energy Act (1954)	NRC	Radioactive substances	
Federal Hazardous Substances Act (1960, amended 1981)	CPSC	Toxic household products	
Federal Meat Inspection Act (1967) Poultry Products Inspection Act (1968)	USDA	Food, feed, color additives and pesticide residues	
Egg Products Inspection Act (1970)			
Occupational Safety and Health Act (1970)	OSHA, NIOSH	Workplace toxic chemicals	
Poison Prevention Packaging Act (1970, amended 1981)	CPSC	Packaging of hazardous household products	
Clean Air Act (1970, amended 1974, 1977)	EPA	Air pollutants	
Hazardous Materials Transportation Act (1972)	DOT	Transport of hazardous materials	
Clean Water Act (formerly Federal Water Control Act) (1972, amended 1977, 1978)	EPA	Water pollutants	
Marine Protection, Research and Sanctuaries Act (1972)	EPA	Ocean dumping	
Consumer Product Safety Act (1972, amended 1981)	CPSC	Hazardous consumer products	
Lead-based Paint Poison Prevention Act (1973, amended 1976)	CPSC, DHEW (DHHS), HUD	Use of lead paint in federally assisted housing	
Safe Drinking Water Act (1974, amended 1977)	EPA	Drinking water contaminants	
Resource Conservation and Recovery Act (1976)	EPA	Solid waste, including hazardous wastes	
Toxic Substances Control Act (1976)	EPA	Hazardous chemicals not covered by other laws, includes pre-market review	
Federal Mine Safety and Health Act (1977)	DOL, NIOSH	Toxic substances in coal and other mines	
Comprehensive Environmental Response, Compensation, and Liability Act (1981)	EPA	Hazardous substances, pollutants and contaminants	

^a Abbreviations used: CPSC, Consumer Product Safety Commission; DOL, Department of Labor; DOT, Department of Transportation; EPA, Environmental Protection Agency; FDA, Food and Drug Administration; DHEW, Department of Health, Education and Welfare (many functions assumed by (DHHS); DHHS, Department of Health and Human Services; HUD, Department of Housing and Urban Development; NIOSH, National Institute for Occupational Safety and Health; NRC, Nuclear Regulatory Commission; OSHA, Occupational Safety and Health Administration; USCG, United States Coast Guard; USDA, United States Department of Agriculture.

has been, however, an attempt to distinguish clearly between the different types of information presented (and also provide to all interested parties specific reference to the analysis upon which they are based).

The principles can serve as the basis for consistent regulatory cancer guidelines that the federal agencies can tailor to meet the requirements of the legislative acts they are charged to implement. Similar documents,

Institutes of Health; Angelo Turturro, National Center for Toxicological Research, Food and Drug Administration; Elizabeth Weisburger, Division of Cancer Etiology, National Cancer Institute, National Institutes of Health; Paul White, Health Sciences, Consumer Product Safety Commission; Alvin Young, Office of Science and Technology Policy, Executive Office of the President, The White House. A number of people were members at different times in the deliberations for various tenures. They included: Andrew Jovanovich, Office of Pesticides and Toxic Substances, Environmental Protection Agency; Carl Leventhal, Office of Science and Technology Policy, Executive Office of the President, The White House; Denis Prager, Office of Science and Technology Policy, Executive Office of the President, The White House; John Todhunter, Office of Pesticides and Toxic Substances, Environmental Protection Agency. This document was edited by R. W. Hart, A. Turturro, and E. K. Weisburger.

with varying degrees of specificity, have been produced in the past; e.g., in the late 1970s and early 1980s several agencies of the federal government produced statements directed toward providing a consistent basis for a general federal cancer policy. This document should be seen then, in the broad view, as part of an ongoing process, on behalf of the federal government, that strives to periodically update and review current understanding of carcinogenesis and the scientific process of how this understanding is utilized.

This document is the result of the combined efforts of senior scientists from the following Federal health-related units, operating in the Interagency Staff Group, under the direction of the Office of Science and Technology Policy (OSTP): Center for Food Safety and Applied Nutrition (CFSAN) (formerly Bureau of Foods), Food and Drug Administration (FDA); Consumer Product Safety Commission (CPSC); Environmental Protection Agency (EPA); Office of the Commissioner (OC), FDA; Food Safety and Inspection Service (FSIS) of the United States Department of Agriculture (USDA); Na-

tional Cancer Institute (NCI), National Institutes of Health (NIH); National Center for Toxicological Research (NCTR), FDA; National Institute of Environmental Health Sciences (NIEHS), NIH; National Toxicology Program (NTP); Occupational Safety and Health Administration (OSHA), Department of Labor (DOL).

Context of This Report

This document is written in light of a decision-making process used by many of the regulatory agencies, which involves the assessment of carcinogenic risks posed by chemical substances. The scientific inputs to these evaluations can be best appreciated by examining the decision-making process in some detail.

Risk can be conceived of as being composed of two aspects, each of which can be addressed by science: hazard and exposure.* Although other definitions have been used historically, this document conforms to present usage, and "hazard" generally refers to the toxicity of the substance (for some toxic endpoint) and is deduced from a wide array of data including: epidemiological evaluation of long-term animal studies, shortterm tests, information on mechanisms, and the results of structure/activity relationships. "Exposure" generally refers to the amount of the substance that people come in contact with. The "risk," in a quantitative risk assessment, is estimated by coupling the results of the exposure and hazard assessments. As either the hazard or exposure approaches zero, the risk also approaches zero.

As a first step in assessing the cancer risk associated with the use of a particular chemical substance, the qualitative evidence that a given chemical substance is likely to be a human carcinogen must be evaluated. In this step, as in the whole process, a number of assumptions and approximations must be made in order to deal with inherent limitations found in the existing data bases. Then, estimates of human exposure, and distribution of exposures likely to be encountered in the population are made. Next, one or more methods for estimating the dose-response relationship at doses below those generally used experimentally must also be evaluated. Finally, the exposure assessment is combined with the dose-response relationship to generate an estimate of risk. The various ways in which these steps are conducted and combined and their attendant uncertainties treated, constitute what is generally referred to as "cancer risk assessment," although an assessment can be terminated at different stages to estimate risk for different purposes.

Some legislation calls for action in the presence of any risk. Other forms of legislation use the concept of unreasonable risk, defined, in some acts, as a condition in which the risks outweigh the benefits: e.g., the Toxic Substances Control Act (TSCA) and the Federal Insecticide, Fungicide and Rodenticide Act (FIFRA). A

spectrum of approved responses, from simply informing the public of a risk through restricted use to a complete ban, may be available to bring the risks and benefits into appropriate balance.

This document does not perform a risk assessment nor does it suggest that one method of cancer risk assessment is better than another; but, rather, it attempts to review the science of chemical carcinogenesis and develops from this review a set of general principles. It is not a comprehensive treatise nor a document written for the lay public, but is rather a semitechnical review which tries to evaluate the impact of scientific findings of the last decade on general assumptions or principles important to risk assessment. This is based upon the belief of the group that elucidation of the basic mechanisms underlying cancer and the identification of cancer-causing agents and conditions, when coupled to research aimed at identifying and evaluating the problems created by such agents, should provide the optimal administrative bases for making sound and reasonable judgments. These overlapping approaches to evaluating the problems of cancer, we believe, should form one of the strongest foundations upon which a technologically based society could base its decisions.

Content of This Report

This document is composed of two major components. The first section presents a set of general principles that may be used by regulatory agencies as they review their own specific guidelines for performing cancer risk assessments. The second major section, "State-of-the-Science," addresses the current state of the science concerning carcinogenesis and cancer risk assessment and forms the underlying scientific base of the "Principles" section.

The general principles, as has been suggested for similar guidelines in the National Academy of Sciences (NAS) report on Risk Assessment in the Federal Government (1), can be useful despite an inherent tendency towards oversimplification and the mixing of scientific knowledge with risk assessment science policy. Many components of the risk assessment process lack definitive scientific bases. Often a choice must be made among several different scientifically plausible options. The above-cited NAS report identified approximately 50 components where the uncertainty in the science requires, in part, scientific judgments and science policy decisions to go from one component to the next to reach a decision. This document attempts to leave the majority of these necessary scientific inferences to the scientific regulatory agencies. However, some are so important and fundamental to all cancer risk-assessment procedures, regardless of the source of hazard or the different statutory provisions of the agencies, that they are included in some of the principles. Where this has occurred we have attempted to indicate, in the wording of the principle itself, that a public health science policy decision is embodied in it. For example, Principle 8 defers to the statement that it is reasonable to treat an

^{*}The European community uses the term "risk" where we have used the term "hazard" and vice versa.

animal carcinogen as if it were a human carcinogen. Many scientists would agree that, while there is a significant amount of evidence to support qualitative animal-to-human extrapolation for carcinogenesis, the evidence falls short of establishing this proposition as a scientific fact. (When determining the response of different species to chemicals, many chemicals appear to be carcinogenic in one species or strain and not in another, even when only rodents are being compared.) Nonetheless, this principle has been accepted by all health and regulatory agencies and is regarded widely by scientists in industry and academia as a justifiable and necessary inference. Despite this effort, it is not possible to draw a sharp distinction in every instance between the bases of the principles which are based solely on science and those which embody a choice based also on science policy. Several of the principles, e.g., Principles 8, 11, and 26, clearly reflect certain aspects of science policy.

The second major section of this document is divided into six sections. Each discusses the current information in an area of science in the cancer risk assessment process.

The first section, Current Views on the Mechanisms of Carcinogenesis, highlights some of the recent advances in our understanding of some of the mechanisms of carcinogenesis. A general consensus has evolved which describes cancer as a multi-stage process involving a variety of events which can include metabolic conversion, initiation, promotion, and proliferation. The section highlights some advances in our understanding that have implications in the way we assess cancer risk. At the same time, the section also identifies some of the vast amount of information that remains to be collected and analyzed.

The second section, Short-Term Tests for Potential Carcinogens, examines short-term testing and the relationship between genetic toxicity and carcinogenicity. Tests for gene mutation, chromosomal aberration, DNA damage and repair, and cellular (morphologic) transformation have provided valuable information on the potential for chemical substances to cause cancer. This field is currently characterized by a great deal of activity, and the data generated must be carefully analyzed and verified before use in cancer risk assessment.

The third section, Long-Term Carcinogen Bioassay, contains an evaluation of long-term animal tests which have been a major factor in past assessments of cancer risk. A review of the usefulness in cancer risk assessment of the data developed via this approach to chemical evaluation is made, and potential limitations and pitfalls of such data relative to extrapolation to human populations are discussed.

The fourth section, Current Views of Epidemiological Methods, examines the current state of epidemiological knowledge relating to human cancer. Data collected during the past 50 years permit the analysis of trends in cancer incidence/mortality and provide information related to the various etiologies of cancer. Recent developments in cancer epidemiology are highlighted and their potential impact on risk assessment pointed out.

Much of the discussion concentrates on the methods to be used in such studies in attempting to overcome past, often cited, limitations of this approach.

In addition to these sections which examine factors associated with hazard assessment, a discussion of exposure assessment is also given. As the fifth section, Chemical Exposure Assessment, points out, the field of exposure assessment has been marked by significant progress in the areas of monitoring data, computer modeling, transformation and transport of chemicals, and laboratory approaches to determining the behavior of chemicals in the environment. At the same time uncertainties remain, and any risk assessment will necessarily be affected by these limitations.

The sixth section, Utilizing Scientific Data in Assessing Human Cancer Risk Associated with Chemical Exposure, discusses hazard or dose response assessment and gives a comprehensive framework for integrating this with hazard and exposure elements as inputs into the qualitative and quantitative risk assessment processes. It depends heavily on all the other sections and attempts to make clear the various approaches that are used in risk assessments, noting both the limitations and advantages of each. Common themes within are the uncertainty, the gaps in data, and the questions of interpretation associated with some aspects of the scientific information used in the risk assessment process. A critical evaluation of this information comprises the State-of-the-Science section and led to the principles listed in the first part. These general principles were developed to provide interim guidance in areas of uncertainty until such time that additional scientific experimentation provides the required information needed to improve estimations of risk in human populations. As a note of caution, it should be recognized that the State-of-the-Science section, taken in toto, provides the underlying support for the Principles section. To read the Principles section without a full appreciation of the material in the State-of-the-Science section is to invite oversimplification and misinterpretation.

Finally, a note is needed, chiefly addressed to specialists on the various topics that are briefly discussed. If reports of a broad nature are to be written at all, and general reports are often necessary to present organized thought on subjects of vital interest, it is inevitable that those who write them must spend less time on a topic than someone who concentrates almost exclusively on one specific interest. Therefore, there are certain to be areas which some authorities think are not discussed enough, others which some feel go on ad infinitum. Also, since the sections are an attempt to provide a balanced account of what is generally accepted, derived from a vast amount of information involving the work of thousands of individuals, citing relatively few, some must be slighted.

Both problems are inevitable in a work of this type. Hopefully, deficiencies in the choice of topic for emphasis or failures to acknowledge contributions have not been egregious.

Principles

Preface

The principles contained herein were derived from the information detailed by the members of the Interagency Staff Group. This section attempts to provide, in a nontechnical form, some important general statements relevant to the evaluation of the role of chemicals in carcinogenesis. These statements are intended to serve as a bridge connecting the basic science with multifaceted process of risk assessment.

Since there are gaps in the information available, differences in evaluations and in scientific opinion may exist about certain of the points highlighted as principles. However, these principles derive from a Weltanschauung utilizing a balanced approach with an appreciation of all elements of the problem, from hazard identification and estimation through exposure and risk assessment. It is clearly understood that new information and newly emerging concepts may modify some of these statements. Indeed, an unstated "zeroth" principle is that regulatory judgments should embody an openness to advances in science and emerging scientific understanding. As a consequence, it is necessary that the process which led to this document be a continuing one, with periodic updates as new advances in science dictate. For the time being however, as a result of an arduous cooperative effort, these statements, we believe, represent an up-to-date summary on a number of important topics.

Principles Derived from the Mechanisms of Carcinogenesis

Principle 1. Carcinogenesis is a multistage phenomenon that may involve the genome both directly and indirectly. These stages of carcinogenesis may be, to varying degrees, influenced by a number of variables such as age at exposure, diet, hormonal status, and intra- and interspecies variability, which should be considered when trying to predict human response to potentially carcinogenic agents (see pp. 209–212, 220–224).

Principle 2. Appropriate *in vitro* and *in vivo* tests can indicate that an agent has a certain action such as genetic toxicity or promotion. Such information is valuable and may be useful in evaluating mechanism(s) of cancer induction. However, in the evaluation of human risk, the attribution of observed findings of carcinogenicity to a particular biological effect must rest upon sound evidence that the effect is responsible for the cancer induction. It must be kept in mind that a chemical may contribute to carcinogenesis in multiple ways (see pp. 220–224).

Principle 3. At the present stage of knowledge, mechanistic considerations such as DNA repair and other biological responses, in general, do not prove the ex-

istence of, the absence of, or the location of a threshold for carcinogenesis. The presence or absence of a threshold for one step of the carcinogenic process does not necessarily determine the presence or absence of a threshold for the whole process (see pp. 217–219).

Principle 4. The carcinogenic effects of agents may be influenced by nonphysiological responses (such as extensive organ damage, radical disruption of hormonal function, saturation of metabolic pathways, formation of stones in the urinary tract, saturation of DNA repair with a functional loss of the system) induced in the model systems. Testing regimes inducing these responses should be evaluated for their relevance to the human response to an agent, and evidence from such a study, whether positive or negative, must be carefully reviewed (see pp. 217–220).

Principles from Tests of Cancer Induction

Principle 5. Short-term tests, such as assays for point mutations, chromosomal aberrations, DNA damage, and *in vitro* transformation are useful in screening for potential carcinogens, reaching a judgment on the carcinogenicity of a chemical, and providing information on carcinogenic mechanisms (see pp. 227–230).

Principle 6. Short-term tests are presently limited in their ability to predict the presence or absence of carcinogenicity and cannot supplant data from long-term animal studies or epidemiological investigations, since the tests do not necessarily screen for all potential means of cancer induction and do not necessarily mimic all reactions that would occur *in vivo*. Additional research is required to improve existing tests and develop ones that identify chemicals which act by genetic mechanisms not yet determined or which act by other, nongenetic mechanisms (see pp. 227, 232–233).

Principle 7. Short-term tests should be carefully selected to ensure they have been adequately validated. Several tests with different endpoints may be required to characterize a chemical's response (see pp. 230–232).

Principle 8. In the evaluation of long-term test results, the term "carcinogen" should be used in a broad sense, i.e., a substance which is capable under appropriate test conditions (Principles 10–13) of increasing the incidence of neoplasms (combining benign and malignant when scientifically defensible) or decreasing the time it takes for them to develop. Careful consideration to the relevant issues cited in Principles 4, 9, and 14 should be given prior to a determination that a chemical is an animal carcinogen (see pp. 234–235).

Deference should be given to the IARC principle: "that in the absence of adequate data in humans, it is reasonable, for practical purposes, to regard chemicals for which there is sufficient evidence of carcinogenicity in animals as if they presented a carcinogenic risk to humans." However, this presumption is evaluated along with other relevant information (Principle 25) in making a final judgment concerning human carcinogenicity and should not foreclose further inquiry into the human relevance of animal carcinogens (see pp. 253–254).

Principle 9. Some experimental animal models ordinarily have high incidences of certain tumors. The evaluation of tumor data from such animals can pose special problems. For example, the interpretation of cancer incidence in some strains of rats with testicular or mammary tumors or in some strains of mice with lung or liver tumors must be approached carefully in the light of other biological evidence bearing on potential carcinogenicity (see pp. 240–241).

Principle 10. Protocols for long-term tests should be designed to achieve an appropriate balance between the two essential characteristics of a biological assay: adequate biological and statistical sensitivity (a low false negative rate) and adequate biological and statistical specificity (a low false positive rate). The absence of biases in selection and allocation of animals between control and treatment groups as regards diet, husbandry, necropsy, pathology, and from insufficient quality control, is crucial (see pp. 234–238, 241–243).

Principle 11. It is appropriate to use test doses that generally exceed human exposure levels in order to overcome the inherent insensitivity of the traditional design of the long-term animal test. The highest dose should be selected after an adequate prechronic study and after evaluating other relevant information, as necessary, to determine the highest dose consistent with predicted minimal target organ toxicity and normal lifespan, except as a consequence of the possible induction of cancer (see pp. 236–239).

Principle 12. The diagnosis of pathologic lesions is complicated and requires judgment and appropriate experience. Diagnoses can differ, depending on the tissues and species involved and can change with time as techniques improve and data on bioassays accumulate. Accurate interpretation of tumor data is contingent upon careful attention to gross observation, tissue sampling, slide preparation and histologic examination. Diagnosis of tumors should be guided by evidence of their histogenic origin and stage of progression (see pp. 240–241).

Principle 13. Appropriate statistical analysis should be performed on data from long-term studies to help determine whether the effects are treatment related or possibly due to chance. These should include a statistical test for trend and a test based on pairwise comparisons, including appropriate correction for differences in survival. The weight to be given to the level of statistical significance (the p-value) and to other available pieces of information is a matter of overall scientific judgment (see pp. 241–243).

Principle 14. Decisions on carcinogenicity of chemicals in animals should be based on consideration of relevant biological and biochemical data. Use of background or recent historical control incidence of tissue specific tumors can be an aid, in addition to concurrent controls, in the evaluation of tumor data. Care should be exercised when combining different control groups.

Evidence of probable reproducibility is important. This evidence can consist of independent confirmation of the original findings or may be derived from intergroup comparisons of tumor incidence data, between dose groups, sexes, strains or species. Evidence of dose response increases confidence that the effect is treatment-related; similarly, lack of an observed dose response may reduce the likelihood that the effect is associated with the treatment.

Confidence is increased when: (1) the incidence of tumors is markedly elevated in the treated groups compared to controls; (2) tumor incidence is significantly increased at multiple anatomical sites; and (3) tumor latency is significantly reduced. In addition to tumor incidence at specific sites, the stage in the development of neoplasia should be evaluated. For example, the finding that the majority of neoplastic lesions at a specific site is more advanced in a treated group compared to its control may provide additional evidence of a treatment related effect. Conversely, the finding that the control group lesions are more advanced might argue that a marginal elevation of tumor incidence is not treatment-related. The incidence of preneoplastic lesions in treatment or control groups may, in certain instances, provide evidence for the biological plausibility of a neoplastic response and contribute to the interpretation of a bioassay. Identification of effects from prechronic studies on the target organ(s) can aid in the evaluation of long-term studies. Information on the activity of chemicals at the physiological, cellular and molecular level may be important to the evaluation of carcinogenicity data on a case-by-case basis (see pp. 238–243).

Principles for Epidemiology

Principle 15. The major strength of the epidemiological method is that it is the only means of assessing directly the carcinogenic risk of environmental agents in humans. However, the observational (nonexperimental) nature of most epidemiological studies, as well as the frequent paucity of relevant data, can impose serious limitations on the method (see pp. 243–244).

Principle 16. Descriptive epidemiological studies (based on the measurement of disease rates for various populations), including correlational studies (in which the rate of disease in a population is compared with the spatial or temporal distribution of suspected risk factors), are useful to generate and refine hypotheses, or provide supporting evidence in evaluating relationships detected by other means, but rarely, if ever, provide information allowing a causal inference (see pp. 244–245).

Principle 17. Well designed, conducted, and evaluated analytic epidemiological investigations of either the case-control or cohort variety can provide the basis for causal inferences especially useful for public health decisions (see pp. 245–246).

Principle 18. Elements in interpreting the likely causality of epidemiological observations include the magnitude of the risk estimates (strength of associations); the possibility of their being due to chance (statistical significance); the rigor of the study design to

avoid various kinds of bias, including those related to selection, confounding, classification and measurement; dose-response relationships; the temporal relationships between exposure and disease; the specificity of the associations; their biological plausibility; and the reproducibility of the findings (see p. 244).

Principle 19. A high-quality negative epidemiological study, while useful, cannot prove the absence of an association between chemical exposure and human cancer. Within the scope of the study, specifically for the populations studied (including concomitant exposures), for the levels and durations of exposure to the agents evaluated and for the time assessed following exposure, a likely range can be determined for the estimates of risk and the statistical likelihood of the study to detect an effect can be assessed (see p. 247).

Principles for Exposure Assessment

Principle 20. It is desirable that exposure routes employed in animal health effects studies are comparable to human exposure routes both for the simplification of risk assessment and because there may be important route-dependent differences in molecular, biochemical, and physical parameters in organs (see pp. 209–212).

Principle 21. At present, a single generally applicable procedure for a complete exposure assessment does not exist. Therefore, in the near term, it is expected that integrated exposure assessments (utilizing monitoring data, results from physical and chemical models, and considerations of all routes of exposure through all media) will be conducted on a case-by-case basis (see pp. 248–251).

Principle 22. The depth and accuracy of an exposure assessment should be tailored to provide the degree of knowledge required to support analytical needs. A preliminary assessment using available crude data can often shed light on the upper or lower bounds of potential risks (see pp. 248–251).

Principle 23. An exposure assessment should describe the strengths, limitations and uncertainties of the available data and models and should indicate the assumptions made to derive the exposure estimates (see pp. 251–252).

Principle 24. In general, an array or range of exposure values is preferable to a single numerical estimate (see pp. 251–252).

Principles for Risk Assessment

Principle 25. Decisions on the carcinogenicity of chemicals in humans should be based on considerations of relevant data, whether they are indicative of a positive or negative response, and should use sound biological and statistical principles. This weight of evidence approach should include consideration of all relevant factors and should give appropriate weight to each on a case-by-case basis. Examples of the types of infor-

mation that should be taken into account include: findings from long-term animal studies (see Principle 14); results from epidemiological studies (see Principles 16–19); results from *in vivo* and *in vitro* short-term tests (see Principles 5 and 6); and data from studies of mechanism, including factors such as structure-activity relationships, and known similarities and differences in metabolic and kinetic profiles for different species (see Principles 1–4 and pp. 253–254).

Principle 26. No single mathematical procedure is recognized as the most appropriate for low-dose extrapolation in carcinogenesis. When relevant biological evidence on mechanism of action (e.g., pharmacokinetics, target organ dose) exists, the models or procedures employed should be consistent with the evidence. However, when data and information are limited, and when much uncertainty exists regarding the mechanisms of carcinogenic action, models or procedures which incorporate low-dose linearity are preferred when compatible with the limited information (see pp. 255–258).

Principle 27. The quantification of the various sources of uncertainty involved in cancer risk assessment can be as important as the projection of the risk estimate itself. The sources that might be addressed include the statistical uncertainty associated with the given risk estimate (often expressed as upper and lower confidence bounds); the variability introduced by the selection of a particular low-dose extrapolation procedure (often expressed as an envelope of risk estimates from a variety of plausible models); when risk estimation is based on laboratory-generated data, the biological variability associated with the use of a particular test organism and its scaling or extrapolation to man (see pp. 257–258).

Principle 28. An estimate of cancer risk for humans exposed to an agent can be no more accurate than an exposure assessment that it utilizes. Lack of adequate exposure data is frequently a major limiting factor in evaluation of carcinogenic risks for humans (see Principles 20–24 and p. 254).

Principle 29. While several considerations often enter the risk assessment process, it is important to try to maintain a clear distinction among facts (statements supported by data), consensus (statements generally held in the scientific community), assumptions (statements made to fill data gaps), and science policy decisions (statements made to resolve points of current controversy) (see pp. 257–258).

Principle 30. Differences in human susceptibility, and variable and extreme exposures to chemicals suggest the likelihood that there are subpopulations that are at greater than average risk. Consideration should be given to the identification of high risk populations (see pp. 209–212, 251).

Principle 31. Because of the uncertainties associated with risk assessment, a full evaluation of risk to humans should include a qualitative consideration of the basic strengths and weaknesses of the available hazard and exposure data, in addition to any numerical estimations that are made (see pp. 257–258).

State-of-the-Science

This segment contains the details about the topics emphasized in Part I, and, as such, is a summary of the current science in a number of fields important to adequate risk assessment. Special emphasis has been placed on those areas considered to be particularly relevant to estimation of hazard and risk.

There are six sections: one on the mechanisms of cancer induction is followed by two on testing, one each for short-term and long-term testing. These disciplines important to understanding and estimates are rounded out by a chapter concerning estimation of human hazard, i.e., epidemiology. Following, there is a chapter on exposure assessment, a critical problem in assessing human risk. The last section discusses hazard or doseresponse assessment and risk characterization as well as drawing on all the other chapters to discuss risk assessment as a whole. It is clear that the limited space available in this review does not allow a totally comprehensive treatise on the various subjects involved, and specialists in the various fields may disagree with the importance of certain aspects of the discussion. The focus has been on a balanced presentation of what is consensus in the various fields discussed as they intersect with the need to understand the effects of chemical agents.

Current Views on the Mechanisms of Carcinogenesis*

Introduction

PHILOSOPHICAL PERSPECTIVES. Underlying the elucidation of mechanisms in carcinogenesis is the search for causality in this biological phenomenon in order to better evaluate and control it. Since science is continually advancing, our understanding must be periodically re-evaluated in light of new, generally agreed upon findings. Such a review is especially important in rapidly advancing, highly specialized and interdisciplinary areas that are of significance to public health. The mechanisms underlying induction of cancer represent such an area and are re-examined here in light of the findings of the last few years.

This section first places the understanding of the mechanisms of cancer induction in a historical perspective and provides a synopsis of some current commonly held beliefs. The bulk of the text follows, which focuses on the interaction of cancer-related agents with factors both external and internal to an organism and the consequences of such interactions at the molecular, cellular, tissue and organismic levels relative to the development of cancer. The sequence of events presented is not written to advance any specific mechanism of cancer induc-

tion, but simply to organize the presentation of the response of various levels of biological organization to cancer-related agents. It should also be appreciated that the data presented have been obtained from studies using various research model systems (e.g., cell-free, cellular, whole animal) to isolate various components of the process. They have also been derived from various strains and species treated under a great variety of experimental conditions. This comprehensive approach to the study of cancer is generally accepted for many reasons, primarily the belief that, while biological systems vary, the degree of similarity at the level of their basic processes is greater than at higher levels of integration.

HISTORICAL PERSPECTIVES. Studies in paleopathology have indicated that cancers occurred in prehistoric times (2). The frequency of occurrence of the disease (its incidence) is hard to quantitate in antiquity, however, over approximately the last century, a relatively well-documented period, total cancer incidence per capita has increased (3). The increase in the mean life span as a result of prevention and treatment of other diseases is a significant factor in this increase in incidence, however exposure to carcinogens (e.g., by smoking) is also important (3). Less clear are the effects of other factors, e.g., increasing urbanization (3,4), which would result in reducing exposure to certain agents (e.g., mycotoxins, mutagens in raw sewage), while increasing exposure to others (e.g., asbestos, benzene). The study of the process of carcinogenesis is at least 300 years old, with studies during the first 200 years being primarily descriptive. The influence of occupation and lifestyles in cancer occurrence was first observed by Ramazzini in 1700, who noted that nuns exhibited a higher frequency of breast cancer than other women (5). Subsequently, in 1761 Hill associated the use of tobacco snuff with cancer of the nasal passage (6), and in 1775 Pott noted the occurrence of soot-related scrotal cancer in chimney sweeps (7). Based upon observations in developing systems, Weismann, in 1881, suggested an association between the "germplasm" (the reproductive or life principle) and cancer (8), one of the first experimentally based theories concerning the mechanism of cancer induction. In 1895 Rehn published evidence that aromatic amines were associated with bladder cancer (9), while Unna, in 1894, and Dubreuilh, in 1896, associated sunlight exposure with skin cancer (10,11). In the following 50 years, with the development of appropriate models, numerous agents were found to induce cancer in experimental animals. Over the last 30 years, there has been an increased emphasis on understanding the mechanisms of carcinogenesis and, recently, on the role of factors such as diet in the enhancement or inhibition of tumor formation (3,12-14). Consequently, in the last decade, more progress has been made in understanding the overall mechanisms of carcinogenesis than at any other time in history.

Current Beliefs

From the mass of data on carcinogenesis, a consensus has emerged on a number of important scientific issues.

^{*}Although the full committee participated in the framing of this section, primary responsibility was assumed by Drs. R.W. Hart and A. Turturro.

First, cancer can be induced by radiation, biological, "physical" and/or chemical agents.

Second, on a biochemical and molecular level, there are important similarities among mammalian species.

Third, an estimate of the potency of carcinogens may never be exact and may vary with lifestyle, habits, age, sex, individual genetic differences, ethnic background, test strain and/or species, diet, dose rate, route of administration, vehicle or solvent used (if any) as well as the presence or absence of other agents, and the environmental conditions prior to, during, or after exposure.

Fourth, cancer development is a multistage process that may involve the genome, both indirectly (frequently termed epigenetic events) and directly, which may include the participation of chemicals or viruses, and which may be modulated by higher order functions, i.e., at the organ and organismic level.

Fifth, numerous factors may alter the frequency of cancer induction by altering one or more of these stages.

Sixth, the genesis of a cancer appears to require an alteration in the ability of a cell to elaborate its appropriate genetic program, i.e., in its information processing capacity, with the subsequent fixation and propagation of that alteration.

Seventh, we still lack an in-depth understanding of the mechanisms and stages of cancer induction and expression.

Eighth, only by understanding the stages of tumorigenesis and carcinogenesis, the substances and processes which modulate them, and how these may differ among cells, organs, individuals, strains and species will we ultimately understand the role of substances, radiations, viruses and/or life-style factors in human cancer.

Preabsorption Modification of Carcinogens

Human exposure to carcinogens usually occurs by dermal contact, inhalation, ingestion, or a combination of these routes of exposure. Depending on route of exposure, an agent can be altered as a result of interaction with interactions that are in association with, but not actually part of, an organism. These factors include: (1) direct transformation of chemicals by environmental factors [e.g., solar radiation interacting with an agent on the surface of the skin (15)]; (2) interaction with cellular secretions [e.g., certain inhaled carcinogens being "coated" with lung secretions (16)]; and (3) metabolic conversions by microorganisms [e.g., gut microflora acting on ingested agents (17-24)]. An illustration of the impact that these factors can have is exemplified by the role of the gut microflora in the metabolism of xenobiotics. It has been suggested that the potential for metabolism in the gastrointestinal tract is similar to that of the liver (20). However, hepatic metabolism of nonnutrients is predominantly oxidative and synthetic (conjugations), while reactions by intestinal microflora are mostly of a hydrolytic and reductive nature (20). It also appears that intestinal microbial metabolism is most important for polar compounds that are not well absorbed from the gut and for those that are excreted, free or conjugated, in the bile (20). Examples of this

include the metabolism of cycasin to a carcinogenic aglycone (19) and enzymatic reduction of a number of azo dyes by numerous genera of anaerobic gut bacteria (24). Intestinal microfloral metabolism can give rise to competition or cooperation between mammalian and microbial enzyme systems in sequential reactions in the cells and in the enterohepatic circulation.

Intra- and interspecies comparisons of microfloral metabolism of xenobiotics have been rare. In one study, rat, monkey, and human intestinal microflora exhibited the capacity to reduce several benzidine and benzidine congener-based dyes to potentially carcinogenic aromatic amines under test conditions (17). Since there is a similarity between the biological effects of these dyes and the effects of benzidine on humans (25), this study suggested that, for these dyes at least, gut microfloral metabolism may play a significant role in the etiology of urinary bladder cancer. Diet composition has also been shown to affect microbial metabolism. Diets with high pectin content can alter the metabolic activities of bacteria in the gut, thus directly effecting hepatic covalent binding of carcinogens (26). Further studies in this area, especially in regard to the relation between gut microflora in animals with different feeding habits (e.g., carnivore, herbivore, and omnivore) and the animal's susceptibility to certain classes of carcinogens, would be of interest. Species and individual differences in microfloral composition could quantitatively and qualitatively alter the nature of the agent to which the animal is ultimately exposed and, thus, the biological response.

Organismic and Cellular Metabolism

ACTIVATION, DETOXIFICATION, AND REACTIVA-TION. Carcinogens include biological agents, such as some viruses (27), radiations, such as X-rays and ultraviolet light (28), "physical agents," such as plastic surfaces and wounding (29), chemical agents, both organic and inorganic (30), and combinations thereof. Chemical carcinogens occur in a number of chemical classes including polycyclic aromatic hydrocarbons and their derivatives, aromatic amines, azo dyes, nitrosamines, nitrosamides, halogenated hydrocarbons, alkylating agents, and metals, their salts and metal complexes, as well as in plant and microbial products (30). Ordinarily, not all chemicals belonging to any class are carcinogenic, nor are all those compounds within a class which exhibit carcinogenicity equally potent. Carcinogens may be naturally occurring (31,32), as in food (33), may result from industrial processes (32,34,35), or may represent the consequences of social activities (36). With the exception of a few direct-acting agents, the organic carcinogens (37) and, possibly, some of the metal complexes (38) require metabolic activation in order to exert their cancer-inducing properties. In general, by whatever route chemicals enter the body, the enzymes involved in biotransformation of a chemical to carcinogenically active (as well as to a number of carcinogenically inactive) metabolites are part of the same mechanisms responsible for detoxication of drugs. Included in this interpretation of the detoxication mechanism is the partitioning of a compound and its metabolites into various body compartments and the effects of differing physiology (blood pH, blood flow to various organs, etc.) on the distribution of the compound and its metabolites. For example, the prolonged retention of acidic urine in dog and man, compared to the rat, can result in a very different profile of aromatic amine metabolites than in the rat, due to the effect of a lower urine pH over a longer time on N-hydroxyarylamine conjugates (39). Realistic modeling can be very useful to understand the mechanisms of the effects of an agent. However, a detailed analysis (e.g., pharmacokinetics) can become quite involved, and the simplifying assumptions used in deriving tractable equations may be inimical to realistically modeling the *in vivo* situation. The data required for modeling are often unavailable, and care should be taken in defining the "effective dose" when the mechanism(s) of carcinogenic action is (are) unknown (40). Also included in considering metabolism is the total metabolic potential of the organism. For instance, nasal epithelium can activate chemical carcinogens (41), a consideration especially important to understanding the effects of exposure by inhalation.

The enzyme systems involved in biotransformation can be divided for convenience into two groups that convert a compound into either a phase 1 or phase 2 metabolite. Phase 1 metabolites are primary oxidation or reduction products, which are usually more watersoluble than the parent compound and thus may generally be more easily eliminated from the body. These metabolites may be then conjugated with various soluble intracellular constituents, forming glucuronides or sulfates, to become even more water-soluble and available for excretion from the body. These conjugated metabolites constitute the phase 2 metabolites (42), which are usually excreted. However, deconjugation prior to excretion may release an active carcinogen at a site distant from the site of conjugation (39,43).

The reactions that occur in the formation of either phase 1 or phase 2 metabolites require specific enzymes for their conversion (some of the better characterized of which are listed in Table 2). Interactions between a chemical carcinogen and these metabolizing systems may form a large spectrum of phase 1 and phase 2 metabolites. A generally accepted mechanism for the reaction of metabolically activated agents with the cell has been proposed by Miller and Miller (37), namely, that the ultimate carcinogenic forms of organic chemical carcinogens are electrophilic (electron deficient) reactants that bind with target intracellular nucleophilic (electron-rich) macromolecules such as DNA and proteins. The reactive metabolites formed may also react with other nucleophiles such as glutathione or water (44,45). Through these latter processes, the effects of agents can be neutralized by forming less biologically reactive metabolites that are very polar and may be more easily excreted. The efficiency of this neutralization is an important factor in tumor induction. For example, if detoxification by glutathione is efficient, the effective local concentration of the active metabolites of the agent is usually reduced, which results in decreased

binding to macromolecules. However, the electrophilic properties of some chemicals may be enhanced by glutathione conjugation (46) (although this is exceptional). Thus, determining the details of chemical biotransformation is desirable when assessing the impact of a chemical.

Understanding the metabolism of chemical carcinogens is often complicated by the diversity of the enzymatic reactions. For example, while for aromatic amines and related compounds, the N-hydroxy derivatives may be the only phase 1 metabolites needed for generation of the nitrenium ion-species that bind with target intracellular macromolecules (39,47), the common metabolic activation of nitrosamines and nitrosamides occurs via the formation of diazo intermediates, which subsequently decompose to molecular nitrogen and active alkyl carbonium ions (30). To illustrate the complexity of metabolism of a chemical carcinogen, most of the known metabolic pathways and the phase 1 metabolites formed from benzo(a)pyrene, one of the most ubiquitous of environmental animal carcinogens, are shown in Figure 1. Many of these metabolites can be converted to different conjugates by phase 2 enzymes. Also, the enzymes cytochrome P-450 and epoxide hydrolase exhibit high selectivity for a particular molecular region and its local conformation (48). This results in arene oxides, trans-dihydrodiols, and trans-dihydrodiol-epoxides being formed as specific optically active isomers (enantiomers). These enantiomers exhibit markedly different biological activities (49).

Simply knowing the identity of the major metabolites of a compound may not always allow prediction of its hazard. For example, metabolism of polycyclic aromatic hydrocarbons to trans-dihydrodiols was long known, but thought to be a pure detoxification process (50). However, identification of the anti-isomer of the dihydrodiol-epoxide of benzo(a)pyrene, which is derived from the trans-7,8-dihydrodiol (Fig. 1), as the electrophilic metabolite of benzo(a)pyrene which binds DNA (ultimate carcinogen) has proven that some trans-dihydrodiol metabolites can be enzymatically activated (51,52). Additionally, detoxified metabolites formed in the liver may be reactivated in other organs, e.g. the bladder (39). The metabolic fate of a compound is a function of a number of processes occurring together (activation, deactivation, and reactivation), whose dynamic interaction results in the local concentration and lifetime of the active metabolite(s). The combination of whole animal, organ, and cellular processes results in a local variable dose rate of active species with time. These changes may be significant factors in producing the biological effects seen with a particular agent.

MODULATION OF METABOLISM. Compound metabolism is a function of the processes in tissues described above, in combination with the cellular, organ and whole animal distribution and excretory processes. This dynamic interaction can be modulated by a number of factors. Such factors include enzyme inhibitors, enzyme inducers, metabolite scavengers (see below), starvation, age, sex, stress, tissue ablations, nutritional fac-

Table 2. Some enzymes needed for phase 1 and phase 2 reactions.

Phase	Enzyme(s)	Catalysis		
Phase 1	Enzyme systems containing cytochrome P-450 and the FAD-containing monooxygenase. This system occurs mainly in the endoplasmic reticulum of liver, kidney, lung, and intestine. The enzymes may occur in multiple forms and exhibit different or overlapping substrate specificity.	Epoxidation of aromatic rings or olefinic double bonds, hydroxylation of aromatic rings or alkyl chains; oxidative dealkylation and N-oxidation.		
	Epoxide hydrolase	Hydrolysis of arene oxides and alkene oxides into $trans$ -dihydrodiols		
	Dehydrogenases, flavin-containing cytochrome P-450 reductases and xanthine oxidase	Reduction of nitroaromatics to nitrosoarenes, N -hydroxyarylamines, and arylamines; reduction to azo compounds to amines		
Phase 2	Glutathione S-transferases (these enzymes are known to exist in multiple forms)	Conjugation between glutathione and a variety of electrophilic compounds such as arene oxides		
	UDP-glucuronyltransferases (also found in multiple forms)	Conjugation reactions between glucuronic acid with substrates such as phenol and bilirubin		
	Sulfotransferases	Sulfate ester formation		
	N-acyltransferases and N , O -acyltransferases	N-acetylation and N , O -acyltransfer of aromatic amines and arylhydroxamic acids		

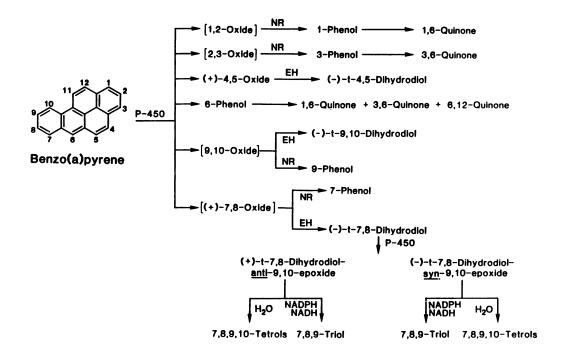


FIGURE 1. Metabolic pathways and metabolites of benzo(a)pyrene obtained from metabolism by rat liver microsomes. Abbreviations: P-450, cytochrome P-450 containing enzymes together with cofactors; EH, epoxide hydrolase; NR, nonenzymatic rearrangement; t, trans.

tors, individual genetic differences, and hormonal status (45,53). Various synthetic chemical agents have been shown to act as inhibitors, inducers, or scavengers (54), and this diversity of effects has been suggested as well for some commonly eaten foods, which contain naturally-occurring inhibitors or inducers, and which also appear to modulate metabolism (3,33). Briefly, there are three general mechanisms accounting for the modulation of metabolism by inhibitors or inducers (54). (1) Stimulation of cellular metabolism results in an in-

creased deactivation and/or activation. For example, a wide range of compounds including polycyclic aromatic hydrocarbons, phenobarbital, flavones, halogenated hydrocarbons, and indoles induce an increase in monooxygenase activity and changes in the relative proportions of the different isozymes. Other enzymes, such as glutathione S-transferases may simultaneously be enhanced. Affinity constants may be altered. As a consequence of any and all these changes, the relative proportions of various metabolites may change, thereby

influencing the ratio of metabolic activation to deactivation (55). (2) Interference with enzymatic activation of chemical carcinogens occurs to form the ultimate carcinogenic metabolite(s). For example, disulfiram is thought to inhibit dimethylhydrazine-induced neoplasia of the large bowel by directly reducing the rate of conversion of the agent into a reactive metabolite (54). (3) Scavenging (neutralization) of the reactive metabolites is altered by nucleophiles, such as glutathione or other defense systems, such as superoxide dismutase. For example, starvation, other nutritional imbalances, or compounds such as butylated hydroxyanisole may drastically alter glutathione levels (56,57). Decreases in glutathione levels may lead to a decreased proportion of the reactive metabolite being detoxified. Conversely, the increased levels of glutathione and glutathione transferase produced by butylated hydroxyanisole may indicate greater detoxification of a reactive metabolite, provided that the glutathione conjugate products are not reactivated.

However, it is the balance between the activation, detoxification and reactivation systems for a chemical carcinogen, and their interaction over time, rather than the change in one of these processes per se, which is important. If monooxygenase activity has been stimulated, resulting in increased reactive metabolite formation, a greater proportion of biologically available carcinogen will result only if the detoxification mechanisms cannot compensate. [An example of compensation using another toxic endpoint involves low doses of acetaminophen, whose reactive metabolite reacts with glutathione, and which will not be hepatotoxic (58). However, larger doses will deplete the glutathione pool and result in hepatotoxicity.] Alternatively, if glutathione levels are decreased, concentrations of electrophilic metabolites may increase.

The use of various chemical modulators has contributed significantly to our understanding of the metabolism and disposition of chemical carcinogens. With extended research into the mechanism of action of these agents, the use of such modulators may, in the future, aid in the treatment of individuals accidentally exposed to high doses of certain chemical carcinogens. However, at present, it would be premature to state that modulators will be generally effective in preventing cancer induction by some chemicals or that such agents could ever be used as an alternative to reducing or preventing exposure to identified carcinogens. An indication for caution is the demonstration that co-administration of aspirin, which modulates the metabolism of N-[4-(5-nitro-2-furyl)-2-thiazolyl]-formamide (FANFT) (a potent rat urinary bladder carcinogen) will decrease the number of bladder tumors induced by FANFT, however, at the same time, it will increase the number of forestomach tumors produced (59).

ORGAN AND SPECIES DIFFERENCES. Because the total enzyme level and substrate specificity of the phase 1 and phase 2 metabolizing enzymes exhibit organ, individual, and species differences (53), the metabolic patterns of both activation and deactivation of a particular

agent could also differ markedly. An example of this is the differences in the monoxygenase activities in nasal epithelium in different species (60). Also, there exist multiple forms of cytochrome P-450 enzymes, each of which exhibits its own specificity toward the substrate metabolized (45). Since glutathione S-transferases and UDP-glucuronyltransferases also occur in multiple forms (45), their particular form in different tissues, strains, individuals or species may also vary. Moreover, the activity of each metabolizing enzyme in a specific tissue of the same strain could vary widely depending on age, sex, nutritional factors, hormonal status, or other factors (32). For these reasons, unless one knows the rates of the activation-deactivation processes and the participating isozymes responsible for these reactions and their levels, it is impossible to predict quantitatively the metabolic activation of chemical carcinogens from an in vitro to an in vivo system, or from laboratory animals to humans. Metabolic comparisons between animal species and humans, when the metabolic pathways are similar, could be useful for qualitative assessments of the relevance of a specific animal test system to human risk. Presently, however, a paucity of comparative metabolism data on either induced or noninduced systems exists since most studies are based on an ontogenetic rather than a molecular-evolutionary approach (61). They, therefore, are of limited use for extrapolation of data between species and organs. There is also expected to be, because of these variabilities, a wide range of responses in different members of a species, leading to individuals with different levels of sensitivity to agents.

Macromolecular Interactions

ALTERATIONS IN GENETIC INFORMATION. Modification of the information content of cells is believed to be one of the events involved in the onset of carcinogenesis (62). Although macromolecules such as RNA (37), protein (63), and lipid-containing membranes (64) will be altered by many carcinogens, and may be important at different stages in carcinogenesis, it is believed that the primary initial site for cancer induction is the DNA. The evidence for this hypothesis rests on a number of observations: (1) the great majority of biological carcinogens that have been examined interact with the DNA of the host cell (65); (2) many, but not all, radiations and chemical carcinogens are mutagens, and mutations result from alterations in the DNA (66); (3) individuals with certain genetic diseases which are characterized by defective DNA repair, such as xeroderma pigmentosum, are prone to cancer at a number of sites and cells from these individuals have increased sensitivity to several radiations and chemical mutagens (67-69); (4) ultraviolet (254 nm) light-induced tumor formation in the gynogenetic fish, Poecilia formosa, is directly related to the amount of DNA damage remaining after different levels of photoreactivation repair of the DNA (70). Factors which influence the expression of cancer cells may or may not act on the DNA during different stages. These factors will be referred to throughout the text, but especially in the subsequent sections on promotion, progression and multiple agents.

The changes in information content of DNA can take many forms such as integration into DNA of viral information for biological carcinogens, chromosomal alterations with radiations, and changes in the base sequence after replication as a result of adduct formation with many chemical carcinogens.

Biological Carcinogens. Many viruses will induce cancer in various organs of one or more species (71). The most progress made recently in understanding the mechanisms of viral carcinogenesis relevant to human cancer involves onc genes, expression control elements, dominant transforming genes, and human viruses.

Studies on oncogenic RNA viruses have provided the background for detailed investigations of genes in the cell with oncogenic potential and for development of methodologies enabling identification and isolation of dominant transforming genes in human tumor cells (72-85). This approach has helped identify genetic elements that may be responsible for certain types of cancer. A major class of RNA tumor viruses, known as retroviruses, has been particularly useful in providing new insight into the cancer process. Retroviruses are unique among animal viruses in their mode of transmission and their intimate association with a wide variety of vertebrate species (86), factors which may have allowed them to become oncogenic in a number of species. Certain members of this group, the acute transforming retroviruses, appear to have arisen by recombination of the viral genome with cellular genes. The acquisition by these viruses of cellular genes, termed onc (for oncogenic) genes (87), is associated with the ability of these viruses to induce rapid neoplastic disease in newborn animals and to transform cells in culture, an in vitro model for carcinogenesis. These genes (the number isolated, at present about two dozen, is increasing) appear to be of major importance to the cells since some have been conserved in widely divergent species (88). The RNA tumor viruses containing them can be thought of as vectors that carry isolated cellular-derived genes critical for malignant neoplastic disease.

The genes in the normal genome of a cell which appear to have given rise to the onc genes, presumably as a result of viral infection, are termed proto-oncogenes. There are human genes homologous to retrovirus onc genes (89-93) and specific onc genes can be identified in many different human cancer cells (94). Some onc genes are rarely expressed, some are expressed chiefly in specific kinds of cells derived from malignant tumors, and some appear to be expressed only in cells of certain lineage or differentiation states (95,96). The specific cellular functions coded by most onc genes are unknown, although a variety of activities have been found. For example, the gene product of one family of oncogenes, src, is a protein kinase, usually tyrosine-specific (97), similar to the kinase often activated by binding of a number of growth factors to their receptors. Interestingly, phorbol tetradeconate acetate (TPA) (an important model compound in tumor promotion) (see below), may substitute for diacylglycerol and permanently ac-

tivate protein kinase C, a tyrosine-specific protein kinase (98), which phosphorylates insulin and somatomedin receptors (99). ras p21 protein seems to have a guanosine triphosphatase activity (100). Sometimes, only pieces of a cellular protein are coded, e.g., erbB codes for the integral membrane and large interior portions of the epidermal growth factor receptor (101) and fgr seems to produce a hybrid protein resulting from the recombination of actin and part of a tyrosine specific protein kinase (102). The onc gene of simian sarcoma virus was probably derived from genetic material encoding platelet-derived growth factor (PDGF), which is a growth factor in the type of tissue which becomes cancerous as a result of this virus (103), while c-myc seems to mediate the mitogenic response to plateletderived growth factor (104) by localizing in the nucleus, perhaps by its DNA-binding properties (105). This is especially interesting in that it has been shown that at least two oncogenes, ras and myc, can work together to transform primary fibroblasts in culture (106) (although Ha-ras-1 can be effective alone if it is mutated) (107). When an onc gene is expressed at high levels, an increase in gene dosage or expression has been correlated with the transforming activity of the virus (90). Alternatively, there is some evidence that the removal of a suppressor or regulatory gene, with the possible involvement of an oncogene as a second step, may be important in human retinoblastoma (108), while erbA (similar to carbonic anhydrase) blocks differentiation of immature erythrocytes and potentiates the effects of erbB (109). It appears that any step in the growth regulation of cells, from receptor through kinase and nuclear changes, can be affected by different oncogenes.

These data also suggest that there is a two-part model for transformation. The first part centers around the stimulation of some growth factor, and the second, an alteration in the cell nucleus. The first part may also involve the interaction of a cell with other cells, as growth factors can be elaborated (110), and the second may be an intrinsic change in the cell itself. Although this is a speculation and there are important gaps in information, a general picture of the mechanism of viral carcinogenesis is starting to emerge which also may be very important in understanding chemical carcinogenesis, since activated oncogenes are seen in chemically induced rodent tumors, e.g., ras in rat nitrosomethy-lurea-induced tumors (111).

Some normal cell *onc* genes can be activated when linked with expression control elements (ECE) derived from the retroviruses (72,112,113). The ECE regulate the level of expression or gene dosage of the retrovirus genome in an infected cell and can be used to "switch on" expression of other genes. Recent studies suggest that when the ECE is linked to an *onc* gene it activates the gene's transforming potential by increasing the gene's level of expression (112). In vivo, the same two factors, *onc* and ECE, which appear to link by chance, have been shown to be responsible for oncogenesis in animals infected with certain retroviruses (112).

The viruses from which the acute transforming re-

troviruses seem to have originated, the leukemia retroviruses, usually cause leukemias or lymphomas, but can also cause a variety of other neoplastic diseases in animals after long latent periods (86). When these viruses infect a large population of cells, viral genetic information can integrate at many loci in the host genome. The stable integrated form of the virus (called the provirus) is bracketed by the same ECE described above. These ECE are virus specific and are generated during the integration process (114). In some instances these viruses will integrate at loci in the host chromosome adjacent to cellular onc genes and will cause neoplastic disease (115). Similar to the situation described above for the acute transforming retroviruses, this specific integration results in elevated levels of expression of a specific cellular onc gene which becomes regulated by the viral ECE. The long latent period required for neoplastic disease induced by these viruses may be associated with the frequency of, or time to, specific integration in the host chromosome of the viral ECE adjacent to a cellular onc gene.

Some radiations and chemical carcinogens might enhance either the frequency of initial integration of the virus or subsequent rearrangement of viral material by inducing chromosomal translocations. In Burkitt's lymphoma, a characteristic portion of human chromosome 8 (carrying c-myc) is translocated to chromosome 14, the site of immunoglobulin heavy chains (114-116). This onc gene is the same one activated by a retrovirus transcription control element in avian bursal lymphoma disease (117,118). Also, c-abl is translocated in chronic myelocytic leukemia patients with a Philadelphia chromosome (a translocation of the gene from chromosome 9 to chromosome 22 adjacent to an immunoglobulin light chain cluster of genes) (116). Such translocations might conceivably result in the juxtaposition of an onc gene and an ECE, altering onc gene expression. The same result might arise from direct genetic insult (via some carcinogen) to the ECE which normally regulates expression of the cellular onc gene.

Considerable study is now being focused on ECE. It has been known for some time that certain nucleotide sequences which bracket structural genes are associated with the initiation and termination of gene expression (120). However, it was first shown with transforming genes that a region termed "enhancer" must be present in order for these start and stop signals to be recognized (72,112,121,122). Although it is not yet known how enhancers function, apparently they can be located in a number of different positions surrounding the structural gene. Moreover, even though the few that have been examined bear little sequence homology, the enhancer sequences from retroviruses have been shown to function to allow expression of a DNA tumor virus, SV40 (122). Also, the sequences seem to be utilized in normal differentiation, as shown by the evidence that immunoglobulin heavy-chain class switching in a pre-B cell line is accompanied by gene rearrangement (123), and, in the development of Burkitt's lymphoma, the activation of the translocated c-myc gene seems to occur

by an enhancer in the heavy-chain immunoglobulin locus (124). It is possible that translocation into a region controlled by an enhancer or mutations in gene enhancer or activator sequences of cellular *onc* genes could participate in expression of a transforming phenotype.

Another area of considerable study during the past two years has been the direct isolation of dominant transforming genes from human tumor cell lines (83-85). A biological assay referred to as transfection was used early in tumor virology to demonstrate that host genomic DNA containing dominant viral onc genes could be transferred to normal cells (86). The early studies were developed and refined by using cells transformed with acute transforming retroviruses; they demonstrated clearly that the integrated provirus could be transferred as part of the cellular genomic DNA in specific cell lines capable of assimilating foreign DNA into their chromosomes. By using this technology it has been possible to transfer dominant transforming genes from a variety of tumor cell lines from different species, including human. These include genes from human lung cancer, colon cancer, bladder cancer, and breast cancer cell lines (83-85,125-127). By utilizing recombinant DNA techniques, it has also been possible to identify and isolate (in certain instances) these dominant transforming genes from the transformed recipient cell and to show that these isolated genes are of human origin (83-85). In the molecular cloned state, the genes have high oncogenic potential. It is of particular importance that some of these genes (for example, genes associated with human bladder, lung, and colon cancers) are related to a specific family of retroviral onc genes (128-130). In the case of a human bladder cancer gene, a single base change in normal human gene results in expression of its oncogenic potential (131-133). Also, in the case of chemically induced tumors, there is suggestive evidence that the activation of an oncogene can occur by a single base change (111). The study of RNA tumor viruses has provided an initial understanding of how viral onc genes cause cellular transformation. The correlation with human onc genes provides an enormous leap in our understanding of causal factors in human neoplastic disease.

Recently, retroviruses of the kind that cause leukemia in animals have been isolated from humans. These human leukemia viruses (HTLV) are known to be associated with several types of T-cell leukemias in man (134,135). When they are transmitted to normal human T-cells, they induce a morphology, behavior, and phenotype similar to those naturally occurring in transformed cells isolated from individuals with T-cell leukemia and lymphomas (136). This virus, the presence of which correlates with clusters of the disease, thus may represent a true human cancer virus (137,138). Other viruses, such as herpes viruses, especially Epstein-Barr virus (EBV), hepatitis B virus, and papilloma viruses, have also been associated with human cancers (139). Although these viruses do not fit all the presently recognized criteria for a true human tumor virus, they may nonetheless influence the development of certain human cancers. Thus, EBV appears to be a stimulus for proliferation of B-cells which may be an early event in the development of African Burkitt lymphoma (140). Herpes simplex, a DNA virus, is associated with cervical cancer (141), and hepatitis B virus to liver cancer (142). Papilloma viruses cause benign skin growths (warts) and some strains have recently been shown to be associated with squamous cell carcinoma (143). Much current work is focused on a possible similar role for papilloma viruses in cancers of the genital tract and accessory organs (144,145). How, and even whether, these and other viruses play a role in various types of cancer or interact with various "physical" and chemical carcinogens is poorly understood; but the belief that DNA is a critical target for the biological carcinogens is strongly supported by current studies. New methodology to prepare antibody—using chemically synthesized peptides from nucleic acid sequences—makes it possible to generate antibodies against several different onc gene products (146) and to characterize the molecular properties of the transforming proteins in transformed cells. This method should aid in developing a better understanding of how chemicals and viruses interact with one another.

Radiation. Radiations, both ionizing (the corpuscular radiations and X- and gamma-ray photons) and nonionizing (ultraviolet light) radiations, have been shown to act as carcinogens (28). Ionizing radiations are ubiquitous, with cosmic rays and naturally occurring radioisotope decay being examples of major sources of background radiation. UV is also ubiquitous, being a component of sunlight.

Carcinogenesis by ionizing radiations is generally related to the spatial distributions within cells of the ionizations they produce (28) as well as total dose, dose rate and fractionation, and target tissue (147). The deposition of energy which results from the interaction of radiation with biological media creates ions, free radicals and excited molecules (147). The free radicals are thought to be most important product since: hydroxyl free radicals are involved in some of the biological effects of radiation (148); chemicals that are cellular freeradical scavengers inhibit the ability of ionizing radiations to transform cells in vitro (149); and chemicals involved in protection from free radicals (e.g., selenium, which is important in the action of glutathione peroxidase, an endogenous scavenger system) inhibit radiation-induced transformation (147). (These data also indicate a role for modulators of metabolism in radiationinduced carcinogenesis, since enzyme systems may be modified by metabolic changes.) Free radicals will react with many cellular macromolecules, but the production of DNA single-strand breaks, double-strand breaks, chromosomal aberrations, and the production of mutations by ionizing radiation, argue that the DNA is the critical cellular target for the biological effects (although the relationship of any of these phenomena with carcinogenesis is not definitive) (147). Although experimental dosimetry is much better defined for radiation compared to most chemical agents, the molecular lesion(s) significant for carcinogenesis is (are) unknown and may be different for different radiations [one example of a possibly significant lesion is the reaction of the hydroxyl radical with the deoxyribose in DNA, which results in DNA strand breakage (130), although many other lesions are formed]. The absence of "radiolabelled adducts," common for many chemical agents, has hampered analysis.

UV photocarcinogenesis is wavelength-dependent, with UVB (photons with a wavelength of 280-320 nm) being most effective on mouse skin (151). Although the photoproducts produced by UV irradiation in cells are myriad (152), reaction with the DNA has been shown to be the critical factor in carcinogenesis. There are a number of lines of evidence which suggest this. First, in patients with actinic keratoses (a premalignant condition) the DNA repair capacity for UV-induced damage (UV excision repair) in lymphocytes was found to be less than in matched controls (153). Also, individuals with xeroderma pigementosum, who exhibit a predisposition for sunlight-related cutaneous malignancies (although having a number of genetic complementation groups) generally are deficient in the ability to repair damage induced by UV (67). By use of antibodies, DNA has been shown to be altered immediately and directly by UV irradiation (154). The persistence of UV photoproducts after photoreactivation, a repair mechanism which specifically corrects pyrimidine dimers in the DNA induced by UV, is directly correlated with the capacity of UV-irradiated tissue to develop into tumors (70). Finally, the action spectrum (relationship of intensity of a biological response to frequency) of UVB photons is the same for their carcinogenic action and the production of pyrimidine dimers in mouse skin (155).

It is important when making estimations of risk for chemical agents that we realize that many of the tenets upon which these estimations are made are based upon information from radiation carcinogenesis. For example, the hypothesized mechanisms of the action of radiation, while not the primary concern of this document, have served, and continue to serve, as the basis for some of the primary models for chemical carcinogenesis (156,157). Much evidence that damage to the cellular DNA is a step in the induction of tumors comes from studies using radiation. Similarly, various models for high-dose to low-dose extrapolation have their origins in studies using radiation and the best data for these models have been obtained from such studies.

Chemical Carcinogens. The interaction of chemical electrophiles with DNA, as discussed in the section on metabolism, may lead to covalently bound adducts, intercalations, strand breaks, phosphotriesters, crosslinks, apurinic sites, apyrimidinic sites, deaminations, and/or hydrations, all of which are forms of DNA damage that may alter genetic information (54,158–170). Studies of chemical carcinogenesis have focussed on covalent adducts. Examples of this are the observations that: simple alkylating agents can methylate or ethylate any of the nitrogens or oxygens in either of the four bases in the DNA as well as the sugar residues and

phosphate backbone of the DNA (171); the epoxide derivatives of aflatoxin can attack the N-7 position of guanine (172) (as well as other sites); activated aromatic amines attack the C-8 and N-2 position of guanine (173,174); and the dihydrodiol-epoxide derivatives of benzo(a)pyrene and certain other polynuclear aromatic hydrocarbons attack the N-2 position of guanine and, to a lesser extent, form adducts with adenine and cytosine (175,176). Activated derivatives of these and other carcinogens form similar lesions in the DNA of a number of diverse tissues and species (176,177).

The biological significance of these alterations in the DNA is not always clear, and different alterations may have very different biological effects. Some alterations may cause mutations (178,179). For example, modification of the O-6 position of guanine will cause this base to be misread, during DNA replication, as adenine (180). Also, the major guanine adduct of benzo(a)pyrene, through a transversion, may be misread as a cytosine or thymine in replication (181,182). Alterations in the control of DNA are possible. The amount of methylation of DNA, which may be used in the control of genetic expression (183), can also be altered by some agents. Some of the carcinogenic metal complexes may work through alteration of the enzyme that replicates DNA (DNA polymerase) to cause alterations in the genome through miscoding of DNA bases after replication (this will be discussed in detail in the section on fidelity/infidelity of replication).

"Physical" Carcinogens. This class of carcinogens encompasses a number of very diverse phenomena including skin abrasion, scar formation, implantation of nonreactive materials (foreign bodies), schistosomiasis, etc. (the term physical carcinogen also is sometimes used to refer to radiation, i.e., a physical agent). Common factors of this class seem to be the need for the development of a chronic fibrosis, or a least an inflammatory response, and the observation that chemical properties, e.g., of a foreign body, are not important (29). An interesting mechanism, with a number of steps, proposed for foreign body carcinogenesis may be relevant to the whole class. During tumor induction, there is an acute inflammatory response with an outgrowth of capillaries as cells adhere to the surface of the foreign body. This response is induced by a number of substances (including mitotic stimuli similar in some ways to growth factors), elaborated by cells in the vicinity of the foreign body. Fibroblasts increase in number, which leads to tissue deposition of collagen (fibrosis). After a latent period, uncontrolled proliferation begins (184). The initiation event seems to occur away from the foreign body surface, inducing an initiation cell through changing the environment of the cells in the vicinity of the foreign body (29). However, the inflammatory response results in an active cellular proliferation of a number of vascular-derived cells (this step can also occur in the inflammatory response generated by the other "physical" carcinogens). By its presence, the foreign body causes the macrophages to become quiescent and the reactive tissue to develop into a chronic fibrotic state, so that there is surface-independent maturation of the initiated cell (29). Asbestos, which is frequently considered a member of this class, may induce cancer by a different pathway since, unlike other "foreign bodies," fragments of asbestos fiber are incorporated into and activate macrophages, as well as slowly dissolve components (185). However, the end result is a fibrosis, i.e., asbestosis, which, though more diffuse, is similar in many ways to fibrosis induced by the foreign bodies (29). The size of the asbestos fiber and the possibility of chemical effects because of fiber dissolution of adsorbed molecules make direct comparison with foreign body cancers difficult.

It is not clear how, if at all, this class of carcinogens interacts with the genome. Speculations include: no interaction with the genome, with the recruitment of omnipresent previously initiated cells by a proliferative stimulus into neoplasia; the environment of the inflammatory response, especially the production of free radicals by macrophages, causes chromosomal damage and rearrangement resulting in transformation; and a stimulation of proliferation may result in mutation as a result of infidelity of replication (see below); however there is little information in the area.

General Statements. The interaction of electrophiles, or the products of irradiation, with DNA can play a role in the induction of a tumorigenic phenotype through either of two mechanisms: directly, by altering genetic material through a somatic mutation (178) or indirectly, by altering gene expression (183). Somatic mutation, for example, can arise from the direct interaction of DNA with electrophiles, free radicals derived from carcinogens, or with activated oxygen produced by metabolism of the agent. The alteration of gene expression can arise from (1) a direct mutational event in a regulatory gene controlling expression (e.g., base transition) (178,186); (2) changes in gene expression resulting from an increased rate of abnormal base incorporation due to enhanced DNA synthesis, or abnormal methylation of nucleic acid bases (183); or (3) induction of genetic transpositions or gaps in the DNA leading to an alteration in gene expression (higher order alterations) (190). Direct alterations in the informational content of cells seem to be common for many carcinogens. The potential involvement of an onc gene with either direct or indirect alteration of the genome has been discussed in a previous section.

Understanding the significance of these changes may now be possible with the recent development of new methods to determine the structural alterations caused in DNA by chemical carcinogens and radiation (188). This has occurred as a result of recent advances in spectroscopic and spectrometric instrumentation, especially in the areas of nuclear magnetic resonance spectroscopy and mass spectrometry. Significant progress has also been made in understanding the effect of carcinogens upon DNA's higher-order structure (189–191). For example, recent studies have demonstrated that while methylating and ethylating agents do not induce major steric changes in DNA, aromatic amines induce "base

displacement" (192). Additionally, the carcinogen 2-acetylaminofluorene appears to induce a flip from a conventional right-handed DNA helix to the left-handed Z form (175,192), which may induce changes in gene regulation (191). Such changes, while not induced by all chemical carcinogens could, if they occur in vivo, suggest mechanisms by which chemicals induce major steric alterations in the structure of DNA. The biological consequences of such changes, however, are uncertain. Even newer techniques are being developed, such as radioimmunoassays using monoclonal antibodies against specific forms of DNA damage (193) and reversed labeling methods that are able to detect a single damaged base per cell (194), that may eventually allow routine estimation of DNA damage in humans and better estimation of its biological effects.

Modulation of Altered Genetic Information

For many carcinogens, it is necessary for cellular proliferation to occur before the damage induced by a given agent is expressed. For instance, a cell replication must occur within 48 hr to express X-ray-induced transformation in cell culture (195). Replication must also occur in vivo in liver (196). One way to think about the process is that a cell is first "primed" for alteration by interaction with a carcinogen; cell replication then results in at least one daughter-initiated cell, i.e., a cell irreversibly altered so that it can interact with the proper stimulus to form a malignant cell.

The alteration of genetic information that produces the initiated cell is modulated by a number of factors. Two of the most important are: the amount of DNA damage (adducts, strand-breaks, etc.) that is removed from the "primed" cell and its daughters (DNA repair) and the accuracy of DNA synthesis (fidelity/infidelity of replication), both as a result of damage to the genome and as a result of changes in cellular parameters (ion concentrations, etc.).

DNA REPAIR. DNA damage is believed to be one of the primary factors in chemical carcinogenesis. The bulk of evidence for this assertion comes from studies implicating UV and ionizing radiation as etiologic agents in cancer induction (198). For example, as discussed above, individuals with xeroderma pigmentosum who are defective in repair of UV-induced and some forms of chemically induced DNA damage (68) exhibit a high incidence of light-related cancers (199). More generally, the available data are equivocal as to whether there is an increased risk of development of cancer of the internal organs (69,200). Problems with interpretation of these observations include: chemically-induced and UVinduced damage may induce cancer by different means; organ-specific differences exist for cancer induction; and the time to tumor may differ significantly between organs and death of the individual due to complications arising from the skin tumor prior to the onset of cancer in an internal organ or tissue. Also, however, the databases may have been too small to support any conclusion and a larger study may resolve any problems.

Because of the importance of DNA damage, a number of short term bioassay systems that measure it, directly

or indirectly, have been developed to screen for potential carcinogens. One of the most important modulators of the amount of DNA damage an agent causes, DNA repair, has been studied in numerous *in vitro* and *in vivo* biological systems (201).

From these studies, various pathways for repair of numerous forms of DNA damage have been identified (Table 3) (202-251). Many of the processes of repair in mammalian cells are similar to those described in bacteria, and thus it is assumed that these systems have been conserved during the course of evolution. It has been speculated that the primary role of these systems in evolution was to protect the genetic information of the species from both endogenous and naturally occurring exogenous DNA-damaging agents, especially long enough for the organism to replicate (249). All species examined thus far possess at least one form of DNA repair (219).

Since the nature of DNA lesions formed by agents which are genetically toxic and the methods of their repair are so similar across species, the use of prokaryotes to screen for this particular form of genetic toxicity seems reasonable. However, the greater complexity of the mammalian systems requires that care must be exercised in the interpretation of bacterial data. For example, chromatin structure in mammalian cells appears to make some regions of the DNA more accessible, and others less accessible, to adduct formation and repair (250-252). Furthermore, significant differences in the repair efficiencies between tissues (253) and species (254–264) have been observed. In terms of general understanding of the significance of these differences, it is known that among the placental mammals there is a good correlation between the maximum achievable lifespan of a species and the ability of primary fibroblast cell cultures from these species to perform unscheduled DNA synthesis following UV exposure (262,264). It is not known whether there is any such relationship for DNA damage induced by chemical electrophiles. Also, in animals, the organization of cells into tissues presents other complicating factors. One example, based on tissue interactions, involves the complex effects of xenobiotic liver metabolism on the activated agent concentrations in other tissues. Another involves tissues that are less active mitotically than others. The longer cells are given to perform DNA repair prior to DNA replication, the less likely it is that a transformation event will occur (199,261). Still another example may be the maintenance of many cells in stationary phase (G_0) in a tissue. Those cells could be triggered into DNA replication, and run the risk of mutagenesis, by an agent delivered in a dose sufficiently high to induce cell killing (see below, section on hyperplasia) (267). Since the latter possibility has been used to explain the apparent threshold effects seen in various longterm low-dose vs. high-dose carcinogenesis experiments using nitrosamines (by presuming that the high dose induces cell killing and resultant division leads to effects that are only seen with a high dose) (265), understanding factors such as these are important in assessing risk.

Table 3. Mammalian DNA repair mechanisms.

Reaction	Action	References
Nucleotide excision repair	Removes bulky, noncoding lesions from the DNA in a manner similar to but not identical with bacterial nucleotide excision repair	(167, 201–207)
Base excision repair	Permits reinsertion of the proper base into the gap left in the DNA by the action of enzymes that effect the excision of inappropriate bases from DNA as the free base (DNA glycosylases), avoiding scission of the DNA backbone, as well as a system similar to the bacterial one requiring strand scission. Included are direct demethylation by transfer	(208,209,212-217) (209-211, 218-221)
	of a methyl group to an acceptor protein and AP site repair (direct repair of a removed base)	
Strand break repair	Rejoins single- and double-strand breaks with the addition of few or no additional nucleotides through the action of a sealing enzyme	(222–228)
Photoreactivation	Light-activated mechanism specific for the breakage of the UV-induced covalent bond attaching two pyrimidines in a cyclobutane-type ring	(228–234)
Recombination repair (post-replication repair)	System once believed to occur in mammalian systems but now controversial, by which bulky, noncoding lesions are transferred to DNA synthesized after damage	(235–238)
Replication bypass (post-replication repair)	Process that may function in mammalian cells as an alternative to recombination repair in which the bulky lesions are bypassed and the gap created filled	(239, 240)
Inducible DNA repair systems	Still speculative (for mammalian systems) repair system in which DNA damage triggers the induction of enzyme systems to remove the damage	(241–247)

Other complicating whole-animal effects include diet, age, and hormonal status, all of which impact on DNA repair (266-268).

Many biochemical mechanisms are involved in the repair of DNA damage, and many factors may modify their effectiveness. One mechanism is a nuclear enzyme system which polymerizes activated ADP-ribose residues to form poly(ADP-ribose) (269). The biological function of ADP-ribosylation is not clear; however there is a relationship between it, DNA damage, and DNA repair, which may involve the capacity of the system, demonstrated in vitro, to utilize damaged DNA (270). This system also seems to influence the production of sister chromatid exchanges (271) and has some relationship to mutagenesis and carcinogenesis, although not a simple one (270). Another mechanism, recently brought to light by the identification of a series of enzymes [collectively referred to as the DNA glycosylases (214-218,272-278)] active in base excision repair (Table 3), has indicated that there is much greater specificity for the various kinds of DNA damage than was previously thought. Over a dozen of these enzymes have been isolated from bacterial and mammalian systems, each involved in the repair of a specific form of DNA damage. The apparent lack of nonspecific DNA damage recognition enzymes in adult mammalian tissues and the high degree of specificity of those repair systems that have been studied suggest that there is specific repair of many forms of DNA damage, including those induced by man-made substances. Another recently discovered mechanism involves an inducible antimutagenesis repair system in bacteria (279). An inducible system, considered part of post-replication repair, was also found in mammalian cells (243,244). Low doses of certain mutagenic agents appear to induce a response in the liver that enhances the repair of damage subsequently induced by those agents. For example, in rat liver, an inducible protein transfers the intact methyl group, added to the guanine base by the potent carcinogen dimethylnitrosamine, from the DNA to a cysteine residue. Prestimulation of the animal with small doses of the compound can enhance this inducible repair system by as much as threefold (221).

As an example of the relation between knowledge of mechanisms and resolution of science policy questions, one can consider the relationship of DNA repair systems and a threshold phenomena in carcinogenesis. In simple terms, the threshold concept maintains that there is a level of exposure to an agent below which there is either no observable effect and/or no adverse biological effect. usually as a result of some reserve (280). The practical manifestation of this concept is a dose-response curve which does not appreciably differ from zero response at some nonzero dose. The concept is used to evaluate many toxic phenomena, but has been a source of controversy in the evaluation of carcinogenicity. The existence of DNA repair systems has been used by some authors to support speculation that a threshold exists for the biological effects of chemical carcinogens, by suggesting that low levels of DNA damage could be totally removed, thereby removing the effects of a low dose of carcinogenic agent. Such an extrapolation is not supported by present data, as is shown below.

The following analysis will concentrate on the question of a threshold for DNA damage and it must clearly be recognized that DNA damage is only one part of a complex chain of events leading to carcinogenesis which may be effected in multiple ways by an agent. A threshold, or lack of threshold, for DNA damage does not necessarily determine a threshold, or lack of threshold.

for carcinogenesis.

First, no DNA repair system reduces DNA damage to zero. Consistent with this observation is the observation that DNA damage accumulates in postmitotic tissue, as a function of age even in the absence of known chemical exposure (282). The repair proficient systems found in dividing Escherichia coli, when fully induced, can reduce mutagenesis by up to 5000-fold, but the mutation rate in exposed cultures is still 10 times that which is produced spontaneously (283). The fragmentary data from mammalian cells suggests that there may be an inducible system (282), but it would be unreasonable to assume that replicating eukaryotic cells would be at less risk than prokaryotic cells from low doses of a mutagen. Second, not all DNA repair systems are error-free. While the data on inducible error-prone bypass mechanisms in mammalian systems are meager (282), virus reactivation, which is likened to a mutagenic effect, has been well established in eukaryotes (245,282). This suggests that mammalian cells also possess an error-prone bypass system. Third, major variations in DNA repair efficiency occur between cells, tissues, strains and species exposed to similar doses of the same carcinogen (254-268). This heterogeneity limits generalizations about the effectiveness of repair systems, constitutive or inducible, relative to the completeness of repair or the notion of a "safe" threshold. Tissue differences in repair might result in a dose-response relationship indicating a threshold for repair in one tissue, but a relationship indicating no threshold for repair in another, thereby providing little hope for uniform protection within an organism. In tissue or cell culture, repair appears to vary between cells. Thus, even if biochemical analyses, which normally measure the average repair capacity of a heterogenous population of cells, demonstrate that a tissue or cell culture is repair proficient, the possibility of repair deficient subpopulations cannot be disregarded.

It is clear that DNA repair will not produce a threshold for DNA damage. However, it is not clear what the significance is of the DNA damage which results from exposure to exogenous agents. Since some damage results from normal metabolism and, probably, exposure to chemicals in foodstuffs (33) over the life of the animal, the importance of the additional burden induced by a carcinogen above this background is hard to determine. As noted above, since DNA damage is only a part of the complex process leading to the production of cancer, this determination probably can only be made when a clear understanding of the mechanism(s) of carcinogenesis is achieved.

FIDELITY/INFIDELITY OF DNA REPLICATION. Another modulator of the alteration of genetic information is the fidelity of DNA replication. Although it is not proven that the role of DNA replication in initiation is to stabilize incorporation of the effects of DNA damage in the genome ("fixation") (283), it remains the most attractive possibility. The fidelity of DNA replication (284) will help determine the integrity of genetic infor-

mation in the initiated cell.

Before one considers the ways fidelity can be altered, it is important to understand the consequences of error accumulation in DNA in nondividing somatic cells (249). In dividing cells, one is concerned with replicative error propagation; that is, the transmission of mistakes from one generation to the next. Mechanisms that enhance the frequency of such mistakes could involve alterations in the DNA synthetic apparatus and have been referred to as intrinsic mutagenesis (285). In nondividing cells, one is concerned about the effect of accumulated unrepaired DNA damage on cell function. When considering mechanisms in carcinogenesis, attention is normally limited to either dividing cells or to resting cells destined to undergo at least one further division cycle. With our present understanding, a cell which fails to undergo further division is not usually considered pivotal with respect to neoplasia. However if amitotic cells become, through some mechanism, able to divide, or if nondividing cells are involved in the control of dividing cell populations, the accumulation of damage in them can be significant. For instance, nondividing neurons control the level of thyroxine, and it has been shown that this hormone is an important factor in X-ray transformation of cells (286). Therefore, although usually not appreciated, the effects of agents on nondividing cells should not be ignored.

In dividing cells, a repaired or inadequately repaired DNA lesion, such as an adduct, when present on the DNA template at the time of replication, could result in a genetic alteration, e.g., base substitution, such as replacing a guanine with a cytosine. However, damaged DNA need not always cause significant base substitutions during DNA replication. First, parental cells containing the damaged DNA may fail to divide and thus be progressively diluted during cellular proliferation. Second, due to the redundancy of the genetic code, as well as other factors, approximately 25% of incorrect base substitutions may not result in amino acid substitutions. On the other hand, a number of modifications of the DNA template or of the DNA polymerases have been shown to cause incorrect substitutions during copying by DNA polymerases in vitro (287,288). If these modifications are not corrected by subsequent DNA repair, it is a reasonable expectation that the unrepaired lesions will in time result in either mutations or cessation of cell reproduction (263). Alkylating agents and carcinogens have been shown to induce misincorporation in vitro by modification of DNA templates and by removal of purines from the DNA (289-294). Also, the induction of an error-prone repair pathway (SOS) may cause problems since the underlying concept of this repair pathway is that it can alleviate potentially lethal damage only at the expense of inducing mutations (279).

Even if the template is undamaged, other factors which can be altered by carcinogens, both directly and/or indirectly, may cause the template to be copied incorrectly. Some factors that have been found to diminish the fidelity of DNA synthesis *in vitro* in two different systems (289) and may have to be considered *in vivo* are: (1) ratio of incorrect to correct nucleotide substrates

(294-300); (2) type of incorrect nucleotide substrates (296-300); (3) different metal activators for DNA polymerase (288,291,295); (4) nonactivating metal mutagens and/or carcinogens (288); (5) depurination of template (293).

During replication, the fidelity of DNA synthesis is altered by changes in the reaction conditions. The error rate of mammalian DNA polymerases in vitro has been shown to be dependent on the ratio of correct to incorrect nucleotides with both polynucleotide and natural DNA templates (295,296). There is evidence that alterations in cellular nucleotide pool sizes and content are mutagenic (297-299), and the cellular homeostatic mechanisms for precise maintenance of deoxyribonucleotide concentrations may be altered by various chemicals. Furthermore, agents may directly alter some of the nucleotides in the pools available for replication, leading to possible misincorporation (300,301).

Lindahl and Nyberg (302) estimated that the in vivo rate constant for thermal depurination is approximately $3 \times 10^{-11} \text{ sec}^{-1}$, suggesting that as many as 10,000 purines may be lost from the genome of a mammalian cell per generation time (20 hr). Furthermore, this rate constant may be significantly increased after alteration of bases by alkylating agents (302). The intensity of this formation of damage suggests that during replication, the cellular mechanisms for correcting depurinated sites on DNA might not be able to repair every site before the polymerase complex involved in replication encounters a damage site. Thus, replication over a damaged site may be a secondary result of cellular interaction with a carcinogen. Lastly, mutagenic/carcinogenic metals can affect the fidelity of DNA synthesis in vitro (289). Thus, various agents that do not damage DNA directly or enhance the production of metabolites that can induce such damage may exert their effect by altering the fidelity of DNA replication. Methods to assess or screen for such agents in mammalian cells are generally lacking. The end result of these processes is that a daughter cell can have an irreversible alteration in genetic information (283), even though the cell which interacts directly with the agent does not. This altered cell may be similar to normal cells in many respects, but may be capable of giving rise to a cancer, i.e., be initiated, and may lead to carcinogenesis if, during the lifespan of the animal, it encounters an appropriate stimulus.

As previously noted, it is not clear what role DNA synthesis has in initiation, though it has been shown that manipulation of the rate of cellular proliferation can modify tumor incidence. This has been demonstrated: (1) in partial hepatectomy for animals treated with methylnitrosourea, dimethylnitrosamine (283), benzo(a)pyrene and radioactive particles that localize in the liver (303); (2) with chemical treatment combined with diet changes, drugs or natural physiological agents such as hormones or bile acids which alter proliferation rates (283), e.g., retinoids, which stabilize the differentiated nondividing state of epithelial cells among their other biological effects, have been used to inhibit mammary and colon carcinogenesis (304,305); (3) in stimu-

lated hyperplasia resulting from a toxic effect (306); and (4) regeneration in skin using skin abrasion (307) (much of these data are hard to evaluate however because the treatments induce a chronic inflammatory response and it is not clear whether one is studying the effects of proliferation or the inflammatory response).

Cytotoxicity could be a stimulus for cellular proliferation which may result in hyperplasia (308) (usually termed restorative or reactive hyperplasia). A chemical which may work through such a mechanism is carbon tetrachloride, which is carcinogenic in mice but which has little or no activity in tests which measure alteration of the genome (the compound was positive in yeast, but only at doses which were almost lethal, casting doubt on the relevance of the result) (309). Replication may act to "fix" changes in cells previously "primed" by some unknown stimulus or it may result in replication errors through some of the mechanisms suggested above. The possibility that hyperplasia can lead to carcinogenesis has led to the speculation that a cytotoxic mechanism is of primary importance in the carcinogenic effect of all carcinogens which do not appear to act directly on the DNA that are tested at a high dose (310), with a suggestion that a dose which does not lead to cytotoxicity would have no toxic effect. This hypothesis overlooks the obvious problem that cytotoxicity is by no means the only way an agent which does not appear to act directly on DNA can alter either replication or gene expression. Certain compounds can stimulate hyperplasia without significant cytotoxicity (311). Furthermore, it has been shown many times that neither the induced hyperplasia (312) nor the irritant properties resulting from promoters, such as PMA (313), are sufficient to explain the promotion effect. Thus, a cytotoxic action should be considered simply as an additional possible mechanism of carcinogenesis and not equated to an explanation for carcinogenesis for all nondirectly acting agents tested at high dose.

Whatever mechanism(s) is (are) important in the formation of initiated cells, it is clear that cells irreversibly altered in a heritable fashion are induced during this process and that these are the progenitors of further new cell populations during the carcinogenic process. The properties of these cell populations are discussed in the next section.

Cancer Formation

How initiated cells with altered genetic information result in a cancer seems to be the least understood area of carcinogenesis. The process can be divided into two operational stages, promotion and progression. Promotion will be used to describe the process(es), usually initially reversible in test systems, but later irreversible (314), that results in cellular proliferation and terminates when one or more neoplastic cells, i.e., cells with the ability to grow autonomously, is (are) formed. Neoplastic progression will refer to the next step(s) in which cells become able to form tumors and become malignant with some potential to metastasize.

PROMOTION. A promoter is experimentally defined as an agent which results in an increase in cancer induction

after long-term application subsequent to treatment of an animal with an initiator. Under otherwise identical conditions, neither initiator nor promoter alone induces many tumors, nor does a regimen with reversed order (treatment with promoter followed by an initiator) (312). At a particular dose level and treatment schedule, a carcinogen that does not require supplemental promoter activity is termed complete. Many or all complete carcinogens act as both an initiator and promoter in their interaction with the organism, and some act as initiator or promoter for different tissues in the same organism at the same time (315,316). Also, the promoter action of a complete carcinogen may not act by the same mechanism as promoters used in initiator-promoter paradigms (313). Separation of these two aspects of carcinogenic action is frequently difficult. However the term remains a convenient description for one part of an experimentally isolated complex process. For the most part, the phenomenon of tumor promotion has been most carefully delineated in skin and liver carcinogenesis. It remains to be established more firmly with cancer at other sites, though there is suggestive work in colon, breast, kidney, central nervous system (283), and bladder (311,317) carcinogenesis. At present, given the lack of information on mechanisms of carcinogenesis in humans and the myriad interactions of tissue and the biological effects of promoters demonstrated in experimental systems, it is premature to designate agents as acting through only a promotion-type mechanism, although a number of agents seem to have some promoter action.

This experimental definition will not clearly differentiate between promotion and progression, since it measures only the production of a malignant tumor. In practice, however, laboratory studies of promoter action are usually ended after a focal mass of cells (e.g., papilloma, nodule) is observed (311). The appearance of the first neoplastic cells has been proposed as a definition of the end of promotion (283). This definition is useful, since it emphasizes the frequently seen reversible effects of promotion, i.e., regression of the direct results of a promoter (e.g., papilloma) with termination of its use, while clearly indicating that the role a promoter has in carcinogenesis is to induce a presumably irreversible event, a neoplastic cell. A similar definition is suggested by the separation of promotion into two stages, based on work in skin promotion in mouse (314). Stage one is partially reversible, while stage two, which requires cell proliferation, is initially reversible and then becomes irreversible. When induced, many of the focal masses are reversible, especially in skin and liver (283). How these nonneoplastic masses originate is not completely known, but they seem to arise through the differential stimulation of cells altered by the initiator to respond to the promoter (283). Of the persistent masses, in all systems in which they are induced, almost all do not become neoplastic (196).

Promoters are usually thought of as exogenous compounds. Many of them are plant products (311,318) such as certain of the phorbol esters, e.g., 12-O-tetradeca-

noylphorbol-13-acetate (TPA) or phorbol tetradeconate acetate, which is also known as phorbol myristate acetate (PMA). However, there is also evidence that certain compounds normally present in the body (endogenous) may also be promoters (319,320). Promotion by such endogenous compounds may be difficult to demonstrate, since production of the compound in the body must usually be altered (which normally has many physiological ramifications) to observe an effect, as opposed to exogenous compounds, whose levels are controlled by direct application to a tissue. Another type of interaction that seems to have promoter action occurs in foreign-body tumorigenesis (197). Although it has been claimed that a single event determines the sarcoma which results from the implantation (183), it is clear that there is a step where preneoplastic cells of probable vascular origin (produced in a surface-contact independent step) require a surface-contact dependent step to become neoplastic (183). Like other promoters, removal of a foreign body before the last step will, as a rule, prevent the sarcoma from developing. Thus some surfaces, as well as exogenous and endogenous compounds, seem to be able to act as promoters.

Despite recent significant progress, the mechanism of action of PMA, the model promoter, is not completely clear. There is good evidence PMA binds to a receptor (321,322), that activates protein kinase C (101,323) or that may activate the kinase permanently through similarity to diacylglycerol (98), similar to a number of growth factors. Other promoters also interact with a cell receptor (311,324), e.g., estrogens, suggested to be promoters of mammary and liver tumorigenesis (325), are known to produce their main biological actions through receptors (326).

PMA produces profound biological effects in vivo, including edema, erythema, inflammation, and cellular proliferation (a property which allows the promoter to participate in "fixation" if given within a short enough time after the initiator dose). But these same effects can be produced by agents that are not promoters (311). By stimulating specific separate effects of PMA in other ways, one can define, as noted above, at least two stages in promotion (314,327,328), supporting the hypotheses that cancer is a multistep process and that more than one aspect of PMA may be important in promotion. In vitro, promoters have been shown to induce an array of diverse morphological and biochemical changes (311,325,329). An important one may be the interaction of PMA and other promoters with cells to generate free radicals (330-332). A similar stimulation is seen with the use of either ionizing radiation or activated oxygen (332), agents which damage the DNA. The capacity to stimulate radicals seems to be correlated with promoter potency to generate tumors (330) and varies in a dosedependent manner for the strong promoter PMA (332). PMA can also cause dedifferentiation or inhibit terminal differentiation (334), stimulate cellular differentiation (335), alter macrophages (336), induce chromosomal alterations (337) and sister chromatid exchanges (SCE) (338), and induce virus synthesis (339). It is not known

which, if any, of these effects are central to the role of promoters in carcinogenesis, since evidence for all models of promotion is indirect. However, the effects on chromosomes may be especially relevant.

PMA can induce chromosomal anomalies such as SCE, although the concentrations (100 ng/mL) required for this effect are quite high (338). The activated oxygen species produced by the promoters, perhaps through an interaction with membrane components, can induce chromosomal damage and a soluble substance which induces chromosomal abnormalities (a clastogen) in nearby cells (337). However, the promotional effects of PMA are reversible unless it is given for a fairly long time while a chromosomal change, such as an aberration, is almost certainly not reversible. This suggests that a chromosomal alteration could be an event, induced by prolonged exposure to the promoter, which terminates the reversible period in promotion (191,312). This suggestion is consistent with the observation that papillomas, which emerge in a common response to promoter action in skin carcinogenesis during the stage when it is reversible, are primarily associated with a diploid karyotype, whereas malignant tumors usually have karyotypic alterations (340,341). The selection process for cellular characteristics important to neoplasia may occur during the entire promotion process [indeed, selection may be important even during initiation (283)]. Also, the need for prolonged exposure, leading to a relatively large cumulated dose, may be why such high doses have to be given to observe the effect in shortterm experiments in vitro.

Chromosomal changes, although not proven to be responsible for growth independence of neoplastic cells, are implicated in transformation (342). For example, SK-L7 human lymphoid cells markedly increase their cloning efficiency in soft agar and decrease their doubling time from 30 hr to 14 hr upon introduction of an additional chromosome 15 (343-345). More directly, a clastogenic effect is produced by cells from an individual with Bloom's syndrome, a clinical entity in which there is usually a high cancer incidence (337). Specific chromosomal transpositions, as seen in the case of Burkitt's lymphoma, can be associated with changes in the regulation of gene expression and also could be associated with phenotypic changes such as differences in antigenic expression and unresponsiveness to regulators of cell growth. (It must be remembered, however, that this analysis is drawing general conclusions about malignancy from the pathogenesis of rare diseases in persons with a limited life span, which may not be appropriate.)

The hypothesis that a chromosomal change is a causal factor in cancer induced by promoters is attractive, especially when viewed with the evidence concerning the effects of transposition in oncogenes; however, few data directly support this contention (199). Although there is a lack of methods to measure such changes readily in vertebrates, this hypothesis is interesting since such genetic alterations are much more drastic than localized changes in base sequences, may be either site-specific or non-site-specific, do occur at different frequencies,

appear to affect the stability of neighboring genes, and are virtually irreversible. In rapidly multiplying bacterial cells, conventional mutagens do not enhance the frequency of transposition (346); however, factors that trigger transpositions or govern their rate of movement are not yet understood well enough to determine whether mutagens enhance transposition in mammalian cells in vivo. Large-scale changes, such as rearrangements and deletions, rather than the small changes repairable by excision repair that can be detected in the usual tests for mutagenicity may be more important to predicting human cancer (199). It appears that localized lesions in DNA can be carcinogenic (55,347); it is not clear whether such lesions might act either independently or through induction of direct or indirect induction of major genetic alterations or through a combination of both factors (348).

Evidence is starting to accumulate that both oncogene activation and treatment by some promoters have some aspects in common, i.e., the activation of some growth factor-like stimulus. Interesting in this regard is the discovery that metallothionein gene induction by heavy metal ions (such as cadmium) and glucocorticoid receptors both work by activation of "enhancer-like" elements (349), which leads to the speculation that the direct interaction of certain metal complexes with DNA may have effects similar to hormone or growth-factor induction for gene expression. In some cases, the primary stimulus seems to need some second factor, another oncogene involving a nuclear protein in the case of oncogenes, or Phase II promoters, involving replication, to cause transformation. In this light, chromosomal changes might be seen as merely a possible, but not obligatory, method to accomplish the same end, with the final, chromosomal change equivalent to the second oncogene activation or the effect of proliferation and other changes in the case of the PMA stimulation.

Cancers in humans from causes such as exposure to cigarette smoke or exposure to benzidine in the workplace have a very long latent period. It is not clear whether this long period is the time from exposure to the production of a neoplastic cell (promotion) or the time necessary to go from a neoplastic cell to a malignant tumor (progression). Evidence that the carcinogenic risk of cigarette smoking declines with the length of time after smoking stops (350), suggests that most of the latent period is partially "reversible" and therefore probably occupied by a phenomenon similar to promotion.

This is also suggested by a recent model for the development of Burkitt's lymphoma, which seems to be a virally produced cancer (351). According to this model, a lymphoma clone develops in one or more steps from B lymphocytes that have acquired neoplastic characteristics due to EBV transformation, and the fully malignant phenotype occurs only when the lymphoblastoid cells acquire a specific chromosomal translocation that involves the immunoglobin locus. The long latency period seen in the disease has been suggested to arise from the need for a promoter-like stimulus that may

cause proliferation and interfere with the differentiation of the long-lived initiated B cells (341).

Since neither initiator nor promoter alone is sufficient to cause tumors and since promotion appears to be involved in the carcinogenic process, especially in time to tumor induction, low-level exposure to certain carcinogens may only become significant after subsequent exposure to promoters. This bears on the question of a threshold for tumor promoters. Although it has been stated that promoters have a threshold (310), there is little good evidence for this. Studies of the dose-response curves for promoters, e.g., PMA (352), have clearly been inadequate for the purpose of defining a threshold, as is usually acknowledged by the authors involved. Also, there is a question of interpretation. For instance, recent work has shown that bladder stone formation by high doses of melamine is correlated to the carcinogenic effect of this agent in rats (353). (The significance of this observation for carcinogenesis has been brought into question by the discovery that mice, with similar propensity to develop stones and induce tumorigenesis by bladder implantation, have little melamineinduced bladder cancer.) This has been taken by some authors as indicating a threshold for "promotion." Alternatively, the dose used in the assay can be considered to have induced a significant pathological effect (other than tumorigenesis) that was important in generating the cancer seen. Similar considerations are appropriate for agents which induce other functional pathological effects only in high dose, e.g., immunosuppression. It must be remembered, though, that agents which induce these pathological effects may have other actions [e.g., azathioprine is mutagenic as well as immunosuppressive (309)], which could contribute to carcinogenesis at lower doses. Also, it must be kept in mind that the tendency to induce a pathological effect may vary with the well documented variations in individual human response to agents. Important factors in tumor promotion such as dose, dosing schedule, tissue specificity, interaction with initiating stimulus, etc. create substantial uncertainty about any general statements regarding thresholds.

Generally, animal bioassay studies, because of their design and the conditions under which they are conducted, fail to distinguish between initiating or promoting activity or between a direct or indirect action by the agent on the genome. Although it may be useful to group compounds with some criteria; e.g., mutagenic, or having a particular biological effect in a particular strain, one should be careful about implying that something is known about the mechanism of action. The grouping, since it is not based on knowledge of carcinogenic mechanism in vivo, may have no relationship to the multiple routes by which carcinogenesis could be induced in a particular instance (354). It is difficult to be clear about what is the target of a particular agent when the mechanism of effect of any agent is still unknown. However, it is still important to define the kinds of promoters and to examine the patterns of exposure to them to help elucidate mechanism.

SPECIFICITY/ANTIPROMOTION. Tumor promoters often display tissue specificity, having primary effects on only one or a limited number of tissues in a particular species. For example, estrogens seem to be promoters in breast (319) and liver (320) in animal models. Mouse skin and bladder are also specifically responsive to certain promoters (311). An interesting sidelight to this aspect of promotion is that the PDGF-like material stimulated by the sis oncogene is active specifically for the tissue which develops the sarcoma produced by the virus (102). Since some promoters seem to be simulating growth factors, and many growth factors tend to be fairly tissue-specific, this may be an important source for the tissue specificity seen.

The potential for any tissue to give rise to a tumor after initiation may be partly determined by inherent biological processes; treatment to prevent disease may be designed to interfere with those processes, especially with endogenous tumor promotion. Such inherent processes may be altered by diet, stress, or other conditions such as individual genetic differences or altering physiological function. For example, a number of experimental animal studies suggest that a high fat diet can promote carcinogenesis in breast and colon, while epidemiological studies show a positive relationship between dietary fat and breast cancer in some populations (355). However, to confirm that a low fat diet will decrease these types of cancer (13), additional studies need to be conducted in countries with a wide genetic diversity where a high proportion of the population's caloric intake comes from fats. It must be kept in mind that the total carcinogenic response should be considered. For instance, there are certain populations, with certain lifestyles and diets, such as the Japanese, which have a low incidence of one cancer, such as colon cancer, when compared with other cultures (e.g., Americans). However, they frequently have a high incidence of some other type of cancer, such as stomach cancer, which has a low incidence in other populations (3).

Studies have also revealed several compounds, called antipromoters, that inhibit responses to PMA in vitro (148) and prevent tumor promotion in vivo (356). Antipromoters are effective during different phases of the multistep process of promotion. Although "pure" promotion has not been conclusively demonstrated in humans, the existence of antipromoters in other systems has been used to suggest that it may be possible to design cancer prevention methodology based on the interference of promotion with a putative antipromoter in humans. Clinical studies are currently underway to test the therapeutic efficacy of some antipromoters, e.g., synthetic retinoids (357).

While cell culture methods offer possibilities for screening for tumor promoters and antipromoters, it will first be necessary to determine which functions altered *in vitro* by promoters are critical to the promotion process and which are secondary. The species and tissue specificity of tumor promoters make it likely that a number of different cell types will be required for any screening procedure.

NEOPLASTIC PROGRESSION. The progression of a neoplastic cell into a malignant tumor, which does not occur to every neoplastic cell, is the last major stage of cancer formation and is probably accomplished through a number of steps. Many cancers have more than the normal numbers of chromosomes and exhibit poorly controlled and sometimes higher rates of growth, invasiveness and anaerobic glycolysis, factors that may be important to their survival in the animal.

The route by which a neoplastic cell becomes a cancer may entail a sequence of interactions between tumor cells and both specific and nonspecific host defenses (358), as well as a series of events that result in the cell acquiring more of the characteristics of cells in a malignant neoplasm (283). These selection processes are mutually nonexclusive and may involve karyotypic changes (359). The factors that are important in neoplastic progression, especially in the premalignant stage after neoplastic cells emerge and before a malignant neoplasm is formed, may be similar to those in tumor dormancy, a phenomenon in which tumor cells reappear many years after apparently successful treatment. The major mechanisms of tumor dormancy seem to be (1) inhibition of cell division and (2) cell division accompanied by cell lysis. Mitotic control can be exercised by macrophages (360), soluble factors such as lymphokines (361) and interferon (362), nutrient depletion (363), or a hormonal requirement for growth (364). Continuing cell division in a tumor, the major characteristic of a neoplasm, can continue without much effect if daughter cells are lysed about as fast as they are produced. This can result from immunological lysis, nutritional deprivation, or very slow tumor cell production. Any or all these factors may lose their effectiveness during the selection processed for malignancy described above, perhaps in a number of steps.

An interesting model for the mechanism of progression of a neoplasm to malignancy is the one proposed by Gersten, Fidler, and Hart (360,365). After a primary tumor is established, there is tumor vascularization, by an angiogenic factor (363), with invasion of surrounding tissue and blood vessels. This invasion seems to be accomplished through tissue damage by a number of different enzymes, such as collagenase (361) and cathepsin B (362), although the role of tumor cell motility is not clear (365). The widespread dissemination of cancer cells occurs by the lymphatic (carcinomas) and hematogenous (mesenchymal tumors) routes. During lymphatic spread, the regional lymph node is involved in the host immune responses to neoplasia (368). During hematogenous spread, release of tumor emboli in the bloodstream results in most tumor cells being destroyed by interaction with a number of blood elements (369). However, the more that are released, the greater chance of survival to form metastases. Arrest and subsequent growth of circulating tumor cells or implantation in another site seem to be a function of tissue and metastasis, and there are some data that the site is selected by the membrane properties of the metastatic cells (370). Metastatic cells then leave the blood vessel and form micrometastases which induce blood vessel proliferation leading to metastatic growth (365).

Progression seems to be modulated by a number of factors. Important among these is the immunological system. Many tumors induced by radiation and chemical agents are antigenically distinct (371-373). To overcome the immunological defenses of an immunocompetent individual, the tumor could (1) simply not be immunogenic, e.g., contain only antigens expressed normally in the body, or (2) suppress the immunological response. Lack of immunogenicity seems to be very common in spontaneous tumors in animals (373,374). This also seems to be the case for humans since, at present, only cells from presumably viral-induced tumors such as Burkitt's lymphoma (375) may be considered to be truly immunogenic (although there is evidence of some specific antigens associated with cells from human tumors grown in culture, e.g., colorectal tumor cells (376,377). A tumor could also induce immunodeficiency, either general or selective. Immunosuppression could be accomplished by a variety of mechanisms, including production of circulating immune complexes (378) (to inhibit antibody response) or a rise in suppressor T-cells (379) (to inhibit T-cell response). Production of an immunodeficiency seems to be important for highly immunogenic tumors, such as UV-induced tumors, which are decisively affected by suppressor T-cells in the formation of primary tumors (379), and viral tumors, which are profoundly influenced by immunocompetence (139,380). The immunodeficiency associated with UVinduced tumors is a selective one, perhaps because of a UV-induced deficit in antigen processing by a macrophage-like cell that subsequently produces the T-cell

Clinically, the term "immunodeficiency" usually refers to a more general type of immunodeficiency that can occur during the later stages of carcinogenesis (381). This may be of importance mainly in metastasis, since it has been shown that immunosuppressive treatments and agents greatly increase the incidence of distant metastases in rats with highly immunogenic tumors [though the effect is reversed for tumors with low immunogenicity (382)]. Its role may be wider, however, since it has been suggested, from evidence about lymphoma in immunosuppressed individuals, that the immune system may be important in the selection of a particular chromosomal alteration, i.e., may be involved in promotion (139).

Implicit in this analysis is the fact that factors that can alter modulators of progression can also alter progression. For example, factors such as stress that result in a mild immunodeficiency may also mimic the effects of immunosuppression in production of lymphoma. A cancer is a product of a long series of events, and it is a reasonable assumption that modulators may interact in a positive or negative fashion with this sequence at any point.

An interesting approach to this complex problem was the development of a general model for the temporal development of tumors, rooted in evolutionary theory. Based upon the observation of tumor heterogeneity, even when derived from a single clone, the "shifting balance" theory, which emphasizes that many genes, with multiple pleiotropic effects mediate the expression of tumor genotype, was applied to tumor evolution (383). This "micro-evolution" maintains the flexibility for rapid population adjustment to conditions because of a wide variety of phenotypes. "Macro-evolution" occurs when a new niche is available, after survival of a "catastrophe" for the cancer cells (resulting from clinical treatment or immunological attack), or when a new adaptive level emerges from previous micro-evolutionary changes. The implications of this theory are intriguing and suggest that cancer prevention may be more effective than cure.

Multiple Agents

Since people are exposed to many different agents at the different times in different sequences, the effect of multiple agents on carcinogenesis is of major concern. However, there is little information of general import in the field. Models for interaction are generally limited by lack of information on dose-response curves for carcinogens in the area of interest (384). The great number of permutations of possible agents and doses makes understanding interaction of multiple agents very difficult.

In general, the action of two or more agents can be additive (if the agents are given in a dose range where the biological response is a linear function of dose) or multiplicative (if the response is a simple exponential response to dose), synergistic (greater than expected) or antagonistic (less than expected) (384,385).

One obvious example of synergism is the tumor initiator-promoter interaction described above. Tumor incidence is much less when either agent is given separately than when the two agents are given in the proper order. Another possible example is the relation between cigarette smoking and asbestos exposure in carcinogenesis. The effect of both carcinogens is much greater than a simple additive model would predict and is closer to multiplicative (386). Although the mechanism for this phenomenon is not known, there are several possibilities: (1) asbestos increases the uptake of the carcinogenic substances in cigarette smoke [as shown when benzo(a)pyrene is used as a model compound (387)]; (2) asbestos stimulates metabolism of carcinogen(s) in smoke (which has been shown for benzo(a)pyrene, one component of smoke) (388); (3) asbestos works as a promoter with cigarette smoking the initiator (389); or (4) some combination of the above mechanisms (390).

One example of antagonism is the interaction of 3'-methyl-4-dimethylaminoazobenzene (an azo dye) and certain polycyclic aromatic hydrocarbons (PAH), such as 3-methylcholanthrene (MCA) (391,392). Some carcinogenic PAH, by stimulating metabolism, inhibit azo dye carcinogenesis. Although the study and understanding of the interaction(s) of multiple agents is not easy, it becomes much more difficult (conceptually as well as experimentally) when it is not clear whether more than one agent is involved. An example is the interaction of immunosuppressive agents and EBV. Al-

though immunosuppressive agents are presumed not to induce tumors, their use may allow the expression of EBV induced carcinogenesis, as has been suggested for renal transplant patients who have a substantially raised incidence of tumors, especially lymphomas (139). Since EBV is ubiquitous, exposure to an immunosuppressive might result in an increased number of tumors in a bioassay (i.e., be carcinogenic), although in fact it is simply allowing expression of tumors that arise from a viral infection. Other agents may alter the modulators at any stage in carcinogenesis and result in either more intense effects of a ubiquitous agent or the expression of a tumor derived from long-lived, irreversibly initiated cells.

These examples illustrate just a few of the possibilities of interaction. Some agents may stimulate metabolism, interact chemically with other agents, or be so cytotoxic that tumors induced by another agent are killed. Other agents may alter modulators of different stages of carcinogenesis or stimulate the expression of tumors that would normally not be expressed in the lifetime of the animal. At present, in the absence of a well established and comprehensive theory of carcinogenesis, the complex interaction of agents is best understood by elucidating the action of each compound. Then the effects of each compound on the organism can be evaluated and their interaction estimated.

Summary

Although the study of the phenomenology of cancer was initiated many years ago, analysis of the mechanisms involved in carcinogenesis was started much more recently, and the level of activity has risen significantly in the last decade. This review has attempted to place much of the data concerning cancer into a broad perspective. Exposure to a carcinogen begins at the interface of an animal with its environment, and an agent can be modified by factors associated with an organism at that interface. An example of this is the reduction of benzidine-based azo dyes to carcinogenic benzidine by gastrointestinal microflora. Since the microflora can vary with species as well as with food habits, this provides one source of variation in the activation of agents.

There is a wide variety of processes that are involved in the activation and detoxification of most agents. The complexity of these processes is illustrated by the major products of the metabolism of benzo(a)pyrene, a carcinogen which needs activation. Lifestyle, exogenous agents, diet, age, etc., can act on several levels to alter the balance of activation and deactivation. Examples of this are the stimulation of cellular metabolism producing more reactive species, the inhibition of a particular pathway resulting in a smaller amount of reactive moiety, and the alteration of cell scavengers that absorb reactive metabolites produced by metabolism, and lower the overall concentration of reactive moiety. Activation and deactivation are functions of species, tissue, and even individuals, providing another source of variation in response to a carcinogen.

The interactions of carcinogens with DNA, which seem to be important for the carcinogenic action of many

initiators and complete carcinogens, vary across the different classes of carcinogens. Biological carcinogens, such as the acute transforming retroviruses, seem to insert genetic information which allows the expression of certain cellular onc genes whose products are important for transformation. Some of these products seem to have effects like certain growth factors. The discovery that the dominant transforming genes of certain human cancer-derived cells are similar to certain classes of onc genes derived from the acute transforming retroviruses has been a breakthrough in understanding certain causal factors in human neoplasia. Radiations damage DNA, but, with the exception of ultraviolet light (UV), the irradiation products are not very well characterized, and the role of the damage is not clear. It has been shown that a particular DNA damage is involved in the carcinogenic action of UV light, but how this occurs is also not clear. The specific damage that many chemical carcinogens produce in DNA is much better characterized, but which lesions are important in carcinogenesis and how a lesion can interact to produce a cancer are not known. "Physical" carcinogens include a number of very different agents and processes which seem to have in common the production of a chronic fibrotic reaction.

Two modulators of DNA damage are DNA repair and fidelity of replication, which help to determine the effects of a carcinogenic stimulus that survives through replication in initiated cells. Recent work on DNA repair has suggested the importance of the ADP-ribosylation system and has determined that there is a set of enzymes, the DNA glycosylases, that confer specificity in base excision repair for different types of DNA damage. Also, post-replication repair has been found to be more complex than thought previously, which may help explain how a specific lesion influences DNA. Since DNA repair is modified by many factors and is tissueand species-specific, this provides another source of variation in the final response of the organism to a carcinogen. An example of how knowledge about mechanisms impacts on risk assessment is the question of how DNA repair is related to biological thresholds for effects of carcinogens. Statements that DNA repair results in a threshold for carcinogenesis cannot be substantiated.

The fidelity of DNA replication can be altered by a damaged template or, more indirectly, through changes in the reaction conditions that result in miscoding. Any of the reaction conditions could be altered by a carcinogen or by a change in physiological state induced by a carcinogen, providing a route by which the DNA could be altered without the formation of direct binding between the DNA and a carcinogen derivative.

Cancer is a phenomenon with many steps. In an experimental paradigm, two major stages can be demonstrated: initiation, the induction of an irreversible alteration of genetic information, and promotion, a reversible process terminated by the final irreversible alteration of initiated cells into neoplastic cells. Each of these stages can be subdivided. The progression of neoplastic cells into a cancer, which has not been easy to

separate experimentally from promotion, is the final step in carcinogenesis.

Promotion can occur with both exogenous and endogenous compounds (and, perhaps, certain surfaces) and is characterized in a number of systems by a focal proliferation of cells of which a small fraction will persist after the promoter stimulus is removed. Few focal proliferations in any experimental system will become neoplastic. Although the aspect of tumor promotion which is critical for tumorigenesis is not entirely clear, an interesting possibility is the induction of chromosomal alterations. The relationship of this process to viral carcinogenesis and the mechanisms by which alterations may lead to a neoplasm are explored. However, a causal relationship of chromosomal changes to carcinogenesis has not been proven. Tumor promoters are tissue and species specific, perhaps providing some insights into how various tissues and species are differentially affected by carcinogens.

Neoplastic progression, the process in which a neoplastic cell becomes a malignant tumor, seems to involve several steps, none of which is yet well understood. In that complex process, the role of the immunological system as one of the important modulators of progression is discussed. An interesting recent approach has been to consider the progression of tumors from an evolutionary, ecological point of view.

Evaluation of the effect of a number of agents acting concurrently has barely begun. Models for summation, potentiation, or inhibition of effects are poorly developed, especially when considering the range of effects that agents have on a number of metabolic and physiological endpoints. Variation with species, tissue, and individual at every stage in carcinogenesis produces the added uncertainty if two or more compounds interact at many levels.

Short-Term Tests for Potential Carcinogens*

Introduction

Cancer can be induced by a variety of radiations, biological, "physical" and chemical agents and is manifested in a variety of phenotypes. Notwithstanding our lack of understanding of the mechanisms by which cancer is induced and expressed, there is an urgent need to identify the agents that cause and promote these diseases. The rapid identification of potential chemical carcinogens is one of the major goals of short-term testing.

The hypothesis that conversion of normal cells to neoplastic cells results from heritable genetic changes the somatic mutation theory of cancer—was proposed early in this century (393) and forms a basis for the development of many short-term tests. The relationship of mutation and carcinogenesis has been stressed for

^{*}Although the full committee participated in the framing of this section, primary responsibility was assumed by Dr. R. Tennant, assisted by Drs. R. Langenbach, M. Shelby and E. Zeiger of NIEHS.

some time (394). However, early studies relating mutagenesis and carcinogenesis were generally unsuccessful, due in part to the need for metabolic activation of most carcinogens. Pioneering studies by the Millers and associates (395) on metabolic activation of chemicals and the coupling of metabolic activation systems and mutable targets (396,397) allowed further elucidation of the possible relationship between mutagenicity and carcinogenicity. During the past 15 years, many short-term systems that detect mutagenesis and other genetic effects have been developed and attempts have been made to correlate the results of these tests with carcinogenesis. The tremendous number of chemicals that remain to be evaluated for potential hazard and the mounting cost of currently acceptable rodent carcinogenesis assays amply justify the continued effort to develop and utilize short-term tests, and to establish a mode for their application in assessing the potential chemicals to induce cancer.

The search for tests with the capability to identify potential chemical carcinogens has yielded a variety of assay systems with a diversity of endpoints. The existence of such a large number of assay systems (approximately 100 have been described) (398,399) reflects both limitations and differences between assays, as well as our uncertainty about the mechanisms of induction and progression of carcinogenesis. As described in the section on current views on the mechanisms of carcinogenesis, the induction of cancer may have many potential mechanisms (gene or chromosomal mutation, heritable changes in DNA transcription reflected in induction or cessation of gene expression, etc.), but no causal relationship has yet been established between any mechanism of induction and any specific neoplasia. In addition, a variety of factors or conditions, including tumor promoters, hormonal levels, rate of cell proliferation, cell type specificity, and genetic predisposition, are known to influence the expression of neoplasia, regardless of the nature of the inducing agent. Because of the relatively small number of known human carcinogens, the predictive capability of the short-term assays is usually assessed by validation with the many more chemicals which are carcinogenic in long-term animal tests.

This section describes the major categories of shortterm assays for chemical carcinogens, attempts to evaluate the assay systems, and recommends further studies required to define their predictive ability. The term "genetic toxicity" is used to indicate any effect that can be attributed to an interaction between the test agent and the genetic material of the test organism. There have been attempts to classify carcinogens on the basis of whether they show evidence of direct interaction with DNA or act by some indirect effect ("epigenetic") (400). However, the assay systems conventionally used to make this distinction may not detect all types of chemical-DNA interactions, nor do they adequately accommodate the tissue or cell-specific metabolism or target specificity which many carcinogens demonstrate. It is thus not possible conclusively to make a clear distinction between genetically toxic and "epigenetic" on the basis of currently available *in vitro* tests. However, the fact that certain factors or substances do appear to induce tumors without appearing to react directly with the DNA indicates a need for development of short-term tests complementary to those which detect direct effects on the DNA, if all types of chemical carcinogens are to be detected.

Many of the tests for genetic toxicity are also useful in the identification of potential germ cell mutagens; however, this represents a distinct application of the tests, and the reader is referred to the recent report of the National Academy of Sciences (401) for further information on this subject. In addition, screening of chemicals for genetic activity can identify agents that may be involved in the aging process or atherosclerosis (33,402).

Short-Term Test Systems

There have been numerous descriptions of assays that have been developed and evaluated for their capability to detect potential carcinogens, but the details of these systems will not be discussed herein. The reader is directed to the EPA Gene-Tox Evaluations (403–431), the International Collaborative Program (432), and the review by Hollstein et al. (398) for details and further references to the individual systems. The four principal biological endpoints represented by the available tests are gene mutation, chromosomal effects, general DNA damage, and neoplastic transformation. Some frequently used assays are listed in Table 4.

GENE MUTATION. Although it is possible to assay for chemically induced mutation with a variety of methods in a number of organisms and cells, the greatest amount of information on tests for potential carcinogens is derived from a relatively few systems. The system in which the largest number of chemicals has been evaluated is the Salmonella/microsome test (Ames test) (433,434), where several strains of the bacterium Salmonella typhimurium have been genetically altered in order to provide increased sensitivity to potential mutagens. In the Ames assay, reverse mutations in the histidine locus (i.e., from a histidine requirement to independence) are detected. By virtue of the sensitivity, ease and relatively low cost of performance and duration of the test (48–72 hr), this assay has been used worldwide to test for chemical mutagens. Other microbial systems can also be used (Table 4) but have not been used nearly as extensively as the Salmonella assay. Cultured mammalian somatic cells (which are genetically more similar to cells as they exist in humans than are bacteria) such as the mouse lymphoma (L5178Y) (430), Chinese hamster ovary (CHO) (405), and Chinese hamster lung cells (V79) (409), have also been used to detect gene mutations induced by a variety of chemical agents and radiations. The various systems that have been developed are capable of detecting forward and/or reverse mutations. However, none of the specific gene loci conventionally used in the bacterial or mammalian cell mutagenesis systems (his, tk, oua, hgprt, etc., Table 4) appear to be related in any way to the cellular changes

Table 4. Conventially used short-term assays.

Type of assay	Endpoint	Reference
Mutation Microbial systems Salmonella typhimurium	Histidine reversion	(433)
${\it Escherichia~coli}$	Tryptophan reversion	(403)
Mammalian cell systems Mouse lymphoma (L5178Y)	Thymidine kinase (bromodeoxyuridine or triflurothymidine [TFT] resistance)	(430)
Chinese hamster ovary (CHO)	Hypoxanthine guanine phosphoribosyl transferase (6-thioguanine [6TG] resistance)	(405)
Chinese hamster lung (V79)	Hypoxanthine guanine phosphoribosyl transferase (6TG resistance) Na/K ATPase (ouabain resistance)	(409)
Human lymphoblasts	Thymidine kinase (TFT resistance) hypoxanthine guanine phosphoribosyl transferase (6TG-resistance)	(435)
Chromosome effects Many cell types in vitro and in vivo	Chromosomal aberrations Sister chromatid exchanges Micronuclei Chromosome gain or loss Translocation	(408–410)
DNA damage Unscheduled DNA synthesis (UDS)	Radioactive nucleoside incorporation	(431)
Repair deficient bacteria	Differential growth inhibition	(411)
Mammalian cell transformation Cellular		(426)
Syrian hamster embryo (SHE), Balb/c 3T3, C3H/ 10T1/2	Altered cellular morphology and growth pattern, growth in soft agar, tumorigenicity	
Viral enhanced/mediated	Same as above in appropriate host	(426)
Syrian hamster embryo (SHE)/Simian adenovirus type 7 (SA7), Fischer rat embryo cells/Rausher leukemia virus		

that occur in the induction of neoplasia. These loci are used to measure the capability of a chemical to interact with the DNA of an organism in such a way as to give rise to mutagenic effects. Although it has been widely assumed and theorized that neoplasia arises from mutation, it is known that heritable phenotypic changes also occur in mammalian cells in the process of differentiation and growth and such changes arise without the induction of gene mutation. Consequently, it is necessary to use additional biological endpoints that reflect the capacity of chemicals to cause changes through other types of interactions with the genetic material.

CHROMOSOME EFFECTS. Changes in the structure or number of chromosomes occur frequently in tumor cells (436), and cells from individuals exposed to radiation show evidence of chromosome damage (437). Cytogenetic short-term tests have demonstrated that a number of carcinogens can cause structural alterations of chromosomes (aberrations) in the bone marrow of exposed animals or in cultured cells derived from various tissues (410). There is increasing evidence that associates spe-

cific chromosomal rearrangements (translocations) with specific neoplastic phenotypes (438). Although no definitive causal associations have been defined, the use of molecular and recombinant DNA techniques promises to aid in resolving the relationship between translocation of large segments of chromosomes and the specific DNA sequences involved. Another assay often used in conjunction with the search for chromosomal aberrations is the induction of sister chromatic exchanges (SCEs) (439,440) that are believed to result from the induction of recombination by DNA damage. However, the association between an increase in the frequency of SCEs in cells in vitro or in vivo and heritable changes has not been clearly defined.

It is also possible to identify the production of chromosome fragments (micronuclei) in reticulocytes or erythrocytes (441). Induction of micronuclei has been associated with chemical exposure and is also believed to be indicative of the ability of chemicals to induce chromosomal aberrations, but it is also not yet clear that the induction of micronuclei is related to heritable

changes in cellular phenotypes.

A long-range potential value of tests for chromosomal effects is that they may be used to assess the comparative (trans-species) toxicity of chemicals. Since the sampling for cells is nondestructive to the exposed animal or individual, and because the assays can be conducted on cultured cells, comparisons are possible between various species and humans. Additionally, the monitoring of human blood cells for chromosomal effects may be a valuable method to assess human exposure to hazardous chemicals (442). However, such comparisons must be conducted with the understanding that chemical disposition (pharmacokinetics) may vary with species thus giving rise to species-dependent target cell populations.

DNA DAMAGE AND REPAIR. These tests do not measure mutation per se, but DNA damage, or the cellular repair of such damage, induced by the chemical treatment of microbial or mammalian cells. The most extensively used method is to detect non-S-phase DNA synthesis (unscheduled DNA synthesis, UDS) in hepatocytes (431,443), although other cell lines (431) have also been used. Techniques that measure DNA strand breaks (414) or binding of chemicals to cellular DNA in vivo or in vitro (444) are also used as assays for DNA damage. Other assays use strains of E. coli or B. subtilis and are based on differential growth inhibition of DNA repair proficient and deficient strains (411,445).

CELLULAR TRANSFORMATION. Exposure of cultured mammalian cells from a variety of tissues and species to carcinogens (biological, chemical and radiations) has been shown to result in certain heritable phenotypic changes. A variety of these changes has been described (426,446), but some specific morphological changes have been shown to be highly associated with the capacity of the cells to induce tumors when inoculated into appropriate recipient animals. Because of this ability of transformed cells to produce tumors in recipient hosts, these assays have been considered by some to be the shortterm systems most relevant to detection of chemical carcinogens (397,426,442). A number of cellular systems derived from rodent embryos have been used to identify chemical carcinogens (Table 4), some of which use concomitant infection with transforming or nontransforming viruses (426,446). There is no clear understanding of the mechanism of in vitro transformation, and it is likely that the various heritable alterations can occur through one of several possible mechanisms. Recent innovative advances at the cellular and molecular level are having a major impact on our understanding of this process (447,448). As the implications of studies on oncogenes and transfected transforming genes become better understood and are incorporated into methods for detecting carcinogens, there is the promise of significant improvement in the accuracy with which these systems can be used to detect carcinogenic chemicals.

IN VITRO PROMOTION SYSTEMS. The existence of chemicals which appear to cause (or promote) cancer by mechanisms which may or may not directly involve the

genome has necessitated the development of short-term systems that can detect these types of chemicals. However, it should be emphasized that chemicals may have both initiating and promoting activity, and the two properties are not mutually exclusive. The V79-metabolic cooperativity assay (449) has been the most thoroughly studied in vitro system developed to detect chemicals with promoting activity. A basis for this assay has been attributed to be the interruption of cell-to-cell communication, although the exact mechanism is unknown. Further studies with this and other promotion systems are needed before they could be reliably used as test systems to detect chemicals that act as promoters of tumorigenesis.

IN VIVO SHORT-TERM BIOASSAYS. Several short-term in vivo bioassays to detect carcinogens and/or promoters have been developed. Of these assays, the mouse skin papilloma system and the strain A mouse lung adenoma have been used for testing, while the liver foci bioassay is still in developmental stage. These systems are generally viewed as intermediate in biological relevance between in vitro systems and the long-term animal bioassay, because in vivo metabolism and/or chemical disposition are provided for by in vivo exposure. However, as with the in vitro tests, the relevance of the endpoints measured to the development of malignant tumors remains an open question.

The mouse skin papilloma system, first reported by Berenblum (450), is based on a multistage (initiation and promotion) hypothesis of cancer induction and has been developed further in other laboratories (314,327,352,451,452). The system relies on the induction of benign papillomas, although carcinomas do develop in the later course of the assay. Variations in experimental test design reportedly allow chemicals to be classified as: initiating carcinogens, promoters, complete carcinogens (chemicals with both initiating and promoting activity), co-carcinogens, and co-promoters (452). Several strains of mice have been used in the assay, but the SENCAR stock appears particularly sensitive (452). The mouse skin papilloma assay is the in vivo system in which the greatest numbers of chemicals have been tested (453); however, many of the chemicals investigated have been polycyclic aromatic hydrocarbons, and the usefulness of the system to test other classes of chemicals, or chemicals with organ specificity other than skin, is not presently clearly known.

The strain A mouse lung adenoma system was developed by Shimkin and colleagues (454–457). Although lung adenomas do occur spontaneously in mice, the strain A mouse, unlike other strains, has nearly a 100% lifetime incidence (457). Therefore, the assay appears to rely on decreasing the time-to-tumor onset by the test chemical, although Stoner and Shimkin consider the test chemical to be inducing rather than accelerating the process (456,457). A wide spectrum of chemical classes has been tested in the lung system (457), and good correlations of results and the tumorigenicity of the chemicals have been reported, although one study using diverse coded chemicals did not find a high cor-

relation (458). The system has been criticized because the adenoma is a benign growth and an analogous tumor does not appear in humans.

The induction of altered foci in rat liver systems have also been studied for use as short-term in vivo assays for carcinogens and promoters. However, these systems are relatively new and are still in a developmental stage compared to the mouse skin papilloma and lung adenoma assays. The rodent liver systems have been used by Peraino et al. (459), Pitot et al. (460), and Solt and Farber (461), to study the initiation/promotion phenomena. The endpoints that have been measured include the appearance of foci in the liver which have increased gamma-glutamyl transpeptidase, basophilia, or diaphorase; or foci with decreased glucose-6-phosphatase, adenosine triphosphatase or iron storage (462). Chemicals can be tested either for initiating or promoting activity in the liver assays. Since defined protocols have not yet emerged, further development is required before the possible value of the approach in a routine testing program can be assessed. In addition the relationship of the altered liver foci to hepatocellular neoplasms remains to be determined.

Factors in Test Evaluation

METABOLIC ACTIVATION. Most target cells used in short-term tests have limited endogenous metabolic capability for xenobiotics and, therefore, must be supplemented with an exogenous metabolic activation system. The metabolic activation systems employed in shortterm tests may be one of the major limitations in these assay systems, because a chemical may show a different metabolic profile in vitro from what would occur in vivo. From a testing point of view, a major concern is that specific activating enzymes or pathways may be decreased or lost during preparation of the metabolic activation system, thereby giving a false negative result with the test chemical. Alternatively, enhanced activation of certain chemicals would also be possible (false positive). Essentially, three types of metabolic activation systems have been used with target organism host-mediated (415,463), intact cells (464,465), and organ homogenates, usually 9000g supernatants called S9 (33,434). Most short-term assays have used rat liver S9 preparations because of the ease of preparation and storage, capability to activate many chemical classes, and the relative compatibility (nontoxicity) with most target organisms used in genetic toxicity testing. However, it is also known that liver S9 preparations may cause different responses in vitro from what would occur in vivo due to factors such as: organotropic differences, chemical induction of S9 preparations, and destruction of certain activation enzymes during S9 preparation, decrease in conjugation capability, etc. Therefore, the inability of S9 preparations to mimic in vivo activation has provided the impetus to modify S9 utilization and/or investigate other metabolic activation

Intact cells are another exogenous metabolic activation system used in short-term tests (465). These cells can be primary cultures prepared from various organs

(liver is most common), embryonic cells that have been cultured only a short time, or in certain cases, established cell lines. For such an activation system to function, the test chemical must be taken up by the activating cell, be metabolized, and the activated product be transferred to the indicator cell. Thus, a negative response with a cellular system may be a function of the uptake of the test substance and the transfer and stability of active metabolites. Since most cells lose significant metabolic capability with time in culture, certain enzyme systems may be functional at reduced levels. However, this loss of activation capability can be at least partially overcome by the use of freshly prepared primary cells.

In the host-mediated assay, the indicator organism is introduced into the peritoneal cavity or injected into the circulatory system of an intact mammal (415,463). The animal is then treated with the substance under test, sacrificed, and the test organism removed and examined for mutation induction. A wide range of indicator organisms has been used (415). Because an organism foreign to the host is commonly used as the indicator organism, problems arise such as interaction between the host and the indicator organism, including immunological interactions, and the limited time the indicator organism can be kept in the animal. Therefore, this system is unable to detect many carcinogens and chemicals which show in vitro mutagenesis.

Attempts to improve metabolic activation systems are certainly worthwhile and have increased our ability to detect carcinogens by in vitro methods. One must accept, however, that it is not possible to mimic in vivo metabolism in a petri plate and the negative in vitro results may represent false negatives and positive in vitro results may represent false positives. These possibly erroneous test results due to metabolic activation can sometimes be resolved through the use of short-term in vivo rodent assays. A testing scheme has been proposed by Ashby (466) whereby short-term rodent assays are employed to study further chemicals falsely identified in the in vitro systems.

STATISTICAL EVALUATION OF SHORT-TERM TEST DATA. The overwhelming majority of short-term test data in the literature has not been statistically evaluated. In general, decisions of "positive" and "negative" are made on a subjective basis or on the basis of simple statistical procedures. Recently, several statistical procedures have been proposed for the Salmonella assays (467–469), the mammalian cell mutation (470) assays, and discussions of statistical approaches appear in the respective Gene-Tox reports, but there is no consensus as to which particular procedures should be used for data evaluation.

FACTORS IN THE CORRELATION OF ASSAYS WITH CARCINOGENESIS. Because short-term tests for carcinogens employ neither the organism (humans) or the biological endpoint (cancer) of concern, there is a need for a great deal of extrapolation of test results; hence, the demand for proving the validity of the tests is great. However, after more than a decade of effort to dem-

onstrate the utility of short-term tests in predicting carcinogenicity, it is still difficult to find sufficient test results to assess the performance of an assay within or among chemical classes, the activity of a chemical in a variety of tests, or the performance of a group of the tests on even a small group of chemicals. This deficiency in the existing data results largely from two factors. First, there has not been enough of an effort to evaluate systematically a large number of chemicals in a group of selected assays, and second, there is a marked deficiency of genetic toxicity results on noncarcinogens. This latter point presents an immediate obstacle to the evaluation of short-term tests for carcinogens, since in the absence of an adequate data base on noncarcinogens, it is not possible to determine the performance of a single test or group of tests in discriminating between carcinogens and noncarcinogens.

Definition of Some Terms. The reliability of a shortterm test system may be defined as its ability to predict accurately a chemical's in vivo carcinogenicity. The standard against which short-term test results are most often compared is the rodent cancer bioassay. A shortterm test which correctly identifies, when used without bias, a high percentage of known carcinogens and noncarcinogens is assumed to be capable of predicting the carcinogenicity of chemicals of unknown activity with the same degree of accuracy. The short-term assay's performance can be expressed by three criteria: sensitivity, specificity, and accuracy. Sensitivity is the proportion of carcinogens giving positive results; specificity is the proportion of noncarcinogens giving negative results; and accuracy is the overall proportion of correct results. Unfortunately many, if not all, previous test system evaluations, have been characterized by an inadequate number of noncarcinogens, and, hence, a determination of specificity and accuracy is severely limited. Also, the accuracy of short-term tests could be altered by the ratio of carcinogens and noncarcinogens in the test sample (471). Additionally, the expected accuracy of a test battery (see below) is influenced by the number of tests in the battery and the number of individual tests required to be positive before the chemical is considered to be positive in the battery (472).

Noncarcinogens and Test Analysis. The inadequate identification of, as well as the use of, noncarcinogens, as has been noted, is one of the key problems. Many chemicals have been identified as noncarcinogenic merely on the basis of their chemical structure or perfunctory carcinogenesis tests. To date, relatively few chemicals used as noncarcinogens in short-term tests have been tested in lifetime studies using adequate chemical exposures. A major source of information on noncarcinogens is that of the NCI/NTP rodent bioassay where two sex/species are exposed at sufficient concentration of the chemical and for sufficient duration to insure that the chemical has been tested adequately. However, few of these noncarcinogens have been tested in most genetic toxicity assays (473). In addition, factors that determine a "positive" or "negative" short-term test response for a test chemical can vary from laboratory to laboratory or even experiment to experiment within a laboratory. Often parameters such as control background levels, acceptable range of positive control responses, statistical significance of test chemical response, etc., all of which are important in determining whether a chemical is "positive" or "negative," can be subjective. Such uncertainties could lead to an inaccurate estimation of the sensitivity and specificity of short-term tests.

Test Endpoints and Carcinogenicity. The development of the Ames test and subsequent short-term tests was predicated on the assumption that most carcinogens were mutagens and, conversely, that mutagenic chemicals will be carcinogenic. Implicit in this reasoning is the hypothesis that nonmutagens will not be carcinogenic. However, this hypothesis put forth by Ames et al. (397) and supported by others (474–478), that carcinogens are mutagens and the majority of noncarcinogens are not mutagens, may be based on biased data. The majority of the data used to justify this statement comes from studies where the identity of the chemicals and their carcinogenicity were known to the investigators before they were tested. Because the investigators knew the identity of the chemicals and their "expected" mutagenicity, the test protocols could be modified until the desired results were obtained. While these practices may lead to a true picture of the mutagenicity of a chemical, and are necessary for the assay system development, they probably tend to overestimate the carcinogenicity/mutagenicity correlations.

The correlation of data obtained for other endpoints (i.e., UDS, cell transformation, SCE induction) with carcinogenesis data also is subject to the same limitations. However, the correlations between results from these other systems and carcinogenesis are further complicated by the lack of an adequate data base on chemicals of different classes, and, in some cases, a lack of demonstrated interlaboratory reproducibility of test results.

A method which may establish more objectively the correlations between short-term test results and rodent (or human) carcinogenicity is one where chemicals, coded so their identity is unknown to the persons conducting the assay, are tested, and all the test results are judged by the same criteria. Thus, the one conducting the test does not know whether the short-term test results and conclusions are "right" or "wrong" until after a determination is made. This approach may minimize the proportion of carcinogens detected, but it also eliminates preconceptions about the activity of the chemical. In actual practice, however, additional testing using modified protocols can be done on chemicals whose structure would lead one to believe that the original test conditions were not appropriate. For example, some benzidine-based dyes require reducing conditions, followed by an oxidative system, rather than a strictly oxidative system for their activation. Therefore, choice of test systems should vary with chemical class or nature of the test chemical, but determinations of response should be independent of knowledge of the chemical's identity.

Attempts to Evaluate Short-Term Test Performance. Though not definitive, there have been several systematic efforts to evaluate the short-term assays, singly or in combination, for their ability to detect or predict chemical carcinogens. The largest effort to date was conducted by the Gene-Tox Program of the U.S. Environmental Protection Agency and is a multiphased effort to review and evaluate selected bioassays for mutagenicity and related endpoints using the published literature. Initially, the literature published from 1969 to 1979 was reviewed to assess test system performances, to determine the chemicals tested, and the ability of the various systems to discriminate between carcinogens and noncarcinogens. Reports on the individual systems have been published (403–431). During the second phase, a computerized data base of over 2600 chemicals is being analyzed to determine which tests or combination of tests may be used for routine screening or need further development; determine if specific types of genetic damage are related to chemical class; discriminate between carcinogens and noncarcinogens; and predict heritable mutagenic risk. To aid in evaluating system performance, the Gene-Tox Program convened a panel to review and evaluate the literature on chemicals designated as carcinogenic and to develop a list of carcinogens and noncarcinogens which could be used in determining system performance.

The International Program for the Evaluation of Short-Term Tests for Carcinogenicity (432) was a coordinated effort designed to determine and compare the ability of a series of short-term tests to identify carcinogens and noncarcinogens. Sixty laboratories participated, using their own laboratory protocols, to test coded samples of chemicals supplied from a common source. Test results from this collaborative study led to the following general conclusions. First, while short-term tests are useful in the assessment of potential carcinogenicity, no single test or group of tests was capable of detecting correctly all carcinogens and noncarcinogens. Secondly, there are *in vitro* eukaryotic assays that may complement the bacterial tests by identifying carcinogens not detected in bacteria.

Following the completion of the collaborative study discussed above, a subsequent study was initiated to focus specifically on the evaluation of *in vitro* eukaryotic assays with potential for identifying carcinogens not detected, or poorly detected, in bacterial mutation tests (479).

Other systematic attempts to evaluate the short-term assays include an analysis of their ability to detect pesticides (480) and a series of reports prepared for the International Commission for Protection Against Environmental Mutagens and Carcinogens (ICPEMC) (463,481). The NIH/NTP is currently conducting a systematic genetic toxicity assay of chemicals, both carcinogens and noncarcinogens, tested or under test in the rodent, two-sex/species carcinogenesis assay. The major categories of genetic toxicity are represented, and the chemicals are tested under code.

Practical Assay Utilization

ASSAY SELECTION. The proposed use of short-term tests to detect or predict chemical carcinogens is based on two factors: first, that they require less time to conduct than a rodent carcinogenicity study; and, second, that they are less costly. Currently used tests require from a few days to several weeks to conduct, and the costs may range from \$1000 to \$20,000 per chemical.

Since there has been little success in establishing a cause-effect relationship between specific genetic changes and cancer, all categories of induced genetic change may be potential mechanisms by which cancers arise, and this is reflected in the variety of endpoints detected by short-term tests. Current short-term testing practices generally involve groups of tests selected on the basis of theoretical prudence rather than on an empirical demonstration of their ability to predict carcinogenicity. By theoretical prudence, it is meant that a range of organisms (bacteria, yeasts, cultured mammalian cells, intact rodents) and endpoints (gene mutations, chromosomal damage, DNA damage, and in vitro neoplastic transformation) are employed in an attempt not to miss any effect that might be related to the carcinogenicity of the test agent. However, until the time that basic research on mechanisms of cancer induction provides us with a better understanding of the process, it will be necessary to base testing strategies on a combination of theoretical prudence and correlative evidence obtained by comparing short-term test results with carcinogenicity test results.

In the selection of short-term tests, factors other than ability to discriminate between carcinogens and noncarcinogens must be considered. Reproducibility of test results is a paramount consideration that applies not only to reproducibility within a laboratory but also to agreement of test results among laboratories. When testing chemicals, the ability to reproduce test results qualitatively is more important than quantitative reproducibility. Quantitative differences in test responses represent the degree of response and may be attributed to protocol variables. However, qualitative differences are more serious and affect the judgment of whether a test chemical is "positive" or "negative." Therefore, short-term test selection should be based on reproducibility as well as reliability, sensitivity, specificity, and accuracy. In addition, the degree of development of a particular assay system should also be taken into consideration when selecting tests for use in a screening program. This includes a consideration of the number of laboratories performing the assay, degree of protocol standardization, and a consideration of the number of chemicals and chemical classes which have been tested in the assay.

ASSAY APPLICATION. The application of short-term tests to chemical safety evaluation raises questions at several levels of the process. As discussed above, assay performance must be known to determine which tests are useful. Next, it must be decided how many tests to

use and in what scheme or order they will be utilized. Tier and battery testing approaches have been proposed. In the tier approach, the choice of assay system varies depending on the activity of the chemical in the previous system, beginning usually with the *S. typhimurium* assay. Therefore, decisions are made after each test result is obtained. With the battery approach, the test chemical is assayed in a group (battery) of systems and a decision on the activity of the chemical made when all results have been obtained.

Once test results from the short-term test scheme have been obtained, it must then be decided how to utilize the results. Within the framework of testing or evaluating chemicals for potential carcinogenicity, where possible regulatory action on the chemical may be involved, short-term test results can be used to influence the process at numerous points, including: the nomination and/or selection of the chemical for prechronic and chronic studies; the selection of prechronic studies to be conducted; the decision, used in combination with prechronic results, to proceed with chronic studies; the evaluation of chronic toxicity results, as a source of additional information; the design of postchronic studies, again as an additional source of information, to further elucidate mechanisms of activity.

In the industrial setting, short-term tests can be (and are) used extensively in product development, process improvements, product registration, evaluation of occupational environments, etc.

It is generally agreed that a scheme employing multiple tests is a better approach to carcinogen screening than the use of a single test. The tests used should not be limited to prokaryotic or lower eukaryotic test systems and should include assays for more than one genetic endpoint. A problem that is immediately encountered when multiple test systems are used, however, is a mixture of positive and negative results. The resolution of this problem is not easy, and several authors (482-484) have attempted to deal with the issue. For the present, there are at least three approaches to dealing with such conflicting test results. One may (1) use a weight-of-evidence approach to call a chemical positive by some predetermined criterion; (2) use one's best scientific judgment to reach a conclusion on a chemical-bychemical basis; or (3) simply postpone judgment until the time that a sufficiently large data base is available to permit an unequivocal conclusion from any pattern of positive and negative results. This last choice is rarely acceptable, and current practice generally utilizes some combination of the first two. The systematic accumulation of additional knowledge on cross-system testing of various classes of chemicals should provide insight into the proper choices of short-term systems in assessing potential carcinogenic activity.

FUTURE DIRECTIONS. Short-term tests for genetic toxicity and other effects continue to hold promise of improving our ability to identify potential chemical carcinogens. However, there is at present insufficient data available to justify the use of current tests to supplant

animal bioassays. To improve the interpretation of short-term test results, there is a need for a systematic effort to obtain across-test results using optimal test protocols and coded chemicals, with particular emphasis on noncarcinogens. It is clear from available results that no one assay system is adequate, but it is not yet possible to determine which combination of test systems is most appropriate for most carcinogens. Studies are needed to determine if human cells or tissues would be better predictors of human carcinogenicity than the currently used rodent systems. Some carcinogens, by virtue of their intrinsic mechanism of action, will not be identified by available tests, and we must continue to search for assay systems that accommodate the various molecular mechanisms. Concomitantly, we must continue to improve our understanding of the results of rodent assays for carcinogens. Where there is a dichotomy of results between genetic toxicity or other short-term test systems and the results of rodent carcinogenicity, appropriate follow-up studies in the appropriate systems should be conducted to determine the basis for such differences.

Summary

While many assay systems have been developed and proposed to detect or assess the genetic toxicity of chemicals, they can be generally grouped into four categories: gene mutation, chromosome effects, DNA damage, and transformation. The internal momentum of the field has resulted in the emergence of a few systems or tests in each category that have been used extensively to assess potential carcinogenicity, and the various assays within each grouping have characteristic strengths and weaknesses. The test systems must be demonstrated to have reliability, sensitivity, specificity, and accuracy and to be reproducible both within and between laboratories. The various systems also measure different biological endpoints, and there is no reason to expect that all chemicals will cause an effect in all systems. Since no causal relationship has been established between any specific mechanism of induction and any specific type of neoplasia, we are presently limited in our ability to predict potential carcinogenicity of a chemical, particularly when there is apparent disagreement between different in vitro test results.

The consensus of available information suggests that short-term tests, when properly used and validated, can provide strong indications of potential carcinogenicity. However, based upon our limited knowledge of the mechanisms of cancer induction, it appears that there will be classes of carcinogens that are not detected by currently available short-term tests. Chemicals which act by a genetic mechanism but which have a high degree of organ, species, or sex specificity may be predictably difficult to detect in any *in vitro* genetic toxicity system, since the short-term system may not adequately simulate the basis for these specificities. Additionally, chemicals which appear to act by nongenetic mechanisms would not be detected in currently available short-term tests.

Long-Term Carcinogen Bioassay*

Long-Term Carcinogen Bioassay: Guidelines for Protocols

BACKGROUND. The current carcinogen bioassay in rodents has its origins in research efforts, begun in the 1920s, that sought to determine the carcinogenic potential of chemicals as well as to delineate the mechanism by which they produce cancer (485,486). Animal strains were developed and tested for their specific sensitivity or resistance to certain types of cancer, the central focus of these studies being the understanding of the mechanism of carcinogenesis (485,486). While these experiments led to the induction of cancer in laboratory animals and revealed many new carcinogens, most of the investigators did not intend their highly specialized test systems to be used as routine methods for testing large numbers of chemicals. Only in the mid-1960s, with the advent of the bioassay program at the National Cancer Institute (NCI), did large-scale systematic testing using a relatively standard protocol really begin (487). As a consequence, only recently has there been a reasonably adequate data base on a large number of diverse chemicals tested in a relatively uniform way (488,489). The lack of such a data base, during the earlier developmental period, contributed to some degree of confusion and speculation as to how carcinogens should be tested and how the results from such tests should be interpreted. One finding was clear; known human carcinogens, with the single exception of arsenic, are carcinogenic in appropriately conducted studies in some animal system. [Arsenic has recently been reported to produce carcinomas of the respiratory tract in hamsters (490)]. This does not mean that all chemicals found carcinogenic in animals will turn out to be carcinogenic in humans. Because of differences in the production of critical metabolites and because of other differences between species, a given carcinogen may not produce cancer in all species or in all strains of rodents. Many substances are known that seem to be carcinogenic in one animal species but not in another. But a finding of carcinogenicity in rodents is proof that the chemical is carcinogenic in a mammalian species. And "in the absence of adequate data on humans, it is reasonable, for practical purposes, to regard chemicals for which there is sufficient evidence of carcinogenicity in animals as if they presented a carcinogenic risk to humans" (491). The knowledge and experience accumulated over the last decade, while by no means answering all questions, have helped to focus the issues and have provided a basis for developing some common procedures for conducting and interpreting long-term bioassays.

CRITICAL DESIGN CRITERIA. Several national and international groups have published guidelines on the design and conduct of long-term bioassays (491–502), and the issue continues to be pursued. For example new guidelines are currently being developed by the NTP

Ad Hoc Panel on Chemical Carcinogenesis Testing and Evaluation (503). Essentially, the long-term test involves testing compounds in animals to obtain information relevant to humans. When assessing the results of toxicological testing of any chemical, the limitations of the tests must be borne in mind. Of necessity, animal systems are used, and the results then extrapolated to humans. Such extrapolations may not always be accurate either qualitatively or quantitatively. In some areas of cancer testing, uncertainty still exists concerning the most appropriate experimental design. For example, there are varying opinions on the optimum duration of a study and the number and types of species of test animals that are considered appropriate. It is probably unwise to adopt rigid "standard" test methods for all chemicals; judgment must be exercised to select an appropriate model for a particular chemical (499). Obviously, due to limitations of sample size (the number of animals on test), low incidence effects can go unrecognized. Consequently, although the results of carcinogenicity testing will, in many cases, give good indications of possible hazard, they do not eliminate the need for continuing careful observations of humans (492). In fact, most known human carcinogens, such as 2-naphthylamine and chlornaphazine, were first discovered by observing humans, and only later were they found carcinogenic in animals. The reverse was true for other compounds such as diethylstilbestrol (DES), vinyl chloride, or bis(chloromethyl) ether.

The following constitutes a series of major issues which should be carefully considered in the design of a carcinogen bioassay.

Species and Strain of Animal. Ideally, the appropriate animal species and strain is one, which, for the chemical tested, is able to predict human response accurately. However, the fact is that only three species have been routinely used in carcinogen bioassay (504). All three are rodents: the mouse, the rat and the hamster. Only these species generally meet the requirements that the animals be readily available, that the bioassays are likely to be relevant to humans, and that the bioassays be cost effective and relatively quickly performed. Included in these requirements are: the life span of the species must be of reasonable length, long enough to allow time for tumors to develop and short enough to be economically feasible (2-3 years); the animal should be small and should live and breed well in captivity; there should be similarity to humans in regard to metabolism and pathology responses; strains should be those most likely to be susceptible to cancer from the particular chemical yet not hypersensitive. Only these three species and a few strains currently meet these specifications, however, there are potentially many more inbred or hybrid strains which can provide a wide range of diversity in their metabolism of and response to carcinogens or carcinogenic precursors. There is still no scientific consensus over whether inbred or outbred strains of rodents are better for long-term studies (504-509). While the use of inbred strains may produce more precise and more stable biological re-

^{*}Although the full committee participated in the framing of this section, primary responsibility was assumed by Drs. R. Scheuplein and W. G. Flamm.

sponses, more prosaic strains may more accurately reflect human response, are less expensive, often hardier and are less prone to deterioration due to genetic drift. The NTP Ad Hoc Panel on Chemical Carcinogenesis Testing and Evaluation strongly recommended but did not insist that NTP bioassays be restricted to inbred or hybrid strains (503). When the test substance is chemically related to other human carcinogens, the strain chosen can be that used for the carcinogenic chemical analog. Extensive experience in using a particular strain is often a strong reason for continuing its use despite its acknowledged limitations (501-510). One major advantage is the availability of abundant data on spontaneous tumor rates at specific organ sites.

Because of genetic differences in tumor susceptibility, the use of only a single strain of animal rather than an array of strains represents an experimental compromise, but an essential one given the large cost of a single long-term study. The possibility of a false negative result is reduced by the current recommendation to use two species instead of one in a carcinogen bioassay. More than 90% of the time these species are the rat and mouse. The Fischer inbred (F344) rat and the B6C3F1 hybrid mouse are commonly chosen for carcinogen bioassay, in large part because of the experience gained by the NCI from their use (488,503,511). Despite its long history, the continued use of the B6C3F1 hybrid mouse by the NTP is currently under review because of the difficulty in interpreting the significance of proliferative liver lesions (488,492,493,503,512,514). The merits of using only a rat strain instead of both a rat and a mouse strain have also been recently discussed (514). This issue and others relating to species selection are considered in the NTP Ad Hoc Panel's Report (503).

Animal Care and Diet. Stringent control of environmental conditions and proper animal care techniques are mandatory for meaningful results (496). Access to animal facilities should be limited to essential personnel to prevent excessive traffic and the transmission of disease. Factors such as housing conditions, intercurrent disease, drug therapy, impurities in diet, air, water, and bedding, and animal care facilities can significantly influence the outcome of animal experiments.

The control of intercurrent infectious diseases or parasites is facilitated if rodents are bred and maintained in conditions free from specific pathogenic organisms. Animals from outside sources should not be placed on test without an adequate period of quarantine. Bedding to be used in long-term studies should be absorbent, sterilizable and free of pesticides, other toxic contaminants, or enzyme-inducing substances. Animals should be housed in quiet, well-ventilated rooms, having controlled lighting, temperature, and humidity. It is advisable to balance dose groups over rows, columns, and racks to allow for potential location effects across dosage levels. Alternatively it may be advisable to rotate cages or racks periodically to balance potential sources of variability. Eye lesions (retinal and lenticular degeneration) have been observed with animals kept in cages in closest proximity to light. Experiments should not be

initiated until animals have been allowed a one- to twoweek period of acclimatization to environmental conditions.

The absence of biases in selection and allocation of animals between control and treatment groups, as regards diet, husbandry, necropsy, and pathology is crucial (515). Since mice are reservoirs of asymptomatic viral disease, it is best to house mice and rats separately where possible. However, it is acceptable practice to house rats and mice in the same room if they are free of disease, if they come from the same supplier, and if they are being treated with the same chemical. It is preferable not to mix animals from different suppliers or from two locations of the same supplier. Housing disease-free animals in the same room reduces the chance for differences due to environmental variables. If a population of animals is diseased, then consideration must be given to aborting the study and starting over. Drug therapy should be avoided in bioassays since it is a confounding factor.

Control animals and test animals should be housed in the same room unless there is a possibility of cross-exposure. The availability of hoods, laminar air flow rooms, caging devices, and personnel practices all affect the extent to which experiments should be separated or combined. To minimize the chance for inadvertent cross-exposure of animals to the test substance, it is advisable to test only one chemical in each room. If more than one chemical is tested in a room, the actual cross-contamination of chemicals within the room must be measured. Good laboratory practice need not rule out all flexibility nor restrict the use of judgment or impose unnecessary expenses (516,517).

Cages, racks, and other equipment must be regularly and thoroughly cleaned. Periodic analysis of the basal diet should be conducted for both nutrients and unintentional contaminants, including carcinogens. The results from such analyses should be retained and included in the final report on each test substance. The essential dietary requirements of rodents are relatively high in protein and low in calories derived from fat (about 2-4% of calories). In the administration of substances by gavage using vegetable oils, the amount of fat ingested in rats can reach 30% (509). These facts, coupled with emerging data on the influence of dietary protein and fat on cancer incidence, and the presence of trace levels of carcinogens in animal diets may be important to future re-evaluation of bioassay results. Furthermore, the quality of the agricultural crops that go into animal feed can vary seasonally. Varying amounts of naturally occurring enzyme inducers from plant material in animal feeds may affect the course of a long-term test for carcinogenicity. It is therefore important to document and retain pertinent information on diet source and quality.

When the test substance is administered in long-term studies, stability tests are essential. Properly conducted stability and homogeneity tests, prior to the chronic study, should be used to establish the frequency of diet preparation and monitoring required. Similar monitoring is required of the atmosphere within a chamber in

inhalation tests to determine the necessity of rotating animals within the exposure chamber and the frequency of monitoring required. The stability of the neat chemical, if it is to be used for a long time, as in long-term skin-painting studies, should also be determined.

If diets are sterilized, the effects of such procedures on the test substance and dietary constituents should be known. Appropriate adjustments to nutrient levels should be made.

During carcinogenicity tests, investigators should be aware of potential chemical contaminants in the water used. Although water approved for human consumption is generally satisfactory, the investigator should determine the levels of potential contaminants that are likely to influence the outcome of the study. The analyses of the water supply should be included in the reports of these studies.

Test and Control Groups. The number of animals to use has been the subject of great concern because of the fact that the test animals may be serving as surrogates for more than 200 million people in the U.S., and potentially for far more worldwide. While it is desirable to enhance the sensitivity of the test, it is not feasible to use very many animals. Furthermore, a moderate increase in the group size provides relatively little increase in the statistical power of the test. Therefore, careful attention must be paid to experimental design to assure the maximum reliability of the study and that the results are amenable to statistical evaluation. The use of adequate randomization procedures for the proper allocation of animals to test and control groups is of particular importance. A sufficient number of animals should be used so that at the end of the study enough animals from each group are available for thorough pathological evaluation. For such reasons, it has been recommended that each dose group and concurrent control group should contain at least 50 animals of each sex (488,498,499,503). If interim sacrifice(s) is included in the study plan, the initial number should be increased by the number of animals scheduled for the interim sacrifice(s).

At least one concurrent untreated control group, identical in every respect to the exposed groups, except for exposure to the test substance, should be used. In certain studies, such as with inhalation exposures that require unique housing conditions, it may also be appropriate to include an additional concurrent control group housed under conventional conditions.

Occasionally, a positive concurrent control group is useful, particularly when there is reason to believe that the strain of animals may be resistant to the particular class of carcinogens or that a previously sensitive strain has become insensitive because of genetic drift. A positive control can also provide some information on the relative carcinogenic potency of the tested chemical. In routine carcinogenicity testing, no positive controls need be used (488,498).

Dose Levels, Frequency, and Route of Exposure. If a single bioassy is to be used both for detection and for risk assessment, at least three dose levels in addition to the concurrent control group are recommended. The question of what maximum dose should be administered may be the most controversial issue concerning bioassays (497,503,517).

The highest dose currently recommended is that which, when given for the duration of the chronic study, is just high enough to elicit signs of minimal toxicity without significantly altering the animal's normal lifespan due to effects other than carcinogenicity (503). This dose, sometimes called the maximum tolerated dose (MTD), is determined in a subchronic study (usually of 90 days duration) primarily on the basis of mortality, toxicity, and pathology criteria. The MTD should not produce morphologic evidence of toxicity of a severity that would interfere with the interpretation of the study. Nor should it comprise so large a fraction of the animal's diet that the nutritional composition of the diet is altered, leading to nutritional imbalance.

The MTD was initially based on a weight gain decrement observed in the subchronic study; i.e., the highest dose that caused no more than a 10% weight gain decrement (494). More recent studies and the evaluation of many more bioassays indicate refinement of MTD selection on the basis of a broader range of biological information (503). Alterations in body and organ weight and clinically significant changes in hematologic, urinary, and clinical chemistry measurements can be useful in conjunction with the usually more definitive toxic, pathologic or histopathologic endpoints. The 1984 report of the NTP Ad Hoc Panel (503) contains a helpful discussion of the use of such corroborative information to more reliably determine the MTD (see pages 126-145 of the NTP Report). The rationale for the dose selection, including the data elements rationale for the selection of the dose based on these data.

The Report of the NTP Ad Hoc Panel also discusses the kind of data which may be helpful in selecting the spacing of doses below the MTD. Such doses are ordinarily simple fractions of the MTD; e.g., 1/2, 1/3, etc. Dose selection is influenced primarily by one or more of these types of information: observations in the subchronic studies, particularly the dose-response curve; nonlinear behavior and the doses at which this occurs; and anticipated human exposure levels or relevant exposure guidelines (such as the threshold limit values [TLVS]).

The important contribution of pharmacokinetic studies to the selection of doses below the MTD is the information such studies may provide on the presence of dose-dependent and the time-dependent processes affecting the disposition of substances in the body (503,519-525). Such information may also identify changes in the internal concentrations of the chemical or its metabolites that are not directly proportional to administered dose and may also indicate if accumulation of the chemical or its metabolites occurs with repeated exposures. To be of maximum use in the dose selection process, this information should be obtained early, before or during the prechronic phases of the bioassay. However, because pharmacokinetic data may also re-

veal distinct changes in the animals' treatment of the chemical at low as compared to high doses, such data may also aid in the interpretation of the bioassay.

Because of cost, a single long-term study may be utilized to obtain data on chronic toxicity, on carcinogenic potential and for carcinogenic risk assessment. The present discussion envisions the combination of both the qualitative objective of a carcinogen bioassay, the detection of carcinogenic potential, and the quantitative objective, obtaining dose-response data for risk assessment, into a single bioassay. If only one of these objectives is sought, it may be advisable to select the doses differently. The purpose of using the MTD is to provide maximum opportunity for the detection of a neoplastic response. The purpose of several lower doses is to provide adequate information on the shape of the dose-response curve for risk assessment and to ensure a meaningful comparison with the controls even if the MTD was unfortunately selected at too high a level. The pathological responses observed in the subchronic study and the slope of the subchronic dose-response curve should aid in the selection of the range and number of doses for the long-term study. For example, a steep dose-response curve would suggest closer spacing of intermediate doses than a shallow dose-response curve. The existence of metabolic saturation (as indicated by pharmacokinetic data) at doses well below those identified by pathological criteria suggest a spacing of doses designed to determine both whether the highest dose has any carcinogenic effect and to adequately define the dose-response curve for the saturable process. This may require additional doses. Generally four doses including the control, (occasionally three) will be necessary to adequately define the dose-response curve, assuming an appropriately chosen MTD. Since the MTD is an estimate at best, the value of additional doses is generally difficult to judge. If the estimated MTD eventually produces moderately high or excessively high tumor incidence, additional lower doses may aid in defining the shape of the dose-response curve and may even salvage a study when the MTD is overestimated. If the estimated MTD produces no tumors, or a low incidence of tumors, the additional doses will have been unnecessary as regards carcinogenicity but can help establish safe nontoxic doses. As indicated above, the best guide to the number of doses is the acquisition of knowledge on the pathological and pharmacokinetic criteria upon which a sound, scientific judgment can be based. The National Toxicology Program (NTP) has published more detailed recommendations for prechronic testing and dose selection from its Ad Hoc Panel on Chemical Carcinogenesis Testing and Evaluation (503). Interruptions in the administered dose may permit some cellular repair and reduce the sensitivity of the test. Therefore, if the chemical is administered in the drinking water or mixed in the diet, it should be continuously available. The frequency of exposure may vary according to the route chosen or may be adjusted according to the toxicokinetic profile or stability of the test substance. Protocols for inhalation or for dermal exposure are usually subject to special constraints (498).

The route of administration used for the bioassay is critical for both its qualitative and quantitative objectives—but in different ways. If the route of administration results in the absorption, distribution and metabolic activation (if required) of the substance, it is reasonable to regard the test as relevant for a qualitative demonstration of human carcinogenic hazard. This comparison is obviously most directly made when the route used in the test and the route of human exposure are the same, but as long as the criteria above are satisfied, the results of animal studies are generally considered qualitatively meaningful.

If, because of its palatability, solubility, stability, or volatility, a substance can not be given in feed or water, oral gavage may have to be considered. This is clearly less desirable, however, since single large boluses can result in absorption and pharmacokinetic patterns different from those anticipated from human exposures. Sometimes, the use of gavage can be avoided by gradually increasing dietary concentration of the test substance to gain acceptance, by altering other dietary components to increase the palatability or by microencapsulation.

Certain site specific tumors, e.g., bladder carcinoma following bladder implantation and subcutaneous sarcomas following injection, may not be relevant for human oral exposure. The presence of tumors remote from the site of administration and a knowledge of site-specific metabolism play a significant role in the final evaluation. Some routes of administration may fail to provide adequate metabolic activation or exposure of target tissues and therefore lead to false negative results (498). Such data require careful evaluation.

When an assessment of human risk is desired, it is desirable to obtain sufficient data to permit a reasonably accurate estimate of the dose of the carcinogen reaching the target tissue. In some instances, e.g., for substances used on the skin, this may require a bioassay to detect tumorigenic potential and a separate study of the substance's permeability across the skin to measure the amount absorbed. In inhalation studies, this may require a separate study for determining the amount of inhaled material deposited on, or absorbed by the respiratory tract.

Duration of Study. Test animals should be exposed for the majority of their normal life spans. While there are certain animal test systems which offer the possibility of inducing cancer within only a few weeks or months, such systems at the present time are useful mainly as research tools to explore specific, well-defined questions. They cannot be relied upon to test unknown substances for carcinogenicity and are insufficient as a primary basis for regulation.

Treatment is preferably begun as soon after weaning as possible and continued for the major portion of the animals' lifespans. This is at least 18 months for mice and hamsters and 24 months for rats. In some instances where the cumulative mortality at the planned time of sacrifice is still very low, e.g., less than 10%, a longer

duration may be appropriate. Some protocols call for treatment soon after birth (neonatal) or during fetal development (*in utero*). The rationale is based on the greater susceptibility of certain organ systems to carcinogens during their early development. However, there are logistical problems which preclude using neonatal or *in utero* systems for routine bioassays.

Termination of the study is ordinarily acceptable when the number of survivors in the low dose or control group is reduced to 20–25%. Generally, because of difficulties produced by naturally occurring tumors, longterm studies are conventionally terminated by week 100 for hamsters, 120 for mice, and by week 130 for rats. regardless of mortality. When there is an apparent sex difference in response, each sex should be considered a separate study (496). If the high-dose group dies prematurely from obvious toxicity, this should not trigger termination of the lower-dose groups or controls. A negative test is ordinarily accepted by regulatory agencies if no more than 10% of any group is lost due to autolysis, cannibalism or management problems and if survival of all groups (per sex per dose) is no less than 50% at 80 weeks for hamsters, 96 for mice, and at 104 weeks for rats (499).

Rigid rules for termination are not helpful, the best guide is professional judgment, experience and careful observation of the condition of the animals. The longer the test continues, the greater the opportunity for the development of tumors (526,527). On the other hand, the sensitivity of bioassays decreases with time, because of the natural appearance of age-related tumors in the controls. In theory, it is necessary to know both the incidence and growth rate of naturally occurring tumors and experimentally induced tumors at various organ sites in the sex and strain of animal used in order to determine the optimal time for termination. This information is ordinarily not available until the end of the test and while serial sacrifices are useful, they are considered too expensive for routine testing of chemicals.

Data Collection and Reporting. Animals should be checked carefully at least twice each day. Observations should be sufficient to detect onset and progression of all toxic effects, as well as to minimize loss due to diseases, autolysis, or cannibalism. Body weights should be recorded individually for all animals once a week during the first 13 weeks of the test period and at least once every 4 weeks thereafter. Loss of body weight should trigger careful clinical examination of the animal and its sacrifice, if necessary. Food intake should be determined weekly during the first 13 weeks of the study and then at approximately monthly intervals, unless health status or body weight changes dictate otherwise (499).

Clinical signs and mortality should be recorded for all animals. Special attention must be paid to clearly visible tumor development; the time of onset, location, dimensions, appearance, and progression of each grossly visible or palpable tumor should be recorded.

The pathological examination, macroscopic as well as microscopic, is the cornerstone of the carcinogenicity

study. A thorough gross necropsy with subsequent selection of appropriate lesions for histopathology is mandatory. It is vital that the sampling among the test and control tissues be uniform so that no bias is introduced into the comparisons. Microscopic examination of both test and control tissues is required. While an all-inclusive microscopic examination of all tissues is theoretically desirable, the resource limitations may dictate a more selective approach such as the one recently proposed by the National Toxicology Program or earlier by the National Cancer Institute (528,529). The quality, extent and completeness of the documentary record of the bioassay are of critical importance in establishing its validity for scientific and regulatory purposes. It is crucial that the physical evidence, e.g., the original tissue sections and slides, is accurately prepared and maintained. Sponsors of bioassays that are to be submitted to regulatory agencies should be familiar with additional requirements of GLPs (Good Laboratory Practice) (516).

Interpretation and Evaluation of Long-Term Carcinogen Bioassays

BACKGROUND. Several attempts have been made over the last 10 years to construct a suitable operational definition for a chemical carcinogen (493,494,497,498,530,531). Though none of these have succeeded fully, there is common agreement that a chemical carcinogen is a substance which induces cancer by some chemical/biological mechanism. A chemical carcinogen may be a substance which either significantly increases the incidence of cancer in animals or humans or significantly decreases the time it takes a naturally-occurring (spontaneous) tumor to develop relative to an appropriate background or control group. Either phenomenon is said to represent the effects of a carcinogen (467).

The interpretation of carcinogenicity data has evolved considerably in the last decade. In past years, there were no standardized protocols for testing carcinogens. Typically an identified carcinogen was used as a research tool to study the mechanisms of carcinogenesis, so that many experiments were performed with the compound. This intense study usually dispelled any doubt as to whether the substance was actually a carcinogen. This degree of confirmation gradually decreased as more of the experimentation turned from research on the process of carcinogenesis to detecting as many chemicals as possible that might pose a cancer risk to humans. For this latter purpose, which began in earnest in the mid 1960s with the "screening" bioassavs conducted by NCI, it was necessary to devise "decision rules" by which to declare a chemical a carcinogen (487). Since then, considerable progress has been made in refining basic principles and criteria to guide the interpretation of carcinogenicity (488,489,498,499). Experience has taught that these criteria should be regarded as general guidelines in evaluating rather than rigid rules and that final decisions are best made after careful communication between pathologists, toxicologists and statisticians. Most useful to the process is the experience and the data derived from large numbers of recent tests using a uniform protocol.

With some oversimplification, the task of the pathologist and toxicologist is to decide whether a lesion is cancerous or precancerous and whether it is related to exposure to the test chemical. The statistician, on the other hand, has the task of determining whether any observed tumors are more likely to have occurred by chance or as a result of treatment. Neither of these tasks is simple or completely independent of the other and, more importantly, each requires experience and judgment if sensible decisions are to be made.

Toxicological Issues in Bioassay Evalua-TION. Largest doses (i.e., doses far exceeding human exposure levels) have been strongly recommended by several national and international bodies in order to overcome the inherent low sensitivity of bioassays (498,499,503). These recommendations have been controversial because high doses may themselves produce altered physiologic conditions which can qualitatively affect the induction of malignant tumors. Normal physiology, homeostasis, and detoxification or repair mechanisms may be overwhelmed, and cancer, which otherwise might not have occurred, is induced or promoted. Normal metabolic activation of carcinogens may possibly also be altered and carcinogenic potential reduced as a consequence. If qualitatively different distribution, detoxification, or elimination of the test chemical is produced at the highest dose, a toxic response at this dose may not be indicative of effects at low exposure levels. The cancers which arise from high doses of corticosteroids, estrogens, certain sulfonamide compounds and from some instances of bladder implantation may result from such "secondary" effects (53 \hat{z}). This concern is not easily resolved, for if the dose is not high enough, carcinogens may remain undetected by traditional bioassays.

Use of the Maximum Tolerated Dose. The concept and operational definition of the MTD have undergone modification during the last several years in response to some of these concerns. The 1971 FDA Advisory Committee on Protocols recommended that: "Testing should be done at doses and under experimental conditions likely to yield maximum tumor incidence" (493). However, as the use of these tests for human risk assessment increased, it became vital not only to detect carcinogens qualitatively (i.e., avoid false negatives), but also to estimate effects at low dose levels. Both the National Cancer Advisory Board Subcommittee (NCAB) (531) and the Interagency Regulatory Liaison Group (IRLG) (498) cautioned against the use of a dose so high that it produced "unwanted toxic side effects" (IRLG) or "unphysiologic conditions [which] may in themselves enhance tumor formation" (NCAB). Recent publications have emphasized the need to select the doses not only on the basis of the results of subchronic toxicity studies, but also on the basis of what is known about the pharmacokinetics and metabolism of the test substance (497,503,517). Better techniques to observe and detect significant differences in metabolism and pharmacokinetics throughout the dose range used are

being developed; these should help in a better determination of the MTD (519-525). The desirable goal is that the dose used meets the objectives of maximally enhancing the sensitivity of the test without introducing qualitative distortions in the results.

Promoters vs. Initiators, Current Experimental Status. In the portion of this report titled "Current Views on the Mechanism of Carcinogenesis," the evidence for the belief that cancer occurs through at least a two stage process consisting of initiation and promotion was discussed. The basic question of whether tumor promotion is a general feature of carcinogenesis or is only characteristic of some organ systems is still debatable, although more and more evidence for the initiation-promotion process in a variety of organs is becoming available (533). The question naturally arises as to whether such promoting agents can reliably be distinguished from initiators by traditional bioassays.

A number of examples of promoting agents are well documented; most of the work on their mechanism has been done on skin, e.g., the phorbol esters in epidermal carcinogenesis. Other examples are butylated hydroxvanisole in pulmonary carcinogenesis, phenobarbital in hepatocarcinogenesis and estrogenic hormones in breast cancer. Some known promoting agents also have weak tumorigenic activity and some are also initiators, i.e. complete carcinogens. Carcinogens may act as promoters at some tissue sites and initiators at others. It is very difficult both in principle and in practice to confirm the claim that a given chemical acts by promotion alone. If such a mechanism is postulated for a specific chemical, data on the purported mechanism at the affected tissue sites and dose response data would be needed to support the proposition.

Need for Biological Significance, Other Data. is no simple or generic solution to the problems which arise when attempting to interpret carcinogenicity data. Over recent years, review groups charged with the responsibility to evaluate carcinogenicity data and come to some decision regarding the carcinogenicity of a chemical have recognized the need to consider all of the available scientific information which may bear on such a decision (492,493,497,531). In addition to statistical significance (see Evaluation of Statistical Significance), there should be biological significance as well. The use of a variety of biological information on dose response, tumor progression, tumor latency, tumor multiplicity, findings in other studies, etc. can add confidence to the final assessment. Other often useful information includes historical data on control animals, information from in vitro mutagenicity or neoplastic transformation systems, toxicological information which might indicate that the organ or tissue which is at risk for carcinogenicity appears to be a target for the toxicity of the compound. Mutagenicity, DNA repair, and neoplastic transformation studies can sometimes be used in aiding judgments on carcinogens (484). For example, if the data indicating carcinogenicity are of only marginal significance, negative findings in properly designed and well-conducted in vitro tests, depending upon their relevance to the specific situation, may lead to a conclusion that the chemical is probably not carcinogenic. On the other hand, if the results from *in vitro* testing were clearly positive, and if evidence existed to show that the organ in which the cancer question arose was also the toxicological target of the chemical, the conclusion would be likely to be that the chemical is carcinogenic. To make these judgments, it is important that there be a good interaction and deliberation among scientists from different disciplines.

Human Relevance of Rodent Tumors. The inference stated earlier, "that in the absence of adequate data on humans it is reasonable to regard chemicals that are carcinogenic in laboratory animals as if they were also carcinogenic to humans," should be given great deference as a general principle. But this need not foreclose further scientific inquiry in individual cases if experimental evidence on the mechanism of tumor formation in the animal is available. As science advances and more sophisticated studies are done, it may become possible to demonstrate the existence of unique susceptibilities to chemical carcinogenesis in rodents that have no relevance for humans. All the available scientific information should be used in attempting to reach the most likely correct scientific conclusion. The inferential relevance of observed animal carcinogenesis to potential carcinogenic risk in humans is obviously justifiable in the absence of such mechanistic information.

Issues of Pathology in the Interpretation of TUMOR DATA. As mentioned earlier, a carcinogen may be defined as a substance which increases the incidence of cancer above the background rate. However, from a practical point of view, the most important question is: how does one estimate or deduce cancer incidence from tumor data? For instance, should all cancers in the animal be combined and the total number of tumors in treatment groups compared to total cancers in the untreated controls? Alternatively, should carcinomas be separated from sarcomas since they are derived from different tissue? Should the comparisons between treated and controls be restricted to specific organ sites and, if so, should all tumors be combined, or only certain types? Other questions are: should benign and malignant tumors be separated or combined; if tumors occur at multiple organ sites, can they be combined and how should it be done; should cancers which shorten the life of the animal be separated from those which do not? These questions and many more must be answered before cancer incidence(s) can be estimated. Unfortunately, there is no universal agreement among experts in the field relevant to the rules for calculating cancer incidence that is applicable to "all" carcinogens "all" of the time.

Tumor Number, Type, and Site in the Assessment of Incidence. Generally, most experts agree that the incidence of total tumors at all organ sites is not a very useful expression of cancer incidence, nor is the calculation of the incidence of total benign or total malignant tumors. Most useful appears to be the number of histologically unique tumors at specific organ sites (534).

The grouping of lesions for evaluation should be based on commonality of histogenetic origin. Therefore, sarcomas should not be combined with carcinomas for the purpose of establishing a value for the incidence of cancer at a specific organ site. It is often necessary, at least in the judgment of many pathologists, to combine certain benign tumors with malignant ones occurring in the same tissue and at the same organ site. This practice can make the total neoplastic response (benign and malignant) clearly significant, despite the lack of statistical significance in the tumors diagnosed as malignant. These pathologists believe that truly benign tumors in rodents are rare and that most tumors diagnosed as benign really represent a stage in the progression to malignancy. For some tissue sites, this view is widely accepted. Examples of this are adenomas versus adenocarcinomas in the pituitary, thyroid, lung, kidney tubules, and according to some experts, in mouse liver. In each of these cases, it is argued that the judgment of the pathologist as to whether the lesion is an adenoma or an adenocarcinoma is so subjective that it is essential they be combined for statistical purposes. It is also argued, in these specific cases, that the adenoma is a precursor of the adenocarcinoma. Indeed, the Subcommittee on Environmental Carcinogenesis of the National Cancer Advisory Board recommended in 1976 that these lesions be combined for statistical purposes (531). Most U.S. regulatory agencies have followed this recommendation. Ordinarily the tumor or cancer incidence is calculated as the number of tumor-bearing animals having tumors at a specific organ site divided by the total number of animals that survived long enough on experiment to have been at risk for that specific type of tumor. For reasons discussed in the statistical part of this section, life-shortening tumors should not be combined with nonlife-shortening tumors.

Time to Tumor. A positive result in a carcinogenesis bioassay can be based on evidence of a substantially decreased latency period, as well as on the induction of an increased incidence of malignant tumors. A good discussion of the precautions that should be observed in order to establish decreased latency is given in the 1979 IRLG report (498). Such data, in conjunction with incidence data, can be useful to distinguish between the relative potencies of carcinogens.

Problems in Tumor Diagnosis. A major problem facing those who must make either scientific or public health policy decisions based on carcinogenicity data is that the diagnosis of pathological lesions is subjective and pathologists may disagree on diagnosis. Frequently, the disagreement is not as serious or far-ranging as it may appear to nonpathologists. The disagreement may be focused on an issue where neither pathologist is absolutely certain and each is forced to offer his "best opinion." In practice, for most compounds, NCI and NTP have used panels composed of two to four pathologists to review and analyze the lesions that have been observed (528,534). This procedure helps to provide consistency in diagnosis, but it may not solve all the problems. For instance, most pathol-

ogists do not review the microslides "blind." They know which slides come from control animals and which from treated animals. The extent to which this information leads to bias in diagnosis is not known (528).

A common problem with the interpretation of pathological data concerns the selection of the tissues at necropsy, how the tissues were prepared, the slides mounted and so on. For instance, a few years ago most investigators did not open the skulls and carefully examine the brain. In other cases, only brains appearing abnormal were examined in detail. Currently, the brain as well as other tissues such as nasal turbinates are routinely examined histologically (526,534), though the techniques vary somewhat. As a consequence, spontaneous tumors of these tissues are now being reported. Depending upon how extensively these tissues are being sectioned for histological analysis the observed incidence may vary from low to moderately high. Such modifications in the pathology protocol, if not recognized and understood, could lead to serious errors in judgment concerning the carcinogenicity of a chemical.

Critical to decisions about carcinogens is the biological significance and human relevance of certain types of tumors, particularly the liver tumors in the mouse. This matter has been the subject of heated debate for the past 15 years. Some scientists are certain that the mouse liver is overly sensitive and will respond to almost any toxic insult by developing cancer. Other scientists are just as certain that the mouse liver is a reliable indicator of carcinogenicity and as good a predictor of potential human risk as any indicator available. In one review, 61 of 85 chemicals (73%) that increased the incidence of liver tumors in mice also induced tumors in other tissues of mice and/or in rats (511). Some toxicologists believe that part of the problem stems from work on C3H strains of mice where the background liver tumor incidence is high and false positive errors are possible. Generally, there is consensus that the mouse liver model in principle does have significance in terms of human risk, but unambiguous diagnosis between "benign" or "hyperplastic" liver nodules and malignant neoplasia remains elusive and there is no apparent scientific consensus. A recent review of this issue has emphasized the need for judgment on a case-by-case basis and the possible relevance of other toxicological information (535).

In addition to liver tumors, other tumors, including lymphomas in the mouse, which appear to be of viral origin, have been questioned as to their relevance to human cancer. As a general matter, toxicologists must question the relevance to humans of any experimental tumor which has a known viral background where the virus may be unique to the animal. This includes a variety of lymphomas and sarcomas in rats and mice, as well as mammary tumors in the mouse. Likewise, the relevance to human exposures of the induction of bladder cancer in rodents which are shown to have bladder stones and crystaluria only at high doses of the test substance is questionable.

EVALUATION OF STATISTICAL SIGNIFICANCE. For

the analysis of tumor incidence data, a widely-used method for comparing experimental and control groups is Fisher's exact test (536). This analysis is generally carried out in conjunction with a Cochran-Armitage trend test (537,538) which examines the relationship between tumor incidence and dose. The trend test has the advantage of utilizing cancer incidence data for all doses and as a result is somewhat more sensitive than pairwise comparisons to the detection of carcinogenic effects.

The statistical procedures mentioned above for the analysis of tumor incidence data do not take survival differences into account (515). This can be a problem with chemicals that produce a life-shortening effect. Although "survival-adjusted" tumor incidence analyses (515,539,540) are more complex, they have the advantage of adjusting for intercurrent mortality and of being sensitive to shortened latency period as well as to increased tumor incidence. These procedures require the determination of whether tumors in individual animals are "lethal" or "incidental" with respect to the underlying cause of death. This determination may be difficult if not impossible to make for many animals, but experimentalists should try to distinguish in their records between lethal and incidental tumors.

While it is customary practice in the conduct of bioassays to regard an increased tumor incidence that is statistically significant (e.g., p < 0.05) as reasonable assurance that the effect is real and not due to chance alone, the final judgment regarding a compound's carcinogenicity is rarely based on a single finding of statistical significance (540). One must be assured that the effects observed are biologically significant as well. Other factors that must be taken into account include: the reproducibility of the effect in other doses and/or in other sexes or species; the historical incidence of the tumor in question; the survival histories of dosed and control animals; and evidence of hyperplasia, metaplasia or other signs of an ongoing carcinogenic process.

The 0.05 p-value is a reasonable statistical bench mark with a history of use in the bioassay field. It would be imprudent to disregard it as an index of significance without good reason. It would be equally unwise to regard it as an absolute requirement or a sine qua non without considering the weight of biological evidence. For example, doubling or tripling the incidence of a tumor that occurs at low incidence in the controls may not be statistically significant, but may indicate a biological effect that should not be dismissed. Similarly, the finding of nominal statistical significance (p = 0.05)at a single tumor site may be less important to the final evaluation than knowledge of the target site and the lack of consistency with related biological findings. As stated by IARC (499): "The p-value from a good experiment is an objective fact, subject to public agreement. The weight to be given it and to the available pieces of information is not."

Problem of False Positives and False Negatives. Because of the time and expense involved, most chemicals, if tested at all, are tested in only one bioas-

say. It is vital that this test have good sensitivity, since otherwise many true carcinogens would remain undetected (a false negative). The test must also possess a high degree of specificity, since there are economic losses as well as disincentives for testing if safe, useful substances are mistakenly declared carcinogens (a false positive).

The typical bioassay, due to inherent limitations (low number of animals and biological variation) cannot reliably detect an increased tumor incidence of much less than 10–15% over background. High test doses are one way around this limitation, but as discussed above this can introduce confounding factors if the dose is too high. In addition, the possibility always exists that for a specific substance, the most sensitive species may be inappropriate for qualitative extrapolation to humans. Until more is known concerning the mechanism of cancer and the similarity in response between man and animal, this uncertainty will persist.

Confidence in an apparently negative bioassay increases with the number of animals used, the adequacy of the doses, the extent to which its duration approaches the animals' expected lifetime, and the adequacy of the gross and histological examination of both control and treated groups. Corroborative evidence such as experimentally consistent biological results in different species or sexes, or at different doses, strengthens the likelihood that the bioassay is correctly interpreted. Rigid rules or definitions are not very useful. The ultimate decision must be based on the knowledge of experienced pathologists and toxicologists, the weight of corroborative evidence and a careful statistical evaluation.

The spontaneous tumor incidence can be of considerable importance in the interpretation of results from carcinogenicity studies. If, for example, the effect of a chemical is to double or triple the background tumor incidence, tissue sites with low spontaneous tumor rates are more likely to yield false-negative results than are sites with high spontaneous tumor rates. For example, if a tumor is a rare tumor, even a slight increase in incidence may be biologically significant and may be considered adequate evidence of carcinogenicity (536). This factor must be taken into account in the overall evaluation of the data. For such tumors (e.g., with spontaneous tumor rates of 1% or less), the utilization of historical control data may be particularly useful in increasing the sensitivity of the study for detecting carcinogenic effects. All recent work emphasizes the fact that the evaluation of the bioassay is not a simple nor routine statistical exercise, but a multidisciplinary process requiring the interaction of toxicologist, pathologist and statistician.

Appropriate precautions in the design and evaluation of bioassays can minimize the chance of false positive results. The test substance must be free of carcinogenic contaminants and appropriate doses and routes of exposure must be used. However, careful attention to these and to other factors cannot eliminate entirely the possibility of either false positive or false negative re-

sults. These problems are fundamental to the process of drawing inferences from observed sample data.

Many investigators familiar with data from bioassays believe that in practice the overall false-positive rate does not greatly exceed 0.05 (540,541). Others are not as certain, and all admit that this is an important problem area that must be considered in the interpretation of carcinogenesis bioassay data. The very effort to be as thorough and as complete as possible in the selection of tissue at necropsy, and in the detection of responses from various doses, increases the chance for a false positive error. Typical bioassay protocols require sectioning and analysis of tissues from 20-30 sites in two species, in two sexes, and, at the least, two dosage levels. It has been suggested that the multiple tests for significance applied to these many sites produce an unacceptably high overall false positive rate. The argument is that, if n independent tests are conducted at the asignificance level, the probability of finding a significance somewhere (the false positive rate) is [1 - (1 - $(a)^n$]. For n = 20 and p = 0.05 for a single dose-sex comparison, this probability is about 0.64.

However, recent investigators have shown that this argument overestimates the likely false-positive rate of the bioassay. Gart et al. (536), using historical incidence rates for B6C3F1 mice, found the probability of at least one significant result at the 0.05 level at some site in at least one sex was only about 8%. In a commonly used strain of rat (F344) the same probability was approximately 21%. These findings are well below the 64% value cited above. One reason for this difference is that tumor incidences are discrete count data, and in this situation, the actual significance level of test procedures comparing tumor rates is always less than the nominal level, often considerably less.

This is particularly true for tumors having a low spontaneous incidence. Even the estimated false-positive rates reported by Gart et al. (536) may be too high, since (as noted earlier) the final judgment that a compound is carcinogenic is rarely based on a single isolated "significant" increase in tumor incidence. Generally more stringent evidence is required, such as the reproducibility of the effect in other doses and/or other sexes or species. For example, if a chemical is judged carcinogenic only if it is positive (p < 0.05) at some tissue site for both the low and high doses, the overall false positive rate for a two-sex, two-species experiment has been estimated to be 5.1% (542).

In a recent examination of 25 NTP feeding studies, a simple statistical rule was derived by Haseman (541) which appears to mimic closely the scientific judgment process used in these experiments. This "rule" was as follows: regard as carcinogenic any chemical that produces a high-dose increase in a common tumor that is statistically significant at the 0.01 level or a high-dose increase in an uncommon tumor that is statistically significant at the 0.05 level. The overall false positive rate associated with this particular decision rule (which appears to approximate closely the overall evaluation process) was estimated by Haseman and found to be no

more than 7-8% for the current NTP two-sex, two-species protocol (543).

In summary, in the interpretation of tumor incidence data an investigator should be aware of the false positive issue. In the overall evaluation process, one should be guided by (but not rely exclusively upon) the statistical significance of an observed tumor increase. This awareness will help minimize the likelihood of a false positive result.

Use of Historical Controls. It is extremely important for the interpretation of a bioassay that the variability of the incidence of spontaneous tumors in the strain of animals used be established with the highest precision possible. These variations present problems in interpreting the results of bioassays. NTP has recognized this problem and has continued to accumulate data from untreated and corn-oil gavage control groups for the strains of animals used (509,510). With such data, the toxicologist may conclude that a study yielding statistical significance is not biologically significant or conversely that an effect is real, even though there is no "significant" difference between test and concurrent controls. Of 27 bioassay study reports issued by the NCI from 1978 to 1980, historical controls were used in 14 studies to show that an apparent increase of a specific tumor was not related to treatment (543). In 13 studies, the use of historical controls showed that an increase in tumors was related to treatment. Historical control data can be valuable when used appropriately, especially when the differences in incidence rates between treated and concurrent negative controls are small and can be shown to be within the anticipated historical incidence. However, the use of historical control data is not without its own problems. Over the course of time, the thoroughness of pathologic evaluations has improved and pathologic diagnosis has evolved and nomenclature has changed. There is also the possibility that the genetic susceptibility of the strain has shifted. The sources of variability in historical control data should be identified and, if possible, controlled. Obviously one has more confidence in the most recent historical control data from the same laboratory conducting the current study than in a compilation of pooled older data from other laboratories. Several procedures have recently been proposed for utilizing historical control data in a formal testing framework (543-546). The relative merits of these methodologies are currently being investigated by the NTP.

Summary

Although the identification of carcinogenic substances has long been of interest to the scientific community, until the 1960's the focus of most studies was research, not testing for substances which may pose a public hazard. The National Cancer Institute, and later the National Toxicology Program, have conducted a number of test-oriented studies, the chronic bioassays, and have also attempted to standardized many important parameters in conducting these studies. Type and number of animals, dose level and route, and study duration are fairly uniform in an effort to make the data base as

comparable as possible from test to test.

Certain issues remain as problems. One of these is the use of the maximum tolerated dose (MTD) in long-term studies and the appropriateness of dose route, schedule, and length of exposure. Use of the MTD, if not an ideal solution to the problem of bioassay sensitivity, is appropriate if it is properly determined. Another problem is the interpretation of tumor data. This includes tumor number, type, time to tumor, tumor diagnosis, etc. and the significance of certain tumors. General guidelines have been recommended, with the proviso that they are to be used in conjunction with sound judgment.

Furthermore, although several methods of statistical analyses of long-term test results have been developed, the questions of false negatives and false positives and the use of historical controls, suggest that the evaluation of a long-term test is not a routine statistical exercise, but is instead a multidisciplinary process requiring the interaction of toxicologist, pathologist, and statistician. The utilization of long-term animal studies for quantitative human risk assessment has accentuated and focussed attention on the real scientific uncertainties involves in high-to-low dose and animal-to-human cancer risk extrapolations. Nonetheless such animal studies, with all their limitations, remain the best way of predicting effects in humans when epidemiological data are unavailable.

Current Views on Epidemiological Methods*

Introduction

Epidemiology is generally defined as the study of the distribution and determinants of disease in human populations, and thus it involves two major approaches (547). Descriptive studies examine the "distribution" of disease and are usually employed to generate etiologic hypotheses, while analytical studies are used mainly to test hypotheses and identify the "determinants" of disease. A primary objective of epidemiology is to identify and quantify relationships between exposure to environmental agents and deleterious health effects. These associations may lead to causal inferences (548), which in turn provide the basis for instituting preventive measures for various diseases.

Strengths and Limitations of Epidemiology

STRENGTHS. In contrast to studies in other biological systems, epidemiology directly evaluates the experience of human populations and their response (risk of disease) to various environmental exposures and host factors. Thus, it is often possible to evaluate the consequences of an environmental exposure in the precise manner in which it occurs and will continue to occur in human populations. This includes such important considerations as dose, route of exposure, and concomitant

^{*}Although the full committee participated in the framing of this section, primary responsibility was assumed by Dr. R. Adamson, with Drs. J. P. Fraumeni and R. Hoover of the National Cancer Institute.

exposures to other exogenous and endogenous factors. Through epidemiological studies human cancer has been linked to a number of lifestyle and other environmental hazards, including tobacco products and alcohol, ultraviolet and ionizing radiation, certain occupational and medicinal chemicals, dietary factors, and some infectious agents (549-551).

Epidemiology has played a central role in detecting carcinogenic exposures, and it has complemented studies in laboratory animals in clarifying the carcinogenic potential of specific agents (552). Another strength of the epidemiologic approach is its ability to provide insights into the mechanisms of human carcinogenesis. Thus, epidemiological observations have complemented experimental evidence that carcinogenesis is a multistage process, and that many cancers may result from the cumulative effect of environmental and host factors that accelerate or retard the transition rates at various stages of carcinogenesis (553).

LIMITATIONS. Although epidemiology is the only means of assessing directly the carcinogenic risks of environmental agents in humans, the method has several limitations that are difficult to overcome (547,554). One problem is that evidence of an environmental hazard is usually obtained from persons with high or intermediate levels of exposure. Just as for studies in laboratory animals, detecting causal relationships at low exposure levels is difficult, since the observed associations with disease are usually less pronounced and may have alternative explanations, including those related to chance, errors, biases or confounding variables. To provide a valid basis for risk estimates, large numbers of human subjects are often needed, especially if the exposure is low or rare, or if the excess risk is small compared to that of the baseline incidence rate. Another obstacle to epidemiology is the long latency period between exposure and the development of cancer. This complicates the detection of relationships and, of course, makes it impossible to identify the carcinogenic risks to humans of agents newly introduced into the environment. Another common problem in epidemiology is that of exposure assessment. Often the specific exposure of interest cannot be measured directly so that surrogate measures must be used (e.g., occupation, place of residence). Since exposure data are usually derived from historical records generated for other purposes or from the recollections of subjects, opportunities for either random or biased misclassification of exposure are frequently encountered. In addition, appropriate study groups are often simply unavailable or inaccessible. Furthermore, it may be difficult to implicate specific carcinogens when the environmental hazards involve complex exposures to a variety of agents, the effects of which are difficult to disentangle. Still another difficulty is the inability of epidemiological studies to adjust for unknown risk factors, since control can be introduced only when the risk factors are already recognized. Thus, when a particular factor is related to exposure and disease outcome, it may be confounding and give the appearance of an association when in fact none exists, or

it may inflate or decrease the magnitude of an association. In view of these difficulties, it is not surprising that epidemiological data exist for only a small proportion of the many chemicals that have been shown to be carcinogenic in laboratory animals.

Determining Causality

In interpreting epidemiological findings, one is guided by the magnitude of the risk estimates, their statistical significance (likelihood of being due to chance), and the rigor of the study design to avoid various kinds of bias, including those related to selection, confounding, classification, and measurement (548,555). A determination of causality in epidemiology is bolstered by dose-response relationships, the consistency and reproducibility of results, the strength and specificity of the association, its biological plausibility, and other considerations. Thus inferences from epidemiology, as from other methods of inquiry, are not made in isolation, but should take into account all relevant biological information. Although epidemiological and other observations can accumulate to the point that a causal hypothesis is likely, it is not possible to ever prove causality (in the strict sense, a hypothesis can only be disproven). Nevertheless, a causal hypothesis can be sufficiently probable, as in the case of cigarette smoking and lung cancer, to provide a reasonable and even compelling basis for preventive and public health action.

Epidemiological Investigations

DESCRIPTIVE AND CORRELATIONAL STUD-Descriptive (or demographic) studies are concerned with identifying the distribution or patterns of disease in populations (547). It is basic tenet of epidemiology that diseases, including cancer, do not occur randomly, but fluctuate according to factors such as age. sex, race, time, and geographical location. The use of rates as measures of disease frequency is fundamental in describing patterns of cancer among these population groups. Prevalence, incidence, and mortality rates of cancer define the levels of risk prevailing in different populations and permit comparisons between groups. Descriptive surveys of cancer occurrence have been valuable in stimulating etiologic hypotheses and providing direction for analytical studies, which are then necessary to establish whether risks are associated with particular exposures (556).

Thus, important leads to etiology have come from population-based cancer surveys, which have revealed substantial international variations in cancer incidence, shifts in risk among migrant populations, changes in risk over time, and geographic peculiarities from mapping cancer mortality at the county level (550).

Descriptive studies may utilize the correlational (or ecological) approach, in which the rate of disease in a population is compared with the spatial or temporal distribution of suspected risk factors (556). This type of study may be particularly helpful in developing or refining hypotheses about carcinogenic risks, but falls short of establishing causal relationships. Correlational studies have the advantage of being much less expensive and time-consuming than analytical studies, because

they often utilize mass statistics previously collected for another purpose (554). The primary weakness of such studies, as with descriptive studies generally, is that data are collected on populations, rather than individuals. In other words, the rate of disease and the prevalence of exposures to variables of interest are known for various population groups, but information on the exposure status of persons who have the disease and those who do not within each population is not known. Thus, one cannot infer from the correlations of the population levels that the exposure of concern is associated with the risk of developing disease within each population (555). For example, in early surveys of lung cancer, the international variation in mortality rates and temporal increases among males appeared consistent with the reported patterns of cigarette smoking, but these correlations by themselves may have been circumstantial rather than causal, since a variety of other exposures (e.g., occupational hazards, air pollution) also varied concomitantly with the patterns of lung cancer. It took the analytical studies that pursued these leads to establish the cause-and-effect relationships between smoking and lung cancer. Correlational studies also may provide supporting evidence in evaluating relationships detected by analytical studies or laboratory data. This is illustrated by the more recent temporal increases in lung cancer among females, who have lagged about 20 years behind males in their adoption of smoking habits. Another example is the temporal variation in endometrial cancer incidence, showing a rise and fall in rates associated with the usage of estrogenic drugs for menopausal symptoms (557). Because correlational studies deal with aggregate exposures and disease occurrence at the population level, they are also often seriously limited by the imprecise measurements of exposure and the many potentially confounding variables. However, occasionally the correlational approach may provide evidence that strongly suggests a causal relationship, as with flatoxin, which has been linked to primary liver cancer based on concomitant geographic variation with the intake of contaminated foodstuffs.

ANALYTICAL STUDIES. In order to test etiologic hvpotheses and to identify and quantify carcinogenic risks to man, it is necessary to conduct analytical epidemiological studies (555,556). These studies are the principal means for determining the human health hazards of specific environmental exposures and agents. In contrast to descriptive surveys, data are obtained on disease occurrence and putative risk factors for specific individuals, using mainly the case-control or cohort method. Thus, by grouping exposed individuals and comparing them to those unexposed, after controlling for all other relevant variables, the risk of disease associated with exposure can be estimated. While it is important not to impose unnecessary constraints on epidemiological investigation, there are some methodological guidelines to consider in designing a study. In particular, the study groups should be sufficiently large and the time intervals between initial exposure and tumor onset sufficiently long to identify the lowest excess risk considered

important to detect. Reliable and valid estimates of exposure should be sought, with quantitative measurements to permit dose-response evaluations.

Studies should be designed in a manner that minimizes potential sources of bias and permits detection and control of confounding variables. Because the putative exposure may act either at an early stage or at a late stage of carcinogenesis (or both), it is possible to provide insight into the mechanisms of action by examining risk in relation to several temporal aspects of exposure. The studies should also strive to identify interactions of a particular exposure with other risk factors that may contribute to the carcinogenic process.

Analytical epidemiological investigations are essential to achieve a better understanding of the causes and means of preventing cancer. These studies cover a broad range of exposures with special relevance to public policy. Occupational studies have been a time-tested means of identifying carcinogens, but they require further emphasis to assess industrial hazards suspected on the basis of experimental, clinical, and field observations. Radiation studies also need increased emphasis to clarify the effects of low-level exposure and the shape of the dose-response curve. Drug studies have been useful both for direct evidence of risk associated with taking certain medications and also for insights into mechanisms of human carcinogenesis. They require an expanded effort to evaluate the late effects of estrogenic compounds, immunosuppressive and cytotoxic agents. and other medications suspected of having carcinogenic activity. Because of increasing clues to nutritional risk factors, emphasis is needed to clarify the role of dietary components including fat, fiber, micronutrients, trace elements, and food additives as well as cooking practices. The influence of general environmental pollutants remains an epidemiological challenge and requires more intensive evaluation through analytical studies, some of which may integrate appropriate environmental and body-burden measurements. Finally, multidisciplinary projects combining epidemiological and experimental approaches to chronic disease have recently gained impetus, and should be encouraged to evaluate the influence of oncogenic viruses, dietary and metabolic factors, host susceptibility, general environmental pollution, and other causative factors that may continue to elude detection by traditional observational methods.

Case-Control Studies. Case-control studies identify persons with a particular disease (cases) and a group of similar persons without the disease (controls). Information on past exposure to known or suspected risk factors is then collected from interviews, questionnaires, medical records, occupational logs, or other sources. The frequency of a particular exposure among the cases is compared with that in the control group, after making appropriate adjustments for other relevant differences between the two groups. If the proportion of cases with a certain exposure is significantly greater than that of the controls, an association between exposure and disease may be indicated. The case-control approach is especially well suited to studying relatively

rare conditions, such as most cancers, where the putative exposure is common in the general population (e.g., menopausal estrogens and endometrial cancer), or when the exposure is rare but accounts for a large portion of a particular cancer (e.g., vinyl chloride and liver angiosarcoma) (558).

Cohort Studies. These studies identify a group of individuals with a particular exposure and a similar group of unexposed persons and following both groups over time to determine subsequent health outcomes. The rates of disease in the exposed and unexposed groups are then compared. Information on disease frequency and other factors may be identified from medical records, occupational records, physical examinations, questionnaires, tumor registries, or death certificates. An association between exposure and disease may be indicated if the rates of disease are greater in the exposed group than in the unexposed group. These investigations may be based on current exposure and future health outcomes (prospective cohort study), but more commonly they utilize past exposure information and disease occurrence (retrospective cohort study). Instead of an unexposed comparison group, general population mortality or incidence rates (specific for age, sex, race, and calendar time) are often used to determine the expected number of cases of disease. This method assumes that in the absence of specific exposure the study group would have had the same probability of developing the disease as the general population, but differences in ethnic, socioeconomic and other variables must be considered in evaluating the validity of this assumption. The cohort approach is used mainly when it is possible to evaluate heavy exposures in clearly defined subgroups of the population. Thus it has been especially helpful in assessing the carcinogenic risk from occupational hazards or medical exposures such as radiation and certain drugs.

The statistical measures of association most often used in analytical studies are the odds ratio and the relative risk. The odds ratio is usually associated with the case-control study and represents the odds or probability of disease occurrence in the exposed compared with unexposed individuals. An odds ratio of 1 indicates no association between exposure and disease, assuming that proper adjustments for confounding factors have been made. The relative risk is a measure of association in cohort studies and is defined as the ratio of the disease rate in the exposed group to that in the unexposed group. For this reason it is also known as the rate ratio, and it permits a direct estimate of the risk of disease associated with a particular exposure. When the disease is rare, when incident cases are collected, and when cases and controls are representative of the same populations, the odds ratio is a good estimate of the relative risk (558). The magnitude of the odds ratio or relative risk is a measure of the strength of an association, although they are both subject to unknown or unmeasured biases which may distort their true value. Another term often used in epidemiology is the population attributable risk or etiologic fraction, which is the absolute amount of disease contributed or caused by a specific exposure.

Both the case-control and cohort methods are characterized as having certain strengths and weaknesses, although they complement each other in the testing of specific etiologic hypotheses. Case-control studies provide a more efficient means of studying rare disease, with fewer individuals needed for study as compared with the cohort approach; a shorter time period for study completion and generally lower costs are compared with the cohort method; an opportunity to evaluate simultaneously several causal hypotheses as well as interactions (the extent and manner in which two or more risk factors modify the strength of one another); and a capacity to evaluate the effects of common exposures as well as those rare exposures which may account for a large proportion of the cases. On the other hand, the case-control approach has some problems in directly estimating the risk associated with a particular exposure, except in special circumstances noted above; in reducing certain biases (e.g., selection, historical recall) that affect the comparability of cases and controls; and in providing detailed and precise information on exposures occurring in the past (557). Such investigations also, by definition, can only evaluate one disease or outcome at a time. The advantages of cohort studies are their capacity to estimate directly the risks attributed to a particular exposure, since incidence or mortality from disease is actually being measured; to reduce subjective biases by obtaining information before the disease develops; to determine associations between a particular exposure and multiple health outcomes; and to evaluate temporal relationships such as latency period and duration of effect. However, cohort studies are usually expensive and complex undertakings. They require large numbers of exposed individuals, particularly when relatively rare events, such as most cancers, are being investigated; long periods of follow-up to accommodate the latency period for chronic diseases, such as cancer; and special handling of problems associated with persons lost to follow-up and with biased estimates of risk as from the "healthy worker effect" of occupational studies (547).

Intervention Studies. Also referred to as experimental studies (557), these represent a third strategy of analytical epidemiology, which is especially useful in confirming causal relationships suggested by case-control or cohort studies. This approach may be applied in programs designed, for example, to reduce cigarette smoking and alcohol intake, modify diet, control occupational pollutants, or evaluate candidate preventatives (e.g., vitamin A supplements, hepatitis-B vaccine). Ethical considerations are obviously critical when developing this approach, and after intervention the statistical procedures resemble those employed for cohort studies.

Biochemical Epidemiology

It seems likely that some limitations of cancer epidemiology may be overcome by incorporating laboratory methods in analytical investigations. This has been a valuable routine practice in infectious disease epidemiology for the past century. This approach, sometimes called biochemical or molecular epidemiology (559), has only recently been developed in cancer epidemiology. There is current enthusiasm for these kinds of investigations, since they merge the strengths of observational human studies with newly developed experimental probes to derive information that could not be developed by epidemiology or laboratory study alone. The laboratory aspect may make it possible to define past exposures and subclinical or preclinical response to initiators, promoters, and inhibitors of carcinogenesis, or to evaluate hose-environmental interactions. There is special interest at present in using this technique to clarify carcinogenic risks associated with nutritional influences or specific environmental agents that can be detected in tissues or body fluids. Opportunities are also available to assess specific host factors that influence susceptibility to carcinogenesis, including endocrine parameters, immunocompetence, and genetic markers. Techniques are being refined to detect and quantify particular carcinogens or their metabolites in tissues or body fluids through chemical analyses, mutagenesis assays or immunological detection techniques. It is already possible to measure the interaction of specific agents with cellular target molecules, for example, through adduct formation with proteins and nucleic acids, excretion levels of excised adducts, or markers of altered gene expression (559). The task of identifying the effects of lifestyle and other environmental and host factors is obviously formidable. Biochemical epidemiology represents an innovative approach that may help to elucidate further the causes of cancer and the actual mechanisms of carcinogenesis.

Implications of Negative Studies

Epidemiological observations often provide regulatory agencies with health information which is helpful in establishing sound, defensible rules governing workplace exposures as well as exposure to general environmental contaminants. The clinical and laboratory observations and subsequent epidemiological confirmation linking hepatic angiosarcoma and exposure to vinyl chloride led directly to safeguarding of workers by regulatory limitation of permissible exposure levels. Although, as in other areas of science, epidemiological studies can never "prove" the absence of an association, sufficiently large and well controlled studies that fail to detect a hazard can also be useful. Specifically, for the types of populations studied, for the doses and duration of exposure to the putative agent, and for the assessed time following exposure, likely upper and lower bounds on the estimates of risk can be made, as well as the statistical likelihood of the study to identify an effect. As an example, the results of a large collaborative study of bladder cancer and artificial sweetners were interpreted as indicating that saccharin use was not a significant public health problem (560).

The likelihood of a study to identify an effect, if one is really present, is referred to as the statistical "power" of the study. Its estimation involves consideration of the size of the study group, the number of subjects

exposed, and the level of excess risk which is considered important not to miss. With this information, the power of case-control and cohort studies can be calculated and thus some index of confidence can be attached to the observation of no association. The statistical power of "negative" investigations should be routinely considered when such studies are reviewed.

Regulatory agencies are often in the position of making decisions, not simply on the effect of an exposure on the "average" exposed population, but on the effects among particularly susceptible subgroups of this population (e.g., the concern over the acute effects of air pollution on infants, the very old, and the infirm). The risk of malignancy following exposure to a carcinogen is rarely uniform across all subpopulations but is routinely modified by genetic constitution, demographic characteristics (e.g., age, race, sex), and exposures to other substances. Such risk modifiers can act either to enhance risk, such as the capacity of cigarette smoking to multiply the risk of lung cancer associated with asbestos exposure, or to reduce risk, such as the recently emerging evidence that certain dietary factors may inhibit the carcinogenic process. Epidemiological data are particularly—and often uniquely—relevant to these issues. Because of this, analyses of risk among subgroups of an exposed population are particularly germane to regulatory agencies. At the same time, these types of analyses are challenging to interpret, since variation between groups can easily occur due to chance or the differential operation of some type of bias.

Because of the central importance of epidemiology in cancer risk assessment, it is important to develop and strengthen programs of epidemiological research and to ensure the conduct of studies to look for and measure (if detected) health effects whenever relevant human exposure has occurred. Collaborative studies among Federal agencies and other groups are needed to evaluate urgent issues involving scientific, regulatory or public policy concerns, as well as to stimulate the epidemiological application of technical and data resources that are used by the government and other institutions for different purposes. Because of the international peculiarities in cancer occurrence, collaborative studies with investigators in other countries should also receive greater emphasis to pursue a wide variety of etiologic leads. These national and international collaborative studies can occasionally profitably include intelligent combined analysis of data already obtained from existing studies. In addition, further statistical investigation is needed to develop and test multicause and multistage models of carcinogenesis and to clarify the many issues involved in extrapolating results from experimental testing to the human experience.

Summary

Epidemiological investigations comprise one of the major strategies in creating the scientific base necessary for regulatory decisionmaking. Descriptive epidemiological studies, including ecological or correlational approaches, are useful in generating and refining hypotheses about potential cancer risk factors. Well

designed, well conducted, and well evaluated analytical epidemiological studies of either the case-control or cohort variety can test such hypotheses and provide the basis for causal inferences that are especially useful for public health decisions. The strengths of these studies lie in their capacity to assess directly the carcinogenic risks of environmental agents in humans (often with the dose levels and manners of exposure that are most relevant) and in their ability to provide insights into mechanisms of human carcinogenesis that can assist in extrapolation of risk estimates to dose levels or similar agents not yet studied. The lack of evidence of a hazard from an epidemiological investigation can also be useful in that, within the scope of the study, a likely range can be determined for the estimates of risk, as well as an estimation of the statistical power of the study to have detected an effect, if one were really present. However, the epidemiological method is often hampered by the long latent period that exists between exposure to a carcinogenic agent and the development of cancer, by the inability to control for the confounding influences of unknown risk factors, by problems in assessing specific agents when the human exposures are to mixtures, by the frequent absence of appropriate groups for study, and by a variety of difficulties associated with accurate and unbiased historical exposure assessment or disease ascertainment. In addition, the epidemiological method shares in the difficulties encountered by experimental studies in the direct detection of relatively low-level risks. In general, the strengths and weaknesses of the epidemiological method form a useful complement to the strengths and weaknesses of laboratory approaches to carcinogenesis. This, coupled with the direct assessments of risks and the results of interventions allowed by epidemiology, makes a strong argument for the conduct of epidemiological studies and inclusion of their results in regulatory decision-making, whenever relevant exposure has occurred in human populations.

Chemical Exposure Assessment*

Introduction

For the purpose of this document, a chemical exposure assessment is that process which seeks to define the quantity of a chemical which comes into contact, or may come into contact, with human populations.

This information then can be coupled with toxicity data to allow estimates of the potential risk from a carcinogen. Exposure assessments may also be used to identify and predict the effects of prospective control options or to measure the effectiveness of intervention activities designed to limit exposure. An assessment may address sources of human exposure, the intensity, routes and conditions of exposure, the frequency and duration of exposure, and the segments of population exposed. In certain simple situations an assessment may

only need to consider well defined individual exposures that can be measured directly. In more complicated circumstances, broad systematic surveys and extensive mathematical computations may prove necessary (561–563). Exposure assessment is often the most resource-demanding portion of the evaluations of chemical risks performed by the various Federal agencies.

The great diversity of different sources and routes of chemical exposure complicates a review of this area. "Many authorities agree that the weakest link in our understanding of the environmental health studies is our knowledge of human exposure" (564). In the Federal Government, the source of a specific chemical exposure has served as a guiding principle in defining the responsibilities of different regulatory agencies. Separate responsibilities exist for exposure to chemicals through the ambient environment (air, water, and waste sites), food, drugs, consumer products, and the workplace environment. In each area an array of specialized estimating techniques, data sources, and expertise have been developed. The hundreds of thousands of possible combinations of factors which might be included in a specific exposure assessment make it extremely unlikely that any two exposure assessments of the same chemical will be identical. Each Federal agency tends to focus on those aspects of exposure which are relevant to the laws which it administers (565-585). This section addresses some of the common considerations in methodology, as well as the broad issues and difficulties that are relevant to exposure assessments. Three appendices provide more detail on aspects of exposure assessments dealing with food, consumer products, and potential carcinogens in the environment.

Despite the overall grouping according to source of exposure, it is important to recognize that the path that a chemical follows from its source to the exposed target can be quite complex and may involve a number of different media: for example: (a) chemicals released into one medium (e.g., the air) may later transfer to other media (e.g., soil or water); (b) low ambient levels of a compound may be magnified through bioaccumulation of the compound through the food chain; (c) chemicals may be degraded and new compounds formed during environmental transport and disposition; (d) a chemical used in an article may be released into the environment years later when the article is disposed of as waste; and, (e) workers may carry a chemical home on their work clothing leading to a continuing exposure of their families

Methodology of Exposure Assessment

There is a diversity of procedures for estimating exposure (586). Although exposure estimating techniques are becoming more sophisticated at this time, there is no universally accepted minimum set of specifications for the reliable estimation of exposure (587). Exposure assessment is rapidly developing as a technical specialty, and even recent major reports on risk assessment only mention exposure briefly (1,588-591). There are, however, a number of general approaches discussed below which may be applicable to the conduct of most

^{*}Although the full committee participated in the framing of this section, primary responsibility was assumed by Dr. P. White, Mr. C. Church, and Dr. P. Preuss.

exposure assessments.

As a useful simplification, exposure assessment studies can be divided between those that rely more heavily on direct measurement of chemical exposure and those where exposure is predicted, or "modeled."

Monitoring Studies. The direct monitoring approach is exemplified by the development of practical personal monitors which measure directly concentrations of chemicals in the air which individuals breathe (592,593). An exposure assessment can be based on the range of personal exposure data generated by these monitors when used with an appropriate study group. However, it should be noted that the detection limits attained by personal monitors may pose limits in some studies and that most personal monitoring techniques currently provide information on average exposure but not on the time course or peak levels of exposure.

Monitoring of pollutants in the air and water has probably generated the greatest quantity of data among all exposure study approaches. Carbon monoxide exposure has been examined extensively using personal and population monitoring (594-601). Air pollution in the city of St. Louis was studied for several years (602,603), producing over one million data points per day during much of the program. The plume from a power plant (604) was studied intensively to define the transport and chemical transformation of the emissions from a single point source. Other techniques have been used to monitor emissions from entire cities to refine our understanding of the atmospheric transport of chemicals over large geographic distances (605). Significant monitoring studies have also been conducted for a wide range or organic and inorganic compounds in marine, freshwater, and surface environments (606-619). Extensive studies have been conducted to determine the levels of bioaccumulated pesticides and other chlorinated compounds in edible fish and shellfish.

In the workplace, monitoring is the principle methodology for determining the levels of exposure to specific chemicals. The monitoring is usually site-specific, and generally is not used to estimate the total exposure of the entire workforce. The Occupational Safety and Health Administration (OSHA) and the National Institute of Occupational Safety and Health (NIOSH) have established data management systems for assimilating and aggregating data from workplace measurements. At times, these two agencies collaborate in the preparation and publication of occupational health surveys that describe exposures in certain chemicals across broad segments of an industry or occupation.

There is no monitoring system in place for determining the levels of exposure to substances in consumer products. Exposure estimates are generally based on ad hoc short-term surveys, laboratory studies and mathematical modeling. For example, monitoring systems have provided for many years extensive information, on the levels of pollutants in the ambient air from the combustion of fossil fuel. No such parallel exists for these same pollutants generated from the combustion of fuel indoors. However, several monitoring

studies of indoor air pollutants have now generated data on exposures to combustion products from gas or kerosene appliances in the home (620-622).

Chemical contaminants in the U.S. food supply are monitored through the FDA's Total Diet Studies, an annual program which involves purchase and analysis of approximately 200 foods in grocery stores across the United States. Other monitoring systems used to measure exposure to chemicals in food are described in Appendix A.

Chemicals are also monitored in humans as a means of exposure evaluation (623-635). In a national program, human tissue has been analyzed on a regular basis to monitor selected, persistent chemicals such as DDT and PCBs (634,635). Tissue, secreta, or excreta, or any combination of these, can be assessed and compared to an appropriate reference (636). Such data can be used not only as a measure of overall individual exposure. but also as a measure of the effectiveness of regulatory actions when the monitoring is conducted for sufficiently long times to reveal trends. Chemicals or classes of substances for which there is some evidence that biological monitoring may be useful for detecting evidence of a substantial internal dose on an individual or group basis include: inorganic and organometallic substances, aliphatic, alicyclic, and aromatic hydrocarbons, aromatic amines, ketones, aldehydes, phenols, and pesticides (636-639). In summary, monitoring can provide estimates of individual exposure and data to define the connection between the sources of a chemical and exposure to humans. Monitoring activities can encompass studies of the sources of chemicals, the ambient environment, the human environment of the home and workplace, the food and water consumed, as well as the doses absorbed and retained by the human body.

Modeling is a second broad approach utilized in exposure assessment. Here the term "modeling" is used broadly to describe the construction of a methodology through which diverse data on different factors can be combined to predict levels of human exposure in the absence of complete monitoring data. Models are predictive tools used when direct measurement of exposure would be prohibitively time-consuming and expensive and when the relationship between the sources of a chemical and the eventual exposure is complex. A model may be constructed both to allow interpolation between experimental observations and to make predictions of exposure and circumstances that have not yet been experimentally studied.

Models often begin as an effort to describe and generalize theoretically a phenomenon observed in the laboratory or field. Monitoring data bases are used to construct mathematical relationships among variables of concern (e.g., meteorological parameters and the temporal/spatial distribution of chemicals emitted from a source). For example, the air monitoring data from the St. Louis and the power plant plume studies cited above have been used to build and refine modeling tools for predicting population exposure to contaminants in the air.

Given the wide variety of possible situations requiring exposure assessment, it is not surprising that many different models have been developed. A recent catalogue of the models available for use in regulatory decision-making identified 156 models, most of which can be applied to exposure assessments (640-642). A review of these models is beyond the scope of this section, but descriptions of various types of models are informative and can be found in the references (577,587,619,643-670).

In general, models are most helpful when average, or idealized estimates of exposure are satisfactory. Even though the level of sophistication of a particular model may be high, the predictions may still be crude, because any model is a simplified description of the natural phenomenon it depicts. There is a need for more research to validate most models, since the accuracy of their predictions is often unknown or only roughly known. In addition, the accuracy of the exposure estimates made by models can be no better than the quality of the input data used. The use of models does not compensate for poor data, and in certain instances, may obscure the uncertainty inherent in the exposure predictions. The use of scientific judgment in interpreting results is essential.

MICROCOSMS. The use of microcosms to study pollutant behavior provides an approach that combines features of both monitoring and modeling studies. The transport and transformation processes which occur between the source of a chemical and the eventual human exposure can be difficult and expensive to study in nature, and in the absence of data on these processes, models cannot be constructed. Microcosms, which attempt to recreate portions of the environment in the laboratory, can provide an alternate means for defining these processes. They have the added advantage that many variables can be controlled. Ideally, a microcosm, when properly validated, faithfully mimics the processes occurring in nature. By adding a chemical to such a system, its subsequent partitioning among the components of the environment, together with the associated chemical and biological conversions, can be studied in a single integrated experiment.

Marine, freshwater, terrestrial, subsurface or aquifer, and atmospheric microcosms have been studied (643,671-689). For example, the laboratory microcosm concept has received increasing attention recently in studies which seek to predict residential exposure to products of combustion or air pollutants (690,691). Limited validation of predicted, versus measured, levels of products of combustion in a residence has shown such predictions to be useful in assessing exposure (688,689).

HUMAN FACTORS. In exposure studies using monitoring, modeling, or a combination of approaches, the actual behavior of the human population under study must be taken into consideration. Some examples illustrate this: (1) the amount of time that an individual spends indoors compared with outdoors will affect the risks associated with either outdoor or indoor air pollution; (2) failure of workers to wear protective equip-

ment can increase their occupational exposure to hazardous compounds; and (3) consumer exposure to vapors during the use of various products will decrease proportionally with increases in the amount of ventilation provided in the home.

Human behavior is often evaluated by using information-gathering techniques somewhat similar to opinion polls or marketing studies. In such a study, a randomly selected population is asked a series of questions that define aspects of their behavior that can affect chemical exposure. On an occupational setting, "timemotion" studies of the periods of employee contact with varying work environments can contribute to an exposure assessment.

The exposure to direct and indirect food additives is monitored by the FDA by using a number of complex studies of human eating habits. Surveys of food additive use have been undertaken since 1973 with the help of the National Academy of Sciences, as have studies of the migration of chemicals from packaging materials into foods (575,579,583,585). Other related work has determined the frequency of consumption of individual foods and individual food serving sizes, with the data compiled in such a way as to be compatible with the population data found in the U.S. Census (582). These surveys and studies have produced a complex array of information which then is converted by the FDA into estimates of population exposure to direct and indirect additives in human food (576,584,694). Some of the same data are used by EPA in setting tolerances for pesticide residues in food crops (574).

STRUCTURE-ACTIVITY RELATIONSHIPS. Data now exist on the chemical, physical, and toxicological characteristics of a large number of chemicals, as does information on their transport, transformation, and environmental fate. Thus, some correlations now can be made between the structures of certain chemicals and the properties they exhibit, a process called structureactivity analysis. Under favorable conditions, these correlations allow rough predictions to be made of the characteristics of chemicals which have not been studied, reducing the amount of testing necessary to make at least an initial judgment of the risks which they may pose (658,695–702). Structure–activity relationships can be used to delineate the general properties of interest, but they are not generally a substitute for actual measurements.

The use of structure—activity relationships may allow preliminary exposure estimates to be made even when there is a scarcity of data on a specific chemical. Structure—activity analysis is a relatively new field, and the available tools are still crude. The user must exercise scientific judgment in interpreting the results, because substantial work remains to be done in refining and validating these techniques.

INTEGRATED EXPOSURE ASSESSMENTS. Most assessments performed in support of rule-making consider only a single source or route of exposure. Such assessments are appropriate in the case when only one source or route of exposure is significant. However, certain

recent exposure assessments on chemicals with multiple routes of exposure have considered all routes simultaneously (703). In a pilot effort, an exposure assessment has been prepared which considers six related halogenated solvents and all routes of exposure simultaneously (704,705). This assessment, in combination with hazard data, attempts to provide a risk assessment data base for all six solvents and all routes of exposure. Using this approach, all regulatory options and combinations of options may be considered in seeking the most effective means of reducing risk.

The preparation of such integrated exposure assessments for widely used chemicals is very difficult and resource intensive and requires the contributions of specialists in many disciplines. The difficulty and expense of integrated assessments may limit the broad application of these valuable tools.

Monitoring of the levels of chemical contamination in human tissue, blood and urine, as discussed above, can provide a second method of assessing the total human exposure to certain chemicals depending on metabolism and persistence in the body.

DATA BASES AND DATA MANAGEMENT. Research by the Federal regulatory agencies on human exposure has produced a large body of data which can be used in performing assessments. One recent index identified 319 data bases, many of which could be of value in exposure assessment (641). For example, monitoring data are reported regularly for National Pollution Discharge Elimination System (NPDES) permits, issued under the Clean Water Act, thereby affording information on 133,000 sources of chemicals found in surface waters (706). Monitoring data on air emissions from many sources and ambient air quality data from over 4,000 active air monitoring sites across the country are included in the Aerometric and Emissions Reporting System (AEROS) (707). In addition, many major research efforts (e.g., the St. Louis air pollution study) generate large data bases useful in exposure assessment (602,603).

Many data bases which do not deal directly with chemicals are also useful. The data of the U.S. Census provide information helpful to determine populations at risk. The National Climatic Center maintains detailed weather records, while the U.S. Geological Survey has a massive data base on water on a regional and state basis (708,709). The EPA has been constructing a comprehensive data base on organic chemical manufacturing.

As the number of data bases and models available for exposure assessment grows, it will become increasingly difficult to access and manipulate the large quantities of information and broad assortment of techniques without computer systems. Preliminary exposure assessments, reviews of the relative effectiveness of regulatory control options, and many other assessment-related tasks, which in the past required substantial effort, can be accomplished rapidly with limited accuracy at a computer terminal, calling upon data bases and models as needed (710-714).

Broad Issues Affecting Exposure Assessments

EXTRAPOLATION AND NONREPRESENTATIVE SYSTEMS. Exposure assessments, in a manner often analogous to risk assessments, may require extrapolation from a limited number of sites, to a large population. Such extrapolations carry with them uncertainties that need to be reflected in the final assessment.

In some cases, the exposure measured may not correlate well with the actual experience of the larger populations. In such cases, the nonrepresentative nature of the exposure measurement may give rise to an assessment that is inaccurate. Such errors can arise especially when microcosm measurements and models are used instead of extensive monitoring networks.

QUALITY ASSURANCE. Measurements of chemicals at the trace levels of concern, as needed for many exposure assessments, are difficult exercises in analytical chemistry and can lead to errors and uncertainties which should be considered in preparing an assessment. Until recently, extensive quality assurance information was not a routine part of most research, testing, and monitoring programs. To improve this situation, some agencies have instituted quality assurance procedures which require that records be maintained on the accuracy and reliability of every chemical measurement associated with research, monitoring, and compliance enforcement (715,716). Two examples of such state-of-the-science projects are the EPA-Chemical Manufacturers Association Joint Study on water treatment systems and the Love Canal Monitoring Study (717,718).

The quality assurance portion of the Love Canal Monitoring Study provided information on the accuracy and reliability of the analyses of chemicals at trace levels in air, water, and soil. The results demonstrated the limitations inherent in the measurement of complex mixtures in a variety of matrices. The overall results in the Love Canal studies suggest that the most advanced analytical techniques correctly identify trace levels of an organic chemical in complex environmental mixtures with approximately 70% certainty.

To date, the major efforts on quality assurance have been devoted to chemical analyses of environmental materials. As discussed, other data contribute to the exposure assessment, e.g. information generated through microcosm studies, human behavior, or modeling. There is a need for further validation of such tools to assure their accuracy and reliability in estimating exposure (719-722).

HIGH RISK POPULATIONS. Subgroups of the population may be particularly at risk from exposure to a chemical. These groups may be exposed to unusually high levels of a chemical or may be especially sensitive to the toxic properties of a compound. The existence of subgroups of either type could lead to a compound producing a substantial risk even though the average exposed individual may experience little, if any, risk. A thorough exposure assessment needs to consider the possibility that high risk populations or situations exist. Many, or most, current exposure assessments do not have enough data to allow an evaluation of the effects

of a chemical on high risk populations.

REFINEMENT OF ASSESSMENTS. At the initial consideration of a chemical, only crude data relating to exposure will generally exist. Such crude data may allow a rough estimate of likely exposure; this preliminary estimate may be useful to decision makers in assigning priorities and resources. It is also possible that crude data may allow an estimate of the maximum or minimum exposure that may be expected to occur. Such an analysis may effectively rule out, or indicate the presence of, a significant risk. Specifically, a "worst case" estimate, where all assumptions are chosen so as not to underestimate the possible exposure, may indicate that little risk exists if significant exposures are not predicted.

As more resources are devoted to an exposure assessment and more studies conducted, a refined assessment is generated. Often there will be several stages of refinement of an assessment, and the degree of refinement and accuracy finally required will be related to the certainty needed to enable risk management decisions.

RELATIONSHIPS TO BIOLOGICAL DOSE. The data generated in an exposure estimate must ultimately be coupled with biological data on the hazards presented by a chemical in order to determine the actual risk presented by the exposure. In order to achieve this coupling, the estimated human dose must be related to the doses received in the human or experimental studies that generated the information on the biological hazard. The type of exposure data needed can vary for different risk estimation procedures. For example, for risk estimation using a dose—response relationship that is not linear in the range of human exposure, information on the frequency distribution of different exposure levels in the population is needed, instead of simply an estimate of the average exposure.

This comparison can be complicated by differences in the frequency, duration, intensity, and route by which the two doses were received. This is particularly relevant since human exposures to a single chemical may follow many different patterns ranging from a high single exposure to a low level lifetime exposure. Humans will also often be exposed through several routes: ingestion, inhalation and dermal. Because of the diversity of human exposures, frequently there will not be toxicological data available which were obtained under the same exposure conditions. In those cases, careful review and expert judgment enter into the application of the exposure data.

Recently, there has been considerable interest in the possibility of relating human chemical exposures to hazard studies by utilizing biological indices of the effects of the different exposures. Specifically, it has been suggested that the quantity of a carcinogen reaching and interacting with DNA be directly compared in exposure and hazard studies. This topic receives fuller discussion in other sections of this document. It should be noted that at the present time, data to allow such comparisons are generally not available, and considerable research still is needed in order to assess the practical applica-

bility of this approach.

OVERALL LIMITATIONS OF EXPOSURE ASSESS-MENTS. Many exposure assessments are unable to distinguish all of the parameters that are important in determining possible health effects. These include variations of exposure with time, including, peak versus average exposures, and annual versus daily exposures. In addition, exposure assessments rarely, if ever, are able to estimate the dose that actually reaches a biological site where an effect may occur. As a consequence, some information is often available about the magnitude, frequency, or duration of exposure, but rarely is information available that is adequate to accurately determine all three. In addition, the population whose exposure is being assessed will often be exposed by more than one route from a number of different sources. As a result, estimates addressing single routes or sources of exposure are often inadequate to represent what is actually occurring. Environmental monitoring data, models, and microcosms may sometimes be inadequate, and of limited value, and may lead to a false sense of security if broadly applied without consideration and enumeration of their limitations.

Finally, there are a number of factors that are a part of both a hazard assessment and an exposure assessment which usually are not completely determined. These factors include the age distribution of the exposed population, special subpopulations that may be particularly sensitive to low doses, cumulative exposures, latency, host factors, or populations which may be exposed to particularly high concentrations. As a result, there are generally unknown data and uncertainties associated with exposure assessments at many points. It is important that the assessor describe the data that are missing, the associated uncertainties at each step of the assessment, and the assumptions made when filling the gaps.

At present, therefore, there is a great deal that can be improved in carrying out exposure assessments. Informed decision-making often requires consideration of a complex array of information, rather than a single numerical estimate of exposure. To the extent possible, the ultimate use of the data should be defined and the exposure assessment tailored accordingly. The limits of the assessment, both conceptual and methodological, need to be described, and all the uncertainties enumerated. Quality assurance programs and efforts should be included as well as a description of efforts made to validate laboratory studies and mathematical modeling; in general, careful scientific review of all elements of an exposure assessment should be stressed.

Summary

Exposure assessment is a co-equal component, along with hazard assessment, in the evaluation of human risk. The assessment of exposure to chemicals by direct routes (e.g., breathing emissions from an industrial stack) and indirect routes (e.g., pollutants accumulating through the food chain) depends upon analytical chemical monitoring data, results of modeling estimates, and assumptions needed to fill in data gaps. The field of

exposure assessment is rapidly developing and only recently have attempts been made to develop systematic approaches to the problem. Although current efforts are often associated with considerable uncertainty, the increasing sophistication being brought to bear on the issue holds the promise of greater confidence in these estimates in the future.

Methodologies can be broken up into two broad categories: monitoring studies, which depend more on the measurement of exposure, and mathematical modeling, which use analysis to define the exposure. Microcosms, as samples of the environment, provide data for both approaches. Human factors, however, are important and must be considered.

An important part of the assessment process is the marshaling of the appropriate data bases, and some of the data bases used by three regulatory agencies are listed in appendices.

Broad issues affecting assessments include the question of extrapolation from representative systems to large populations, quality assurance, the question of high-risk populations; and the approach of coupling exposure data to actual biological dose to improvement assessments, are emphasized, with a discussion of the limitations of assessment as it is being done today.

Utilizing Scientific Data in Assessing Human Cancer Risk Associated with Chemical Exposure*

Introduction

Assessment of human cancer risk associated with some specified chemical exposure is a complicated scientific endeavor that requires careful review of all pertinent information by appropriately trained individuals. This process relies heavily on information derived from epidemiological, clinical, and long-term animal studies. Short-term test results and information on structure-activity relationships, comparative metabolism, pharmacokinetics and mechanisms of action also contribute, in varying degrees, to the assessment.

While there is general agreement within the scientific community about the steps involved in the risk assessment process, a number of specific issues related to these various steps are still unresolved; e.g., the methodology for combining positive and negative data when evaluating carcinogenic potential and the selection of a dose-response model for making low-dose risk projections. The execution of any given risk assessment may also be hampered by the existence of critical gaps in the data underlying the assessment. Because these issues and data gaps can be pivotal in the process, it may be necessary to make judgments about these unresolved issues and assumptions to adjust for deficiencies in the underlying data base that will permit risk assessments to be completed. These judgments and assumptions

should be plausible and, to the extent possible, be based on the specific information on hand. However, the validity of these judgments and assumptions often cannot be conclusively established, so the risk assessment process should not be viewed as strictly "scientific" in the usual sense of the word. Instead, risk assessment involves a complex blend of current scientific data, reasonable assumptions and scientific judgments that permit decisions to be made in the absence of complete information.

With the passage of time the scientific data base grows, information gaps are filled, and perceptions change. This process of evolution and maturation results in a need for periodic reformulations and restatements of the appropriate guidelines for risk assessment.

This section describes the basic framework for carcinogenic risk assessment and attempts to reflect what general agreement currently exists within the scientific community. It is intended to provide a bridge between the scientific data base on carcinogenicity developed in previous chapters and the various Federal regulatory agencies that are charged with the responsibility of protecting the general public from exposures to carcinogenic agents.

Steps in the Assessment Process

There are four steps or components that are typically involved in carcinogenic risk assessment. The first, which is often referred to as hazard identification, entails a qualitative evaluation of both the data bearing on an agent's ability to produce carcinogenic effects and the relevance of this information to humans. The second, exposure assessment, is concerned with the number of individuals who are likely to be exposed and with the types, magnitudes, and durations of their anticipated exposures. The third component, hazard or dose-response assessment, uses the information on carcinogenicity from the hazard identification phase together with mathematical modeling techniques to estimate the magnitude or an upper bound on the magnitude of the carcinogenic effect at any given dose level. Finally, one may combine the information from the first three components or steps to characterize the carcinogenic risk associated with the expected human exposure to the compound of interest.

HAZARD IDENTIFICATION. Due to the lack of complete information, there may be some uncertainty involved in determining whether or not an agent poses a carcinogenic hazard to humaas. While information on human exposures and effects is particularly valuable, too often such data either do not exist or are inadequate, and one must rely primarily on information from longterm animal bioassays in assessing the potential for carcinogenic response in humans. In reviewing data, each relevant study must be evaluated as to the limits of inference implied by the experimental design, the results, and the conclusions. Although strict criteria for evaluating carcinogenicity data have not been developed for all circumstances, there seems to be reasonably good agreement among scientific groups as to the general elements to be included as a part of qualitative

^{*}Although the full committee participated in the framing of this section, primary responsibility was assumed by Drs. M. Hogan and D. Hoel.

carcinogenic assessments (309,483,489,531,588,723,724).

Epidemiological studies can, under certain circumstances, provide direct measures of carcinogenic effects in humans. For example, some epidemiological investigations are able to associate cancer incidence or mortality with exposure to specific chemicals, industrial processes, or components of an individual's life style. Analytical studies, i.e., case control and cohort studies, are especially important indicators of possible human carcinogenic risk, but case reports and descriptive studies may also be supportive. Consistent results in independent studies, freedom from bias and confounding factors, reliable exposure data, reasonable follow-up time, dose-related responses and high levels of statistical significance are among the factors leading to increased confidence in a conclusion of carcinogenicity. Even if an epidemiological investigation fails to demonstrate an increased incidence of carcinogenicity among the exposed study members, upper and lower confidence limits on the risk measure used in the study can indicate a range of probable risk that could be incurred by a similarly composed segment (i.e., in terms of age, race, sex, etc.) of the general population.

Evidence from long-term animal bioassays constitutes the second major class of information bearing on the carcinogenicity of chemicals. The primary factors that are considered in the evaluation of carcinogenicity are higher tumor incidence and shorter latency in treated groups relative to controls. Findings arae strengthened by the existence of positive effects in more than one treated group or sex. Additional support comes from positive results observed with different routes of exposure, in replicated experiments, in different animals strains and species, and in multiple organs or tissues. Attention is also given to the magnitude of tumor increase in treated animals, the existence of dose-related trends, the degree and extent of malignancy at a given site as a function of dose and time, the evaluation of concurrent nonneoplastic pathology in treated animals, and the finding of even a limited number of rare tumors in the exposed group(s). In addition the similarity, or lack of it, between contemporary and historical control tumor rates can affect the carcinogenicity evaluation.

Beyond these two major types of evidence, additional information bearing on the qualitative identification of carcinogenic potential may be gained from short-term test results, including testing by genetic toxicity, macromolecular binding and in vitro transformation; short-term in vivo tests such as skin painting experiments; comparative metabolism studies among species; review of known physiological, pharmacological, biochemical and toxicological properties of the chemical; evaluations of contaminants, degradation products and metabolites of the chemical under study; and consideration of the carcinogenic potential of structurally related compounds. At the present time these latter categories of information will often play a supporting or confirmatory role in the process of human cancer hazard identifica-

tion. However, when specific information on comparative metabolic and kinetic data, exposure route-dependent responses, etc. is available, and its relevance demonstrated, it may be useful in determining the likelihood of possible human cancer hazard.

The relevant data discussed above are reviewed and assessed to determine if the chemical is likely to be carcinogenic in humans. The greater the consistency of the information, the greater the confidence that the agent poses or does not pose a carcinogenic hazard for humans.

EXPOSURE ASSESSMENT. Without exposure, there obviously is no risk to the human population, regardless of the potency of the carcinogen under consideration. Thus, exposure evaluation is a critical component of the risk assessment process. One of the most frequently encountered problems when attempting to use epidemiological data in cancer risk assessment is the lack of appropriate historical exposure information.

In an exposure analysis, a group or groups of individuals are identified and described with respect to their population size and composition, and the route (e.g., inhalation, dermal, ingestion), magnitude, frequency, and duration of their exposures. In addition, consideration must be given to both direct exposures (e.g., drugs or occupation) and indirect exposures (e.g., food or air) that results from a chemical's transport through various environmental compartments before impacting on humans.

For a given exposure source the concentration or amount of the chemical in that medium is determined by measurement, estimated by modeling, calculated from physical-chemical properties and other information on the agent, or projected from data on surrogate chemicals. Common values might include ambient airborne concentrations given as parts of chemical per million parts of air or the the number of milligrams of a drug per dose. Measurements, estimates or assumptions then need to be made concerning the exposure of humans to the chemical through a given medium; e.g., the frequency of consumption of a specific food containing a pesticide residue or the frequency, pattern, duration and conditions of use of a consumer product containing a cleaning solvent. Next, assimilation of the chemical into the body is considered; again there is reliance on whatever pharmacokinetic data or estimates are available. Values may be expressed in a number of ways such as the percentage of dose absorbed from the gastrointestinal tract, the movement of so many micrograms of chemical across a given area of skin over a certain time period, or the percent of the inhaled material absorbed in the respiratory tract.

All the above information, i.e., population identification and characterization, chemical concentration in a given medium, the nature and pattern of exposure and finally the assimilation of the chemical into the body, is assembled and analyzed. Information on multiple sources of exposure may then be combined into a single measure of overall exposure.

In some instances, it may not be possible to identify

or isolate the particular chemical or chemicals in a complex mixture or manufacturing process that are likely to pose a carcinogenic risk to humans, and so detailed exposure quantitation becomes difficult. Furthermore, tremendous variability in exposure exists among members of a population. People change jobs or place of residence, convert from well water to municipal water, occasionally take prescription/non-prescription drugs, etc. Each of these differences among persons contributes to the uncertainty associated with any exposure value. To the extent possible, the degree of uncertainty in exposures should be discussed and estimated. Characterization of uncertainty may involve calculation of a "worst-case" exposure or upper bound on exposures, consideration of a range of likely or possible exposure values, or even the computation of confidence bounds associated with a specific model-based exposure estimate.

Dose-Response HAZARD OR ASSESSMENT. Hazard or dose-response assessment is a quantitative exercise that attempts to describe the expected human response to any given level of a carcinogenic exposure. The response is typically characterized in terms of either a specific estimate or an upper bound on the underlying risk. This step in the risk assessment process relies heavily on data from exposed humans and on observations of animals employed in long-term cancer studies. Long-term animal screening studies have traditionally focused on the detection of carcinogenic potential. As a result, they are usually based on exposures at or near the test animal's maximum tolerated dose level. Since these doses are often orders of magnitude higher than those encountered by humans, some form of mathematical low-dose extrapolation procedure is required to estimate the expected response at the exposure levels typically of concern in the environment. In addition, a species extrapolation or scaling factor is usually employed when estimating anticipated human cancer risk from experimental data. Low-dose extrapolation may also be an important consideration for epidemiologically-based assessments, since the available human data may often involve relatively high level occupational exposures.

Low-Dose Extrapolation. Low-dose carcinogenic risk estimation is based on mathematical modeling that attempts to characterize explicitly the unknown, underlying relationship between exposure and response or to place an upper bound on the dose-response relationship.

Linear extrapolation procedures are often employed for estimating the low-dose risk. In the absence of background response a commonly used approach is to draw a straight line from an upper (binomial) confidence limit on the observed response rate at the study exposure level to the origin. If background is present, the procedure can be modified by making specific assumptions about the nature of background in order to take its effect into account (725,726). Two of the more appealing features of this procedure are that it is easy to apply and that it will generate an upper bound on the unknown,

underlying cancer risk in most instances. The most frequent criticisms directed against it are that it may be unduly conservative and that it fails to utilize all of the study data if several exposure levels are employed. Recently, an interesting modification of the linear extrapolation procedure has been proposed by a number of investigators (727,728). They suggested that some type of mathematical dose—response model should be fit to the experimental data, that the estimated response and associated model-based confidence limit for a preselected level of exposure should be determined, and that linear extrapolation should then be used to determine an upper bound on risk for the low-dose level of interest.

When low-dose extrapolation is based on an attempt to characterize the underlying dose-response relationship, one of three general classes of models (i.e., tolerance distribution models, mechanistic models, or time-to-tumor models) is usually employed.

The tolerance distribution models assume that each member of a population has a threshold or tolerance level below which that individual will not respond to the exposure in question. It is also presumed that the variability among individual threshold levels can be described in terms of a probability distribution. For this class of models the probability that a particular individual will exhibit a carcinogenic response when exposed at dose level d is the same as the probability that the individual's tolerance level is less than d. The familiar probit, logit, and Weibull dose-response models can all be generated by adopting different probability distributions to describe the population's tolerance variability (729). The best known of the low-dose extrapolation techniques based on tolerance distribution models is the Mantel-Bryan procedure (730,731), which employs a probit model. Although the Mantel-Bryan procedure was originally regarded as a conservative approach to the estimation of low-dose risk since it employs a fixed, presumably conservative estimate of slope and extrapolates from an upper confidence limit on the observed experimental response, subsequent research (732) has shown that in the low-dose region the probit model tends to produce relatively high "safe-dose" estimates when compared to other extrapolation procedures. As a result of this characteristic and other criticisms that have been raised agianst the Mantel-Bryan procedure (732), its use in quantitative risk assessment has markedly declined.

The class of mechanistic models, i.e., the one-hit, multistage, and multi-hit models, derives its name from the presumed mechanism of carcinogenesis upon which the models in the class are based. Each of these models reflects the assumption that a tumor originates from a single cell that has been damaged by either the chemical or one of its metabolites.

The simplest of the mechanistic models is the one-hit model (729) which assumes that damage to the target cell sufficient to cause a tumor results from a single "chemical hit." This model is essentially linear in the low-dose region. Since it has only a single parameter other than background, it will not always provide an

adequate fit to the experimental observations.

The multistage model, developed by Armitage-Doll (733), is perhaps the most frequently employed of all low-dose extrapolation models currently in use. It reflects the observation that a developing tumor goes through several different stages, which can be affected by the carcinogen in question, before it is clinically detectable. The multistage model will often be approximately linear in the low-dose region, and so its risk estimates are sometimes regarded as being relatively conservative. Furthermore, the procedures most commonly used to generate upper confidence bounds for the multistage model lead to estimates that are linear at low doses (734).

The multihit model (729,735) assumes that the target cell must absorb at least k chemical hits before the cell achieves its carcinogenic potential. Some investigators regard the generalized multihit model, where k can be any nonnegative value, as being more flexible than the multistage model; and its use in risk assessment has been recommended (497). However, there are research results (736) that indicate a number of important, practical problems associated with its application, such as the fact that model hyperlinearity may produce extremely conservative estimates of safe dose levels, or that the multihit model can also indicate a "safe dose" at levels higher than some doses that actually produced deleterious effects.

The last class of models employed in low-dose extrapolation is the time-to-tumor models. These models, which attempt to describe the complex relationship between dose, tumor latency, and cancer risk, include the lognormal distribution (725), the Weibull distribution (737), and the Armitage-Doll (733) and the Hartley-Sielken (738) models. While time-to-tumor models can lead in many instances to a more complete characterization of the underlying carcinogenic process, the quality of the available data may not permit their application or, at the very least, is often not sufficient to allow any discrimination among such models using goodness-of-fit criteria. Furthermore, a recent analysis (739) of a simulated data base containing information on a time-totumor occurrence indicated that low-dose risk estimates generated by a variety of extrapolation procedures occasionally differed from the actual risk by three or more orders of magnitude even when this additional information was included in the modeling process. It also appeared that, on the whole, incorporation of time-totumor data did not substantially increase the precision of low-dose risk estimation relative to modeling based only on quantal response information.

When data on the time of tumor occurrence or detection are available, some investigators have proposed that alternative measures or risk based on the concept of time-to-tumor, such as the "mean-free" dose or "laterisk" dose (740), be used in the cancer assessment process. However, a detailed evaluation of these measures (741) indicated that they suffer from a number of shortcomings and are probably most meaningful when used in combination with the more traditional measures, like

the lifetime probability of tumor.

As the preceding discussion has indicated, uncertainties are involved in the use of any of the commonly employed extrapolation models. Furthermore, goodness-of-fit to the experimental observations is not an effective means of discriminating among models (493). Thus, no single low-dose extrapolation procedure has yet gained universal acceptance within the scientific community.

Model selection is further complicated when background tumor incidence is present and presumed to be induced by a mechanism or mechanisms distinct from that associated with the chemical under study. In this instance the various mathematical models may fit the observable data equally well and still generate low-dose risk estimates that may differ by many orders of magnitude. On the other hand, if background additivity is assumed, i.e., if it is presumed that there is a common mechanism of tumor induction, then all models are essentially linear in the low-dose region (734). Even if only a small portion of the background incidence is associated with the same mechanistic process as the study chemical, linearity will tend to prevail at sufficiently low doses (742).

All of the preceding discussion has implicitly assumed that the carcinogenic dose-response relationship of interest is that which associates the probability of tumor onset with the administered dose or exposure levels. However, in the case of experimentally-derived data, saturation of critical enzyme systems may occur at high dose levels, and so the real dose of interest may be the biologically effective dose that reaches the actual target site. Since the relationship between the effective dose and the administered dose may be non-linear, pharmacokinetic considerations can significantly modify lowdose risk estimates regardless of the model employed (519,520,743,744). Incorporation of mechanistic and pharmacokinetic data as well as information on target organ dose, when it is available, may, therefore, significantly enhance the usefulness of mathematical modeling procedures. In addition, consideration of any data on dose-dependent changes and differences in metabolism and repair, and appropriate adjustment for partial lifetime and intermittent exposure and competing risks might also reduce uncertainty associated with the mathematical modeling process.

Other Estimation Procedures. Traditionally, the approach to quantitative hazard evaluation for noncarcinogenic, toxicological endpoints has been based on the use of safety factors. With a safety factor approach, an "acceptable" exposure level is determined by dividing the no-observed-effect-level (NOEL) from laboratory-based chronic toxicity tests by an appropriately chosen safety factor. In theory, the safety factor that is selected attempts to reflect both the possibility of an increased sensitivity of humans relative to laboratory animals and the variation in susceptibility within the human population.

In spite of its common use, there are a number of potential problems associated with the safety factor ap-

proach. The observation of no treatment-related effects at a given dose level may depend, at least in part, on the number of animals exposed at that particular level. Furthermore, the determination of a NOEL ignores the shape of the dose-response curve, even though it would seem that a curve that has a shallow slope in the experimental NOEL region potentially represents a greater toxicological hazard than one that rises steeply in this region. In addition, there is no biological justification for the general use of any specific safety factor. Another important consideration that would argue against the use of a safety factor approach in cancer risk assessment is the fact that this approach assumes the existence of a true population threshold below which no adverse effects can occur. Even if the concept of individual thresholds could be supported, the well-recognized genetic variability in the human population would effectively prevent the estimation of a general population threshold value. Moreover, given the high level of background cancer present in the human environment, it seems unlikely that one could rule out the possibility that a new chemical exposure, however limited, might augment an already ongoing mechanistic process and thereby produce a collective or additive exposure that exceeds the unknown threshold level (745). While recent attempts to categorize carcinogens as acting directly or indirectly on the genome have renewed interest in the possible use of the safety factor approach, most scientists believe that there is not sufficient biological understanding of these processes to make such classifications for the carcinogenic effect of an agent on the total organism at the present time (746). Even if this distinction is made, however, the implementation of a safety factor approach would still be problematical because of the various limitations of this procedure that have been discussed above.

Another measure that is occasionally proposed for use in cancer hazard evaluation is the relative potency index. Relative potency calculations are based on standard measures of toxicity, such as the effective dose for 50% of the tested group (ED_{50}). Such measures are limited from an extrapolation viewpoint in that they attempt to compress an entire dose-response curve into a single number suitable for comparative purposes. Therefore, depending upon which toxicological measure underlies its derivation, a relative potency index may offer little insight into the relative risk in the low-dose region.

Extrapolation of Low-Dose Risk Estimates to Target Populations. Low-dose risk estimates derived from laboratory animal data still must be extrapolated to the human population. This process is complicated by a variety of factors that differ among species and potentially affect the response to carcinogenic exposures. Included among these factors are differences between humans and experimental test animals with respect to life span, body size, genetic variability or population homogeneity, existence of concurrent disease, pharmacokinetic effects such as metabolism and excretion patterns and exposure regimen.

The traditional approach to the species extrapolation issue has been to use some standardized baseline for making interspecies comparisons. Commonly employed standardized dosage scales include mg/kg (body weight)-day, ppm in the diet or water, mg/m² (body surface area)-day, and mg/kg (body weight)-lifetime. The choice of a particular dosage scale will obviously affect the magnitude of the projected risk. For example, calculating average daily dose on a body weight rather than a surface area basis can reduce the estimated human risk by approximately 6-fold when rat data are used for modeling human response and up to 14-fold for mouse data (588). A number of studies (747-749) have used the few epidemiologic and laboratory data sets available in the literature involving comparable carcinogenic exposures to evaluate the relative reliability of these various scales for animal-to-human extrapolation. While no single scale emerged as the clear choice in all cases, it does appear that the best agreement between observed and predicted human cancer risk is often obtained when a mg/kg-day or a mg/m²-day dosage scale is employed (748,749). However, it must be emphasized that experience in this area is very limited and that the use of any standardized dosage scale for species scaleup is only a crude approximation to a more detailed adjustment that considers the spectrum of factors that can lead to differential species responses. Furthermore, recent research (750) into this issue has indicated that the previously observed (749) strong quantitative correlation of various agents' carcinogenic potencies in mice and rats may be explained by a corresponding correlation of maximum tolerated dose levels for these same agents. The general implications of this finding for quantitative species extrapolation are not fully understood at this time.

In addition to the empirical comparisons discussed above, there are a number of biological issues that bear on the choice of a particular dosage scale in species scale-up. For example, it may be important to know whether the parent compound, one or more of its metabolites, or some combination of the two, is responsible for the observed carcinogenic activity. Similarly, the nature of the biochemical target site and the various factors determining the rate, duration, and extent of the toxicologic reaction may be relevant to the selection process. However, while any meaningful information on the underlying carcinogenic mechanism(s) should, if possible, be taken into account when choosing a dosage scale, such specific information often will not be available.

While no extrapolation across species is involved when epidemiological data are used in risk estimation, the marked variability in human sensitivity must be taken into account. Furthermore, there are a number of other problems that may be encountered with attempting to use epidemiological data to construct estimates of lifetime risk for a subset of the general population. In the absence of specific knowledge about the synergistic or antagonistic potential of the exposure of interest, additivity of effects is often assumed. This assumption can lead to either an under or over estimation

of the true carcinogenic risk presented by the exposure under consideration.

Epidemiological investigations are often conducted for only a limited period of time, and failure to adjust properly for incomplete follow-up of subjects can significantly affect the estimation of life-time risk. Similarly, estimation of lifetime cancer risks is also complicated if the available epidemiologic data involve exposures that fall far short of a normal lifespan. For example, if a multistage mechanism is assumed, then the effect of early termination of exposure will be dependent on the stages of the carcinogenic process that are influenced by the exposure (553,751).

RISK CHARACTERIZATION. The final step in the risk assessment process, risk characterization, typically involves a multidisciplinary evaluation of all relevant data including the qualitative evidence regarding the likelihood that the chemical in question poses a human cancer hazard, available information on potential or actual human exposure, and the quantitative results of dose response and species extrapolation modeling. The strengths and weaknesses, the biological relevance, and the quality of all data used to develop the characterization of the human cancer risk associated with anticipated or known exposures should be considered, and an attempt to qualify or quantify the various uncertainties in the characterization of risk should be made when possible.

As discussed earlier, the qualitative evidence is evaluated by a weight-of-the-evidence approach. Uncertainties in the data are resolved using best scientific judgment. While no overall measure can be given of the uncertainty, the decision-maker needs to be aware of the strengths and limitations of the qualitative data involved in the risk characterization process. The results of a total risk assessment should be expressed in such a way that there is a clear distinction drawn between a chemical for which the qualitative evidence is overwhelming and a chemical for which the qualitative evidence is only marginally persuasive.

As developed in the section on chemical exposure assessment, the exposure assessment may contain numerous uncertainties. For example, the assessment may reflect inherent limitations, such as those associated with analytical monitoring or modeling techniques. Further, assumptions are sometimes necessary in the absence of data in order to generate an assessment; e.g., the amount of soil ingested by a child. In many such instances, "worst case" assumptions are made in an attempt to err on the side of public safety. The exposure assessment should be expressed in such a way that these uncertainties and assumptions are evident to the decision-maker.

Description of uncertainty is also important in the development of the quantitative estimates from doseresponse and species extrapolation modeling. Unfortunately, the description of uncertainty in this step of the assessment process has often been limited to a description of the statistical uncertainty involved in low-dose extrapolation as expressed through the calculation

of confidence limits for risk estimates.

As has already been noted, the choice of a particular low-dose extrapolation model can have a pronounced influence on the estimated low-dose risk. Therefore, it has been proposed (752) that an indication of the variability introduced by model selection be obtained by considering the range in the magnitude of low-dose risk estimates associated with the more commonly employed models

Furthermore, if laboratory data are utilized in estimating low-dose risk, some attempt can be made to describe the biological variability associated with the process of species scale-up by contrasting estimates based on different dosage scales and animal test systems

Finally, it is important in the characterization of human cancer risk to summarize briefly any judgments or assumptions that may have entered into the risk assessment process to insure that they are clearly differentiated from scientific fact (1).

Emerging Areas of Science Expected to Impact on Regulatory Actions*

Although no one can predict with certainty what and when major breakthroughs will occur in science, many emerging areas of science can be expected to significantly affect cancer risk assessment and related regulatory decisions in the future.

Greater knowledge of the metabolic pathways of chemical compounds and the pharmacokinetics of such agents in various animal species should help regulatory agencies to ascertain better whether carcinogenicity tests have been performed in an animal assay system which is relevant to man. Information on metabolic pathways for activation of a chemical carcinogen known to be important in humans but not present in the animal assay system used will influence the data evaluation and interpretation for human risk estimation. Similarly, factors such as diet, age, stress, sex, hormonal status, etc. can influence the metabolic conversion of chemicals to ultimate carcinogens and may be important in hazard evaluations conducted on proposed compounds.

DNA adducts remaining in organs in vivo, in the absence of tumors in a test animal species, may be innocuous; may indicate physiological peculiarities in the test system that prevent progression to overt tumors; or may suggest that overt manifestation to neoplasm has yet to take place. Such inhibitory factors may or may not be present in other assay systems or in man. Alternatively, failure to demonstrate tumor formation may indicate the need for promoter activity or interaction with other modifying agents. Substantial adduct formation without resulting biological manifestations may suggest either that the lesions induced are innocuous or dictate that more testing is needed to access the human exposure experience (i.e., multiple exposure factors, promoting agents, nutrition, etc.). In any

^{*}Although the full committee participated in the framing of this subsection, primary responsibility was assumed by Drs. R. W. Hart and A. Turturro.

event, it is evident that additional research is required to define more precisely what DNA adduct formation means in term of human risk.

Most prescreen assays for chemical carcinogens presently rely primarily on chemical interaction with DNA (point mutation assays, DNA damage and repair). This approach may be inadequate, since certain chemicals may not induce cancer by direct interaction with the DNA. New assays for carcinogenicity are being developed to include animal models where these factors can be taken into consideration in the evaluation of chemical agents.

Since tumor induction is generally considered to be a multistage process, each stage may be under independent genetic control. Test systems are being developed and used that are based on animals believed to be defective in genetic factors similar to those defects that predispose humans to cancer induction. Such test systems may also be used to screen sensitivity to selected carcinogens for subpopulations of humans with similar genetic characteristics.

A relationship seems to exist between altered DNA and carcinogenesis. The presence of carcinogen-altered DNA is being explored as a possible test system to detect persistent damage at the molecular level after exposure to suspect carcinogenic chemicals. Such a system could be used to screen exposed human populations for altered DNA bases using monoclonally derived specific antibodies to modified bases. Such assays may also help place in perspective effects on the DNA arising from endogenous vs. exogenous DNA damaging agents. This screening would be an example of using a marker for altered cells important to the carcinogenic process. Additional bio-markers should be developed for cells in all stages of carcinogenesis. Such tests might be expected to identify the carcinogenic potential of suspect agents and the population(s) at risk for possible future development of cancers.

Certain DNA base alterations may occur in regions of the genome that currently seem to be unimportant for carcinogenesis. It is important to know which alterations are necessary for initiation of a tumor cell and which are masked or not expressed. Information is being assimilated on types of alterations in "control" regions and the relationship of the number of altered bases in these regions to tumor formation. These data should provide information needed to differentiate those insults to DNA that are critical from those that are not. If multicritical sites must be altered in a particular manner, this approach may be used to examine multistage phenomena of cancer. The measurement of risk from particular carcinogens at the cellular level would then be based on a more selective effect than the level of total DNA adducts.

Other areas of research that may in time have an effect on regulatory decisions include the development of model systems to assay for compounds influencing promotion and progression of initiated populations, elucidation of factors controlling cellular differentiation, and tests for chromosomal rearrangements and onco-

gene activities and the development of computer-based techniques for both the identification of the portions of molecules that are important in carcinogenesis and the modeling of animal (the "computer mouse") and human response to tumor induction. Each of these areas of study will not only expand our knowledge of mechanisms but may also permit a more realistic assessment of risk. Besides improving assessment, increased knowledge of mechanism may also result in a greater ability to render carcinogenic agents biologically innocuous, removing hazards.

Summary

The assessment of human cancer risk associated with different chemical exposures is seen to be a complex mixture of currently available scientific data, assumptions and judgments based on prevailing scientific thought, and policy decisions. The process involves the identification of potential human cancer hazards, usually based on epidemiological, clinical, and long-term animal studies, the determination of potential human exposure profiles, the estimation of the unknown, underlying relationship between exposure and response, and, finally, the combination of the results of these exercises to characterize the expected human cancer risk. Some degree of uncertainty is involved in all phases of this process, and as a result, no simple, standardized format has yet been developed for evaluating human cancer risk, even though a number of basic principles have emerged. It is apparent that human cancer risk assessment is still in an evolutionary state and that a number of emerging areas of science will impact on this process in the future.

Appendices

Appendix A: FDA Information Sources for Exposure Estimates

Survey, epidemiological, toxicological, and analytical data used by the FDA in determining the human health consequences of exposure to many of the categorical agents which the agency regulates are derived from a variety of sources. For example, NIOSH and OSHA provide data on occupational exposure and CDC provides data for a variety of exposure sources. The scientific literature and various reports provide a wide variety of animal and human data. In addition, data are obtained on types and amounts of foods eaten, concentration of food contaminants and human health effects.

Food Consumption

Some principal sources of food consumption data include: the U.S. Department of Agriculture (USDA) Nationwide Food Consumption Survey (NFCS); the National Health and Nutrition Examination Survey (NHANES); and the Market Research Corporation of America (MRCA) survey data. Sources of food composition data include: USDA: Handbook 456, Handbook 8, National Nutrient Data Bank file; NHANES: Model Gram and Nutrient Composition files; and MRCA: Lead

content of foods and methylxanthine content of foods (based on data from Total Diet Study and industry). Additional information on some of these sources is provided in the following discussions.

NATIONWIDE FOOD CONSUMPTION SURVEY. results of the Nationwide Food Consumption Survey (NFCS), 1977–78, provide data on the kinds and quantities of foods and nutritive values of diets ingested by men, women and children of different ages classified by various household characteristics. Dietary information obtained for individuals included the kind and amount of each food eaten; the time the food was eaten, the type of service, and the cost. Individuals were also asked if the day's intake was typical and if they were on a special diet, were vegetarians, or took vitamins, minerals, or other supplements. Reported are average intakes per day and per eating occasion for foods most commonly eaten by individuals who participated for 3 days in the NFCS conducted by the U.S. Department of Agriculture.

NATIONAL HEALTH AND NUTRITION EXAMINATION SURVEY. Data were collected for NHANES II through response to questionnaires on medical history, food consumption, and health-related behavior. Data also were collected through direct medical examination. NHANES II was conducted on a nationwide probability sample of approximately 20,000 persons ages 6 months to 74 years from the civilian non-institutionalized population of the United States. Food consumption data is of the form of quantities of food consumed and frequency of eating in a 24-hr recall period.

MARKET RESEARCH CORPORATION OF AMERICA SURVEYS. One of the largest proprietary surveys of food intake is one conducted by the Market Research Corporation of America (MRCA). This corporation conducts the Menu Census survey which is designed to collect detailed information for one year on the ultimate disposition of all food products in terms of their consumption by individuals or their use in preparation of other dishes consumed by the individuals. Only information on frequency of eating is reported; that is, quantities are not included. Each Menu Census consists of 4000 households. Each household reports all food preparation and consumption at home and away from home for 14 consecutive days in daily diaries. Brands and food packaging materials are also described. The sample is balanced within each quarter as closely as possible to the U.S. Census by various demographic characteristics, including region, household size, household income, and other socioeconomic variables. Pertinent comparisons among these three data sources are given in Table A-1.

The Total Diet Studies monitor levels of chemical contaminants and essential minerals in table ready food and consider dietary intake of these substances by various age-sex groups. The Total Diet Study is an annual program which involves the purchase of approximately 200 foods in grocery stores across the United States and the analysis of these foods for essential minerals, toxic elements, radionuclides, industrial chemicals and pes-

ticides. The food list and diets, beginning in April 1982, were revised to reflect the present food supply and the current food consumption habits. The number of agesex groups was increased to eight (infants, young children, male and female teenagers, male and female adults, and male and female older persons) in order to expand the coverage of the U.S. population. Foods were analyzed individually, rather than in commodity groups, to obtain information on the contribution of specific foods to the daily intake of minerals and contaminants.

The diets are based on data from the 1977-78 USDA NFCS and the NHANES II carried out by the National Center for Health Statistics in 1976-80. Two hundred and thirty-four foods are collected from four geographic regions (northeast, south, west, and north central) and analyzed by the Kansas City Field Office Laboratory for 11 essential minerals and more than 120 chemical contaminants. The essential minerals analyzed for are: copper, iron, magnesium, manganese, potassium, phosphorus, calcium, sodium, iodine, selenium, and zinc. The daily intake by weight of these 234 foods has been weighted to represent 100% of the usual diet for the eight age-sex groups. These estimated food intakes are used to assess daily contaminant and mineral intake.

Food Additives

There are two major data bases. The National Academy of Science (NAS) Surveys provide the agency with a data base for the concentration of additives used in food. To ensure confidentiality, the NAS maintains the raw data from the surveys. These data have been obtained from mailed questionnaires that have been sent to over 2000 firms with approximately 1100 responding. The values obtained from the NAS are used as the substance concentration value in estimating intakes.

The surveys approach the question of usage of food additives from two directions: how much poundage is used annually and what is the level of use in each of the firms' products.

The Bureau of Foods' database for food additives comprises food additive safety information for substances added directly to food in the United States. Nearly 1600 items (over 1300 synthetic flavoring substances, nearly 300 direct additives, and some 70 color additives and color diluents) supply data on human exposure levels, chemical structure, and toxicological parameters. The computer database is searchable by software programs that permit sorting of the data on many parameters, including lowest effect levels, test animal species, toxicological effects and organ sites, as well as exposure and chemical structure categories.

This database comprises one module of the Bureau's SIREN (Scientific Information Retrieval and Exchange Network) system, the other two being the food additive information system (FAIS) and the new animal drug information system (NADIS) containing over 1/2 million indexed records across two FDA bureaus.

Census Data

Census data on the count and characteristics of the population are obtained from the Bureau of the Census. The agency has census files for 1960 and 1970 for the

Characteristic	USDA NFCS 1977-78	NHANES II 1976-80	MCRA Menu Census VI 1977-78
Sample size	30,770	20,332	11,150
Length of dietary survey	3 days	1 day	14 days
Dietary methodology	24-hr recall (one-day) records (two days)	24-hr/recall	Daily diary
Collection methodology	Recall: interviewers record: self-administered	Interviewer	Self-administered via mail
Reporter	Individual subject	Individual subject	Main food preparer
Place of recording	Home	Mobile exam center	Home
Food quantities	Yes	Yes	No
Drinking water	Yes	No	No
Processing/packaging information	No	No	Yes
Other surveys in this series	1965–66	1971–74	1957–75

Table A-1. National food consumption surveys.

total population counts for each U.S. county or county equivalent, such as independent city, and within each county equivalent by sex, race, and 5-year age interval. These data are used in calculating rates in the population. Population estimates for the years 1971–1978 are available.

Fish Intake Data

Fish intake data are important to FDA in regulating food safety and has been used in evaluating the hazard of mercury, dioxins, and other contaminants of fish. The sources of national fish production data are the National Marine Fisheries Service (NMFS) and the USDA. NMFS calculates an annual estimate of the per capita consumption of seafood from data on the total catch of commercial seafood. The estimate does not include commercial fresh water fish, i.e., recreationally caught fish sold at roadside stands. The NMFS also supplies a database on the mercury content found in fish. The USDA calculates annual per capita fish intake by adding an estimate of recreationally caught fish and the other type of data to the commercial seafood production to calculate the total fish consumption.

Natural Toxicants

A computerized natural toxicants literature file is maintained which contains reprints of articles relative to the occurrence and health effects of mycotoxins, including plant and marine toxins. In addition, information is maintained on the level of mycotoxins in various commodities in the U.S.

Elemental Analyses

Data from the U.S. Geological Survey on elemental analysis of soils, waters, and vegetation provide background information on geographic differences in elements likely to be in the human diet.

Special Studies

Consumer interview surveys are conducted periodically on topics of special interest, which may grow out of immediate or potential health risks to the public or the need for information not currently available in areas of regulatory concern to the agency. The studies are

designed internally, executed under extramural contract and the data are delivered on computer tape for internal tabulation and analysis.

Literature

A bibliography file of articles written by FDA employees is maintained. This file together with a computerized cross index contains articles on such subjects as analytical methods and food contaminant levels.

Appendix B: Exposure Assessment at EPA

Introduction

An exposure assessment is that process which seeks to define the quantity of a chemical which comes into contact, or may come into contact, with human populations. This integral portion of the risk assessment is a growing technical speciality whose complexity has long been noted but has only recently been addressed using the sophisticated scientific tools available (588,589).

The process involves consideration of the magnitude, frequency and duration of encounters between individuals and the chemical in question. In some instances (e.g., prescribed dosing with drugs), the information is rather precisely known. In the case of exposure to chemicals of environmental concern, however, the assessment process is often associated with a dearth of data and assumptions/approximations must be made to fill in the gaps. For example, while some data may be available on the "direct exposure" from the emissionsof a manufacturing plant, estimates of the "indirect exposure" resulting from potential bioaccumulation in the food chain may be less precisely known.

In recent years, data bases have been compiled, computer models developed, laboratory/field measurement taken and "standard operating procedures" adopted (1,590,591) that have narrowed the uncertainty associated with some of these estimates.

Defining the Exposure

Before beginning an exposure assessment, it is nec-

essary to define clearly the product needed in terms of purpose, breadth, depth and approach. This conceptual scheme is being developed in detail by the EPA (586,587).

Depending on the answers provided to these questions, different resources are available to construct the exposure assessment. Some of the more important resources are discussed below.

Monitoring

In many cases, site-specific monitoring is performed in order to generate qualitative or quantitative data for an exposure assessment, e.g., more than 5000 water, soil, and sediments samples have been analyzed for the presence of 2,3,7,8-tetrachlorodibenzo-p-dioxin in the state of Missouri; groundwater and vertical soil cores have been analyzed for ethylene dibromide in Hawaii, California, and Georgia; and stack emissions have been sampled for heavy metals.

In addition to site-specific information, qualitative and quantitative exposure information is available from the various monitoring networks which have operated over the years to collect data on chemicals in different environmental media. For example, the Mussel Watch Program (612,618) has analyzed shellfish in the U.S. estu-arine waters for a variety of pollutants. Shellfish are filter feeders and are generally efficient bioconcentrators of water contaminants; the residues in these organisms serve as sensitive markers for upstream pollution of waters which feed the estuaries.

In addition, great quantities of analytical chemical information on U.S. waters can be accessed through the computer data base, STORET. These data represent information gathered over the past years from every major watershed in the U.S.

Increasingly, the problem of groundwater contamination is of concern. Little systematically collected information on this environmental compartment is available at this time, although more effort is being expended to identify and quantitate levels of certain chemicals such as aldicarb and ethylene dibromide at selected sites.

EPA has gathered information on atmospheric pollutants for many years. As part of the national efforts to the quality of the ambient air, innumerable measurements have been made at various times and locations on certain pollutants, namely carbon monoxide, sulfur oxides, nitrogen oxides, particulates, hydrocarbons, and photochemical oxidants. Much of this information is available in computer files maintained by EPA, Office of Air Programs, in Research Triangle Park, NC.

In a broader perspective of "monitoring"—where the chemical is in the environment—EPA's Office of Toxic Substances maintains and has access to files which identify chemical substances which are manufactured in or imported into the U.S. Some types of geototal production figures are generally available, along with graphic information on the location of production sites. This information is useful in generating a "material balance" of a chemical (a complete accounting of sources and dis-

position of a chemical in the environment).

EPA often relies on data generated by other Federal agencies (e.g., FDA, NIOSH and the Department of Labor) to estimate the exposures likely to be encountered from eating food or being in the workplace.

Increasingly, interest is expressed in determining more direct mreasurement of human exposure to environmental pollutants. For years, EPA has maintained the National Human Adipose Tissue Network which collects adipose tissue from selected cadavers in such a way as to obtain a rigorous statistical sample of the U.S. population. These tissue samples are analyzed for a variety of organic chemical residues so that trends and potential problem chemicals can be identified. Further, the Agency publishes a summary of studies, on residues in humans, which have appeared in the scientific literature on residues in humans.

An ambitious program underway at the EPA, called the Total Exposure Assessment Methodology (TEAM) study (625-630), combines personal monitor results (593) with analyses of food, beverages, water, human blood plasma and serum, urine, breath and mothers' milk to characterize the individual exposure of large test populations to a variety of organic chemicals over extended periods of time. Such data also will be useful in determining what portion of a particular chemical is actually absorbed by the body and how absorption varies from one individual to another at different exposure levels. The actual dose of a chemical which determines the health outcome from the exposure is often different from the apparent exposure level (753). The goal of the TEAM program is to develop an individual exposure monitoring methodology for use in estimating population exposures, including analytical chemical techniques of proven accuracy and reliability, tools for conveniently collecting detailed information on the activities of individuals on a 24-hr a day basis, and tools for managing, analyzing, and interpreting the data produced by such studies.

In sum, the Agency has broad environmental monitoring data on a number of chemical substances in various media. The information too often is not sufficient in depth, detail, and validation to provide precise data in a given exposure situation. Further uncertainties are introduced when attempts are made to extrapolate these data to absorbed dose. Therefore, while the Agency is indeed building an exposure data base, the range of uncertainty surrounding any exposure situation can be significant.

Modeling

The questions of "indirect exposure" often center on the transformation, transport, and ultimate fate of a chemical once it is released into the environment. While direct monitoring can provide some data in this regard, important detailed questions of chemical kinetics, distribution between media, potential uptake by biological organisms, and the like cannot easily be derived from such information.

Two approaches to filling this gap are through the use of microcosms and through the use of mathematical

modeling.

MICROCOSMS. A microcosm is a recreation of a portion of the environment in the laboratory—a physical model of nature. Ideally, the microcosm faithfully mimics the processes occurring in nature. By adding a chemical to such a system, its subsequent partitioning amongst the components of the environment, together with the associated chemical and biological conversion, can be studied in a single integrated experiment.

Marine, freshwater, terrestrial, atmospheric, and subsurface or aquifer microcosms are available and have demonstrated utility. Much of our knowledge of the chemistry which occurs in the atmosphere has been generated using smog chambers (639,671), i.e., atmosphere microcosms. Marine microcosms are now being used to understand the complex processes which affect the fate of a chemical in the ocean where it can be transformed by the benthate community, and eventually concentrated up the food chain leading to human food (672-675). Terrestrial microcosms are being used to define more precisely the fate of a chemical on soil-plantwater systems (676-678). Subsurface or aquifer microcosms have been created using special drilling techniques which allow a column of material to be removed from an aquifer in an undisturbed and uncontaminated state (689). Such columns are then used to study the chemical and biological conversions underground.

One of the continuing questions raised about microcosms is how well do their results actually mimic what would occur in the "real" environment itself. In recent years, increased effort has been aimed at validating these systems and gaining an appreciation of the limitations of their data. At this time, however, the risk assessor must recognize this inherent potential source of error.

MATHEMATICAL MODELING. One of the most frequently used tools in exposure assessment is mathematical modeling. Most such models begin as an effort to describe theoretically the phenomena observed in the laboratory or field. Large monitoring data bases are used to construct mathematical relationships between variables of concern (e.g., meteorological parameters and the temporal/spacial distribution of chemicals emitted from a source). These relationships constitute the model which, because of the inherent complexity, often requires a computer for its use. Models are predictive tools used when direct measurement of exposure would be prohibitively time consuming and/or expensive and when the relationship between the sources of a chemical and the eventual exposure is complex.

Given the wide variety of possible situations requiring exposure assessments, it is not surprising that many different models have been developed. A recent catalog of the models available for use in regulatory decision-making (640) identified 156 models, most of which can be applied to exposure assessments (641,642). A description of the general features of some advanced models is informative (644).

Ambient air modeling has reached the highest level of complexity with a capability for considering numerous point, line, and area emission sources simultaneously (645-647). Several models (643) predict the complex chemical conversions occurring in air during transport of pollutants. One model allows prediction of the half-life and reaction products for an organic chemical in air (648). Models currently under development address the problems of predicting exposure on a regional basis (649) and in areas with rough terrain (650). The Handbook for Performing Exposure Assessments (754) lists a number of routinely used air models available through NTIS.

Models are also available for the indoor air environment. The breadth and complexity of such modeling varies in proportion to the complexity of the environment being modeled. Even a single room or residence model may become quite complex when considerations of air filter efficiencies, particulate agglomeration/plating-out, nonspecific heterogeneous decay of reactive pollutants, multiple sources, sinks and varying sourcestrengths are encountered. Additional difficulties of assessing infiltration rates, and the partitioning of ventilation rates among discrete volumes in a building have led to the use of compartmented models (651). The contribution of varying levels of ambient pollutants to the indoor environment has been evaluated by a time-segmented model which has undergone limited field validation (693).

As a result of the Great Lakes research program (652-656) and research into the environmental fate of pesticides (657,658), models are available which describe the movement of chemicals in fresh water aquatic environments. Studies on the ocean disposal of municipal wastes have led to models for chemicals in coastal waters (659-661). Recent work on terrestrial microcosms has provided input data for a terrestrial model (662) which includes consideration of plant uptake. A number of models are available which describe leaching and runoff of chemicals to streams (640). The newest areas of modeling are ground water (619,663-667), human microenvironments and activity patterns (668-670), food consumption (576), and multimedia environmental partitioning (755,756).

As models increase in complexity, the ability to predict exposure generally becomes more accurate. This additional accuracy, however, is purchased at the price of additional input data (694). The choice of models available for a particular assessment may be reduced by a limited data base.

In general, models are most helpful when rough estimates of exposure are required. Even though the level of sophistication of a particular model may be high, the predictions are still crude because any model is an oversimplified description of the natural phenomenon it depicts. There is a need for more research to validate most models, since the accuracy of the exposure estimates made by models can be no better than the quality of the input data. The use of models does not compensate for poor input data, but in certain instances may obscure the true uncertainty inherent in the exposure predictions. In sum, the modeling as well as in monitoring the

Agency can obtain valuable information. At the same time it needs to be recognized that this information might be accompanied by potential significant uncertainties. Nevertheless, the use of scientific judgment in interpreting results is essential.

Appendix C: Chemical Exposure Assessment for Consumer Products

Assessing human exposure to toxic chemicals from consumer products is a vital part of CPSC's chemical hazard work and is the element which most distinguishes CPSC's efforts from the activities of other Federal agencies. The range of uses of chemical compounds in consumer products is great and the ways that consumers utilize a single product may vary considerably. For example, the techniques required for assessing exposure to a gas that may be emitted from building materials into home air and for a dye or other agent contained in cloth that may contact the skin have little in common. Most experience in exposure assessment from products has been gained over the past few years. Because of this, CPSC approaches exposure assessments on a case-by-case basis with each study in general requiring original experimental or theoretical work. Nonetheless, the general activity of exposure assessment can be described as a four step process:

- (1) Determine the fact of the release of a compound from products and estimate the quantity of release. The degree of difficulty of this step can vary greatly. For example, it is straightforward if the compound of interest is contained in an aerosol product. It may require extensive experimental study if the compound is contained in a plastic matrix that is subjected to a variety of uses. More often than not in CPSC's experience, experimental studies are required. The presence of a compound in a consumer product does not imply that release actually occurs.
- (2) Relate the quantity of release with the level of human exposure (to the outside of the body) from use of the products. A series of analyses are required for this step. First, use patterns for the products must be determined; both the most typical use patterns and those that could lead to lower or higher degrees of exposure must be considered. Experience of staff, information from industry, and consumer surveys can provide this information. Then, for each specific use, the data on chemical release must be used to estimate the actual amount of contact the user has with the chemical. For a chemical emitted into the home air, data on release rate, use patterns, home volume, and air exchange rate can be combined in a mathematical model to estimate the average and peak levels in the air. On the other hand, an experimental study could be conducted to measure actual air levels from a certain product use. As another example the quantity of a liquid that comes into contact with skin can be measured.
- (3) Determine the entry of the material into the body. This step can be straightforward if the toxicity information on a chemical was obtained by a route of ex-

posure comparable with consumer use. On the other hand, particularly in the case of dermal exposures, data on entry of a compound into the body are frequently lacking. In that case experimental studies, such as painting the skin of an experimental animal with a radiolabeled quantity of the compound of interest, may be conducted.

(4) Combine the data in steps (1) through (3) for a complete estimate of human exposure to a compound. This estimate can then be combined with health hazard information (toxicity) in evaluating human risks; it can serve as the basis required for a mathematical estimation of risk.

These steps are presented as one sequence, however, exposure estimation is often an interactive procedure. One uses available, limited information to make preliminary estimates of risk that may (if sufficiently conservative) eliminate concern about an exposure or may be successively refined by gathering data where it is most needed.

In addition, in certain studies, some of the above steps may be combined rather than considered separately. For example, a chemically treated cloth may be placed in contact with the skin of an experimental animal and the amount of uptake of the chemical by the animal measured. This experiment provides information that could separately be obtained by measuring both quantities released from the cloth and rates of penetration of the chemical through the skin. In another example, personal air monitoring can take into account both release rates from products and patterns of product use.

Exposure assessment for chemicals in consumer products is a developing science which makes innovative use of standard physical and chemical data, demands development of new experimental procedures, and combines relatively "hard" experimental data with "soft" data on widely varying human behavior patterns. While each exposure assessment is now a new undertaking, certain basic techniques (such as air level modeling) are finding repeated application. In the next several years the knowledge now being gained should lead to more readily standardized and more easily performed exposure assessments.

REFERENCES

- Managing the Process: Risk Assessment in the Federal Government. National Academy of Sciences, Washington DC (1983).
- Zimmerman, M. R. An experimental study of mummification pertinent to the antiquity of cancer. Cancer 40: 1358–1362 (1977).
- Doll, R., and Peto, R. The causes of cancer: Quantitative estimates of avoidable risks of cancer in the United States today.
 J. Natl. Cancer Inst. 66: 1193-1308 (1981).
- 4. Leven, M. L., Haenszel, W., Carroll, B. E., Gerhardt, P. R., Handy, V. H., and Ingraham, II, S. C. Cancer incidence in urban and rural areas of New York State. J. Natl. Cancer Inst. 24: 1243–1257 (1960).
- Ramazzini, B. Diseases of Workers. 1700, p. 191. Translation of the Latin text of 1713 by Wilmer Cage Wright, University of Chicago Press, Chicago, IL, 1940.
- 6. Hill, J. Cautions Against the Immoderate Use of Snuff. Baldwin and Jackson, London, 1761, pp. 30–31.
- 7. Pott, P. Chirurgical Observations Relative to the Cataract, the

- Polypus of the Nose, the Cancer of the Scrotum, the Different Kinds of Ruptures, and the Mortification of the Toes and Feet. Hawes, Clarke and Collins, London, 1775, pp. 63–68.
- 8. Weismann, A. The duration of life. In: Essays Upon Heredity and Kindred Biological Problems, Vol. 2. E. B. Poulton and A. E. Shipley (authorized translation), Clarendon Press, Oxford, 1882, pp. 1–222.
- Rehn, L. Blasengeschwülste bei Fuchsin-Arbeitern. Arch. Klin. Chir., 50: 588-600 (1895).
- Unna, P. G. Die Histopathologie der Hautkrankheiten. A. Hirschwald, Berlin, 1984, p. 725.
- 11. Dubreuilh, W. Des Hyperkeratoses Circonscrites (1). Ann. Dermatol. Syphilig. 3rd Series, 7: 1158–1204 (1896).
- 12. Mulvihill, J. J. Congenital and Genetic Diseases. In: Persons at High Risk of Cancer: An Approach to Cancer Etiology and Control (J. F. Fraumeni, Jr., Ed.), Academic Press, New York, 1975, pp. 3–37.
- 13. Committee on Diet, Nutrition and Cancer/Assembly of Life Sciences, National Research Council. Diet, Nutrition and Cancer. National Academy of Sciences, Washington, DC, 1982.
- Doll, R. Strategy for detection of cancer hazards to man. Nature 265: 589–596 (1977).
- Logani, M. K., Austin, W. A., and Davies, R. E. Photooxygenation of 7,12-dimethylbenz(a)anthracene. Tetrahedron Letters 29: 2467-2470 (1977).
- 16. Richards, R. J., George, G., Hunt, J., and Tetley, T. D. The relationship between the haemolytic potential of certain particulates and their reactivity at the lung surface "in vivo". In: The In Vitro Effects of Mineral Dusts (R. C. Brown, M. Chamberlain, R. Davies, and I. P. Gormley, Eds.), Academic Press, New York, 1980, pp. 323–332.
- Cerniglia, C. E., Freeman, J. P., Franklin, W., and Pack, L. D. Metabolism of benzidine and benzidine-congener based dyes by human, monkey and rat intestinal bacteria. Biochem. Biophys. Res. Comm. 107: 1224–1229 (1982).
- Vesell, E. S. On the significance of host factors that affect drug disposition. Clin. Pharmacol. Therap. 31: 1-7 (1982).
- Morgan, R. W., and Hoffmann, G. R. Cycasin and its mutagenic metabolites. Mutat. Res. 114: 19-58 (1983).
- Goldman, P. Biochemical pharmacology of the intestinal flora. Ann. Rev. Pharmacol. Toxicol. 18: 523-539 (1978).
- Lynn, R. K., Donielson, D. W., Ilias, A. M., Kennish, J. M., Wong, K., and Matthews, H. B. Metabolism of bisazobiphenyl dyes derived from benzidine, 3,3'-dimethylbenzidine or 3,3'-dimethoxybenzidine to carcinogenic aromatic amines in the dog and rat. Toxicol. Appl. Pharmacol. 56: 248-258 (1980).
- Nony, C. R., Bowman, M. C., Cairns, T., Lowry, L. K., and Tolos, W. P. Metabolism studies of an azo dye and pigment in the hamster based on analysis of the urine for potentially carcinogenic aromatic amine metabolites. J. Anal. Toxicol. 4: 132– 140 (1980).
- 23. Martin, C. N., and Kennelly, J. C. Rat liver microsomal azore-ductase activity on four azo dyes derived from benzidine, 3,3′-dimethylbenzidine or 3,3′-dimethoxybenzidine. Carcinogenesis 2: 307–312 (1981).
- 24. Cerniglia, C. E., Freeman, J. P., Franklin, W., and Pack, L. D. Metabolism of azo dyes derived from benzidine, 3,3'-dimethylbenzidine and 3,3'-dimethoxybenzidine to potentially carcinogenic aromatic amines by intestinal bacteria. Carcinogenesis 3: 1255–1260 (1982).
- 25. Goldwater, L. J., Rosso, A. J., and Kleinfeld, M. Bladder tumors in a coal tar dye plant. Arch. Environ. Health 11: 814–817 (1965).
- DeBethizy, J. D., Sherrill, J. M., Kickent, D. E., and Hamm, T. E., Jr. Effects of pectin-containing diets on the hepatic macromolecular covalent binding of 2,6-dinitro-[3H]toluene in Fischer 344 rats. Toxicol. Appl. Pharm. 69: 369–376 (1983).
- 27. Klein, G. Viral Oncology. Raven Press, New York, 1980.
- 28. Upton, A. C. Physical carcinogenesis: radiation—history and sources. In: Cancer: A Comprehensive Treatise. Vol. 1. Etiology: Chemical and Physical Carcinogenesis (F. F. Becker, Ed.), 2nd Ed., Plenum Press, New York, 1982, pp. 551–568.
- 29. Brand, K. G. Cancer associated with asbestosis, schistosomiasis, foreign bodies and scars. In: Cancer: A Comprehensive Treatise.

- Vol. 1. Etiology: Chemical and Physical Carcinogenesis (F. F. Becker, Ed.), 2nd Ed., Plenum Press, New York, 1982, pp. 661–692.
- Searle, C. E. (ed.). Chemical Carcinogens (American Chemical Society Monograph, No. 173), American Chemical Society, Washington, DC, 1976.
- Schoental, R. Carcinogens in plants and microorganisms. In: Chemical Carcinogens (American Chemical Society Monograph, No. 173) (C. E. Searle, Ed.), American Chemical Society, Washington, DC, 1976, pp. 626–689.
- Grasso, P., O'Hare, C. Carcinogens in food. In: Chemical Carcinogens (American Chemical Society Monograph, No. 173) (C. E. Searle, Ed.), American Chemical Society, Washington, DC, 1976, pp. 701–729.
- 33. Ames, B. Dietary carcinogens and anticarcinogens. Science 221: 1256-1263 (1983).
- Parkes, H.G. The epidemiology of the aromatic amine cancers.
 In: Chemical Carcinogens (American Chemical Society Monograph, No. 173) (C. E. Searle, Ed.), American Chemical Society, Washington, DC, 1976, pp. 462–480.
- Foucar, K., McKenna, R. W., Bloomfield, C. D., Bowers, T. K., and Brunning, R. D. Therapy-related leukemia. A panmyelosis. Cancer 43: 1285–1296 (1979).
- 36. Higginson, J. Cancer and environment. Science, 205: 1363-1366 (1979).
- 37. Miller, E. C., and Miller, J. A. The metabolism of chemical carcinogens to reactive electrophiles and their possible mechanisms of action in carcinogenesis. In: Chemical Carcinogens (American Chemical Society Monograph, No. 173) (C. E. Searle, Ed.), American Chemical Society, Washington, DC, 1976, pp. 737–762.
- 38. Jennette, K. W. The role of metals in carcinogenesis: biochemistry and metabolism. Environ. Health Perspect. 40: 233-252 (1981).
- 39. Young, J. F., and Kadlubar, F. F. A pharmacokinetic model to predict exposure of the bladder epithelium to urinary N-hydroxyarylamine carcinogens as a function of urine pH, voiding interval, and resorption. Drug Metab. Disp. 10: 641-648 (1982).
- 40. Paxton, M. B., Siegel, D. M., and Nisbett, I. C. T. Comments on the pharmacokinetic dose-response model of Gehring and Blau and its applicability to carcinogenic risk assessment. Report for Natural Resource Defense Council, by Clement Associates, Inc., Arlington, VA, February 17 (1984).
- 41. Dahl, A. R., Hadley, W. M., Hahn, F. F., Benson, J. M., and McClellan, R. O. Cytochrome P-450-dependent monooxygenases in olfactory epithelium of dogs: possible role in tumorigenicity. Science 216: 57-59 (1982).
- Williams, R. T. Detoxification Mechanisms, 2nd ed. Chapman and Hall, London, 1959.
- 43. Powell, G. M., and Curtis, C. G. Sites of sulphation and the fates of sulphate esters. In: Conjugation Reactions in Drug Biotransformation (A. Aitio, Ed.), North Holland/Elsevier, Amesterdam, 1978, pp. 409-416.
- 44. Sims, P., and Grover, P. L. Epoxides in polycyclic aromatic hydrocarbon metabolism and carcinogenesis. Adv. Cancer Res. 20: 165–274 (1974).
- Lu, A. Y. H. Multiplicity of liver drug metabolizing enzymes. Drug Metab. Rev., 10: 187–208 (1979).
- Caldwell, J. Conjugation reactions in foreign-compound metabolism: definition, consequences, and species variations. Drug Metab. Rev., 13: 745-777 (1982).
- Radomski, J. L. The primary aromatic amines: their biological properties and structure-activity relationships. Ann. Rev. Pharmacol. Toxicol., 19: 129–157 (1979).
- Jerina, D. M., Yagi, H., Thakker, D. R., Karle, J. M., Mah, H. D., Boyd, D. R., Gadaginamath, G., Wood, A. W., Buening, M., Chang, R. L., Levin, W., and Conney, A. H. Stereoselective metabolic activation of polycyclic aromatic hydrocarbons. Adv. Pharmacol. Therap. 9: 53–62 (1979).
- 49. Slaga, T. J., Bracken, W. J., Gleason, G., Levin, W., Yagi, H., Jerina, D. M., and Conney, A. H. Marked differences in the skin tumor-initiating activities of the optical enantiomers of the diastereomeric benzo(a)pyrene 7,8-diol-9,10-epoxides. Cancer Res.,

- 39: 67-71 (1979).
- Hutson, D. H. Mechanisms of biotransformation. In: Foreign Compound Metabolism in Mammals, Vol. 4, Thanet Press, Margate England, Vol. 4, 1976, pp. 259–346.
- 51. Conney, A. H. Induction of microsomal enzymes by foreign chemicals and carcinogenesis by polycyclic aromatic hydrocarbons. Cancer Res. 42: 4875–4917 (1982).
- 52. Yang, S. K., McCourt, D. W., Roller, P. P., and Gelboin, H. V. Enzymatic conversion of benzo(a)pyrene leading predominantly to the diol-epoxide r-7,t-8-dihydroxy-t-9,10-oxy-7,8,9,10-tetrahydrobenzo(a)pyrene through a single enantiomer of r-7,t-8-dihydroxy-7,8-dihydrobenzo(a)pyrene. Proc. Natl. Acad. Sci. (U.S.) 73: 2594-2598 (1976).
- 53. Hart, R. W., Fu, P. P., and Chang, M. J. W. Comparative removal of polycyclic aromatic hydrocarbon-DNA adducts in vivo; In: Sixth International Symposium on Polynuclear Aromatic Hydrocarbons: Physical and Biological Chemistry (W. M. Cooke, A. J. Dennis, and G. L. Fisher, Eds.), Battelle Press, Columbus, Ohio, 1982, pp. 39–72.
- Wattenberg, L. W. Inhibitors of chemical carcinogens. J. Environ. Pathol. Toxicol. 3: 35-52 (1980).
- 55. Thakker, D. R., Levin, W., Buening, M., Yagi, H., Lehr, R. E., Wood, A. W., Conney, A. H., and Jerina, D. M. Species-specific enhancement by 7,8-benzoflavone of hepatic microsomal metabolism of benzo(e)pyrene 9,10-dihydrodiol to bay-region diol-epoxide. Cancer Res. 41: 1389-1396 (1981).
- Tateishi, N., Higashi, T., Shinya, S., Naruse, A., and Sakamoto,
 Y. Studies on the regulation of glutathione level in rat liver. J.
 Biochem. (Tokyo) 75: 93-103 (1974).
- 57. Lam, L. K. T., Sparnins, V. L., Hochalter, J. B., and Wattenberg, L. W. Effects of 2- and 3-tert-butyl-4-hydroxyanisole on glutathione S-transferase and epoxide hydrolase activities and sulfhydryl levels in liver and forestomach of mice. Cancer Res. 41: 3940-3943 (1981).
- Hinson, J. A., Monks, T. J., Hong, M., Hight, R. J., and Pohl,
 C. R. 3-(glutathion-S-yl)acetaminophen: a biliary metabolite of acetaminophen. Drug Metab. Disp. 10: 47-51 (1982).
- Murasaki, G., Zenser, T. Z., Davis, B. B., and Cohen, S. M. Inhibition by aspirin of N-[4-(5-nitro-2-furyl)-2-thiazo-lyl]formamide-induced bladder carcinogenesis and enhancement of forestomach and carcinogenesis. Carcinogenesis 5: 53-55 (1984).
- Hadley, W. M., and Dahl, A. R. Cytochrome P-450-dependent monooxygenase activity in the nasal membrane of 6 species. Drug Metab. Disp. 11: 275-276 (1983).
- Hart, R. W., and Turturro, A. Species longevity as an indicator for extrapolation for toxicity data among placental mammals. J. Am. College Toxicol. 2(3): 235-243 (1983).
- Miller, E. C. Some current perspectives on chemical carcinogenesis in humans and experimental animals: presidental address. Cancer Res. 38: 1479-1496 (1978).
- 63. Ketterer, B. Interactions between carcinogens and proteins. Brit. Med. Bull. 36: 71-78 (1980).
- Carroll, K. K. Lipids and carcinogenesis. J. Environ. Pathol. Toxicol. 3: 253–271 (1980).
- Weinberg, R. A. Oncogenes of spontaneous and chemically induced tumors. Adv. Cancer Res. 36: 149-163 (1982).
- Poirier, L. A., and deSerres, F. J. Initial National Cancer Institute studies on mutagenesis as a prescreen for chemical carcinogens: an appraisal. J. Natl. Cancer Inst. 62: 919-926 (1979).
- Bridges, B. A. Some DNA-repair-deficient human syndromes and their implications for human health. Proc. Roy. Soc. (London) B212: 263-278 (1981).
- 68. Maher, V. M., Rowan, L. A., Silinskas, K. C., Kateley, S. A., and McCormick, J. J. Frequency of UV-induced neoplastic transformation of diploid human fibroblasts is higher in xeroderma pigmentosum cells than in normal cells. Proc. Natl. Acad. Sci. (U.S.) 79: 2613–2617 (1982).
- 69. Kraemer, K. H., Lee, M. M., and Scotto, J. Diseases of environmental-genetic interaction: preliminary report on a retrospective study of neoplasia in 268 xeroderma pigmentosum patients. In: Environmental Mutagens and Carcinogens (T. Sugimura, S. Kondo, and H. Takebe, Eds.), A. R. Liss, New

- York, 1982, pp. 605-612.
- 70. Hart, R. W., Setlow, R. B., and Woodhead, A. D. Evidence that pyrimidine dimers in DNA can give rise to tumors. Proc. Natl. Acad. Sci. (U.S.) 74: 5574-5578 (1977).
- Zur Hansen, H. The role of viruses in human tumors. Adv. Cancer Res. 33: 77-107 (1980).
- Oskarsson, M., McClements, W. L., Blair, D. G., Maizel, J. V., and Vande Woude, G. G. Properties of a normal mouse cell DNA sequence (sarc) homologous to the src sequence of Moloney sarcoma virus. Science 207: 1222-1224 (1980).
- Ellis, R. W., DeFeo, D., Maryak, J. M., Young, H. A., Shih, T. Y., Chang, E. H., Lowy, D. R., and Scolnick, E. M. Dual evolutionary origin for the rat genetic sequences of Harvey murine sarcoma virus. J. Virol. 36: 408-420 (1980).
- Roussel, M., Saule, S., Lagrou, C., Rommens, C., Beug, H., Graf, T., and Stehelin, D. Three new types of viral oncogene of cellular origin specific for haematopoietic cell transformation. Nature 218: 452–455 (1979).
- 75. Sherr, C. J., Fedele, L. A., Donner, L., and Turek, L. P. Restriction endonuclease mapping of unintergrated proviral DNA of Snyder-Theilen feline sarcoma virus: localization of sarcomaspecific sequences. J. Virol. 32: 860–875 (1979).
- Spector, D. H., Varmus, H. E., and Bishop, J. M. Nucleotide sequences related to the transforming gene of avian sarcoma virus are present in DNA of uninfected vertebrates. Proc. Natl. Acad. Sci. (U.S.) 75: 4102-4106 (1978).
- 77. Witte, O. N., Rosenberg, N. E., and Baltimore, D. A normal cell protein cross-reactive to the major Abelson murine leukaemia virus gene product. Nature 281: 396–398 (1979).
- 78. Shih, C., Shilo, B. Z., Goldfarb, M. P., Dannenberg, A., and Weinberg, R. A. Passage of phenotypes of chemically transformed cells via transfection of DNA and chromatin. Proc. Natl. Acad. Sci. (U.S.) 76: 5714-5718 (1979).
- Krontiris, T. G., and Cooper, G. M. Transforming activity of human tumor DNAs. Proc. Natl. Acad. Sci. (U.S.) 78: 1181– 1184 (1981).
- 80. Shih, C., Padhy, L. C., Murray, M., and Weinberg, R. A. Transforming genes of carcinomas and neuroblastomas introduced into mouse fibroblasts. Nature 290: 261-264 (1981).
- 81. Perucho, M., Goldfarb, M., Shimizu, K., Lama, C., Fogh, J., and Wigler, M. Human tumor-derived cell lines contain common and different transforming genes. Cell 27: 467-476 (1981).
- 82. Murray, M. J., Shilo, B.-Z., Shi, C., Cowing, D., Hsu, H. W., and Weinberg, R. A. Three different human tumor cell lines contain different oncogenes. Cell 25: 355-361 (1981).
- 83. Goldfarb, M., Shimizu, K., Perucho, M., and Wigler, M. Isolation and preliminary characterization of a human transforming gene from T24 bladder carcinoma cells. Nature 296: 404-409 (1982)
- 84. Shih, C., and Weinberg, R. A. Isolation of a transforming sequence from a human bladder carcinoma cell line. Cell 29: 161-169 (1982).
- Pulciani, S., Santos, E., Lauver, A. V., Long, L. K., Robbins, K. C., and Barbacid, M. Oncogenes in humn tumor cell lines: Molecular cloning of a transforming gene from human bladder carcinoma cells. Proc. Natl. Acad. Sci. (U.S.) 79: 2845-2849 (1982).
- Weiss, R. A. (ed.). RNA Tumor Viruses: Molecular Biology of Tumor Viruses, 2nd Ed. Cold Spring Harbor, New York, 1982.
- Coffin, J. M., Varmus, H. E., Bishop, J. M., Essex, M., Hardy, W. D., Jr., Martin, G. S., Rosenberg, N. E., Scolnick, E. M., Weinberg, R. A., and Vogt, P. K. Proposal for naming host cell-derived inserts in retrovirus genomes. J. Virol. 40: 953-957 (1981).
- Shilo, B. Z., and Weinberg, R. A. DNA sequences homologous to vertebrate oncogenes are observed in *Drosophila melano*gaster. Proc. Natl. Acad. Sci. (U.S.) 78: 6789-6792 (1981).
- 89. Dalla-Favera, R., Gelmann, E. P., Gallo, R. C., and Wong-Staal, F. A human onc gene homologous to the transforming gene (v-sis) of simian sarcoma virus. Nature 292: 31-35 (1981).
- Trus, M. D., Sodroski, J. G., and Haseltine, W. A. Isolation and characterization of a human locus homologous to the transforming gene (v-fes) of feline sarcoma virus. J. Biol. Chem., 257:

- 2730-2733 (1982).
- 91. Dalla-Favera, R., Franchini, G., Martinotti, S., Wong-Staal, F., Gallo, R. C., and Croce, C. M. Chromosomal assignment of the human homologues of feline sarcoma virus and avian myeloblastosis virus onc genes. Proc. Natl. Acad. Sci. (U.S.) 79: 4714–4717 (1982).
- 92. Franchini, G., Gelmann, E. P., Dalla-Favera, R., Gallo, R. C., and Wong-Staal, F. Human gene (c-fes) related to the onc sequences of Snyder-Theilen feline sarcoma virus. Mol. Cell. Biol. 2: 1014–1019 (1982).
- Prakash, K., McBride, O. W., Swan, D. C., Devare, S. G., Tronick, S. R., and Aaronson, S. A. Molecular cloning and chromosomal mapping of a human locus related to the transforminbg gene of Moloney murine sarcoma virus. Proc. Natl. Acad. Sci. (U.S.) 79: 5210-5214 (1982).
- 94. Cooper, G. M. Activation of transforming genes in neoplasms. Brit. J. Cancer 50: 137-142 (1984).
- 95. Gonda, T. J., and Metcalf, D. Expression of *myb*, *myc*, and *fos* proto-oncogenes during the differentiation of a murine myeloid leukaemia. Nature 310: 249-251 (1984).
- Lachman, H. M., and Skoultchi, A. I. Expression of c-myc changes during differentiation of mouse erythroleukaemia cells. Nature 310: 592-594 (1984).
- 97. Barker, W. C., and Dayhoff, M. O. Viral src gene products are related to the catalytic chain of mammalian cAMP-dependent protein kinase. Proc. Natl. Acad. Sci. (U.S.) 79: 2836-2839 (1982).
- 98. Nishizuka, Y. The role of protein kinase C in cell surface signal transduction and tumour promotion. Nature 308: 693–698 (1984).
- Jacobs, S., Sahyoun, N. E., Saltiel, A. R., and Cuatrecasas, P. Phorbol esters stimulate the phosphorylation of receptors for insulin and somatomedin C. Proc. Natl. Acad. Sci. (U.S.) 80: 6211–6213 (1983).
- 100. McGrath, J. P., Capon, D. J., Goeddewl, D. V., and Levinson, A. D. Comparative biochemical properties of normal and activated human *ras* p21 protein. Nature 310: 644–649 (1984).
- Downward, J., Yarden, Y., Mays, E., Scrace, G., Totty, N., Stockwell, P., Ullrich, A., Schlessinger, J., and Waterfield, M. D. Close similarity of epidermal growth factor receptor and verb-B oncogene protein sequences. Nature 307: 521-530 (1984).
- 102. Naharro, G., Robbins, K. C., and Reddy, E. P. Gene product of v-fgr onc: hybrid protein containing a portion of actin and a tyrosine-specific protein kinase. Science 223: 63-66 (1984).
- 103. Doolittle, R. F., Hunkapiller, M. W., Hood, L. E., Devare, S. G., Robbins, K. C., Aaronson, S. A., and Antoniades, H. N. Simian sarcoma virus onc gene, v-sis, is derived from the gene (or genes) encoding a platelet-derived growth factor. Science 221: 275-277 (1983).
- 104. Amelin, H. A., Armelin, M. C. S., Kelly, K., Stewart, T., Leder, P., Cochran, B. H., and Stile, C. D. Functional role for c-myc in mitogenic response to platelet-derived growth factor. Nature 310: 655-660 (1984).
- 105. Persson, H., and Leder, P. Nuclear localization and DNA binding properties of a protein expressed by human c-myc oncogene. Science 25: 718-721 (1984).
- 106. Land, H., Parada, L. F., and Weinberg, R. A. Tumorigenic conversion of primary embryo fibroblasts requires at least two cooperating oncogenes. Nature 304: 596-602 (1983).
- 107. Spandidos, D. A., and Wilkie, N. M. Malignant transformation of early passage rodent cells by a single mutated human oncogene. Nature 310: 469-475 (1984).
- 108. Murphree, A. L., and Benedict, W. F. Retinoblastoma: Clues to human oncogenesis. Science 223: 1028-1033 (1984).
- 109. Debuire, B., Henry, C., Benaissa, M., Biserte, G., Claverie, J. M., Saule, S., Martin, P., and Stehelin, D. Sequencing the erbA gene of avian erythroblastosis virus reveals a new type of oncogene. Science 224: 1456-1459 (1984).
- 110. Owen, A. J., Pantazis, P., and Antoniades, H. N. Simian sarcoma virus-transformed cells secrete a mitogen identical to platelet-derived growth factor. Science 225: 52-56 (1984).
- 111. Sukumar, S., Notario, V., Martin-Zanca, D., and Barbacid, M. Induction of mammary carcinomas in rats by nitroso-methylurea involves malignant activation of H-ras-1 locus by single point

- mutations. Nature 306: 658-661 (1983).
- 112. Blair, D. G., Oskarsson, M., Wood, T. G., McClements, W. L., Fishinger, P. J., and Vande Woude, G. F. Activation of the transforming potential of a normal cell sequence: A molecular model for oncogenesis. Science 212: 941-943 (1981).
- 113. DeFeo, D., Gonda, M. A., Young, H. A., Chang, E. H., Lowy, D. R., Scolnick, E. M., and Ellis, R.W. Analysis of two divergent rat genomic clones homologous to the transforming gene of Harvey murine sarcoma virus. Proc. Natl. Acad. Sci. (U.S.) 78: 3328-3332 (1981).
- 114. Varmus, H. E. Form and function of retroviral proviruses. Science 216: 812–820 (1982).
- Klein, G. The role of gene dosage and genetic transpositions in carcinogenesis. Nature 294: 313-318 (1981).
- 116. Taub, R., Kirsch, I., Morton, C., Lenoir, G., Swan, D., Tronick, S., Aaronson, S., and Leder, P. Translocation of the c-myc gene into the immunoglobulin heavy chain locus in human Burkitt's lymphoma and murine plasmacytoma cells. Proc. Natl. Acad. Sci. (U.S.) in press.
- 117. Hayward, W. S., Neel, B. G., and Astrin, S. M. Activation of a cellular onc gene by promoter insertion in ALV-induced lymphoid leukosis. Nature 290: 475–480 (1981).
- 118. Payne, G. S., Bishop, J. M., and Varmus, H. E. Multiple arrangements of viral DNA and an activated host oncogene in bursal lymphomas. Nature 295: 209-214 (1982).
- 119. Bartram, C. R., de Klein, A., Hagemeijer, A., van Agthoven, T., van Kessel, A. G., Bootsma, D., Grosveld, G., Ferguson-Smith, M. A., Davies, T., Stone, M., Heisterkamp, N., Stephenson, J. R., and Groffen, J. Translocation of c-abl oncogene correlates with the presence of a Philadelphia chromosone in chronic myelocytic leukaemia. Nature 306: 277-280 (1983).
- Darnell, J. E., Jr. Variety in the level of gene control in eukaryotic cells. Nature 297: 365–371 (1982).
- 121. Blair, D. G., McClements, W. L., Oskarsson, M. K., Fischinger, P. J., and Vande Woude, G. F. Biological activity of cloned Moloney sarcoma virus DNA: terminally redundant sequences may enhance transformation efficiency. Proc. Natl. Acad. Sci. (U.S.) 77: 3504–3508 (1980).
- 122. Levinson, B., Khoury, G., Vande Woude, G. F., and Gruss, P. Activation of SV40 genome by 72-base pair tandem repeats of Moloney sarcoma virus. Nature 295: 568-572 (1982).
- 123. Burrows, P. D., Beck-Engeser, G. B., and Wabl, M. R. Immunoglobin heavy-chain class switching in a pre-B cell line is accompanied by DNA rearrangement. Nature 306: 243-246 (1983).
- 124. Hayday, A. C., Gillies, S. D., Saito, H., Wood, C., Wiman, K., Hayward, W. S., and Tonegawa, S. Activation of a translocated human c-myc gene by an enhancer in the immunoglobin heavychain locus. Nature 307: 334–340 (1984).
- 125. Lane, M., Sainten, A., and Cooper, G. M. Activation of related transforming genes in mouse and human mammary carcinomas. Proc. Natl. Acad. Sci. (U.S.) 78: 5185-5189 (1981).
- 126. Cooper, G. M., Okenquist, S., and Silverman, L. Transforming activity of DNA of chemically transformed and normal cells. Nature 284: 418–421 (1980).
- Cooper, G. M., and Neiman, P. E. Transforming genes of neoplasms induced by avian lymphoid leukosis viruses. Nature 287: 656–659 (1980).
- 128. Der, C. J., Krontiris, T.G., and Cooper, G. M. Transforming genes of human bladder and lung carcinoma cell lines are homologous to the ras genes of Harvey and Kirsten sarcoma viruses. Proc. Natl. Acad. Sci. (U.S.) 79: 3637-3640 (1982).
- 129. Parada, L. F., Tabin, C. J., Shih, C., and Weinberg, R. A. Human EJ bladder carcinoma oncogene is homologue of Harvey sarcoma virus ras gene. Nature 297: 474-478 (1982).
- 130. Santos, E., Tronick, S. R., Aaronson, S. A., Pulciani, S., and Barbacid, M. T24 human bladder carcinoma oncogene is an activated form of the normal human homologue of BALB- and Harvey-MSV transforming genes. Nature 298: 343-347 (1982).
- 131. Tabin, C. J., Bradley, S. M, Bargmann, C. I., Weinberg, R. A., Papageorge, A. G., Skolnick, E. M., Dhar, R., Lowy, D. R., and Chang, E. H. Mechanism of activation of a human oncogene. Nature 300: 143-149 (1982).

- 132. Reddy, E. P., Reynolds, R. K., Santos, E., and Barbacid, M. A point mutation is responsible for the acquisition of transforming properties by the T24 human bladder carcinoma cell oncogene. Nature 300: 149-152 (1982).
- Taparowsky, E., Suard, Y., Fasano, O., Shimizu, K., Goldfarb, M., and Wigler, M. Activation of the T24 bladder carcinoma transforming gene is linked to a single amino acid change. Nature 300: 762-765 (1982).
- 134. Poiesz, B. J., Ruscetti, F. W., Gazdar, A. F., Bunn, P. A., Minna, J. D., and Gallo, R. C. Detection and isolation of type C retrovirus particles from fresh and cultured lymphocytes of a patient with cutaneous T-cell lymphoma. Proc. Natl. Acad. Sci. (U.S.) 77: 7415-7419 (1980).
- 135. Poiesz, B. J., Ruscetti, F. W., Reitz, M. S., Kalyanaraman, V. S., and Gallo, R. C. Isolation of a new type C retrovirus (HTLV) in primary uncultured cells of a patient with Sezary T-cell leukemia. Nature 294: 268-271 (1981).
- 136. Gallo, R. C., and Wong-Staal, F. Current thoughts on the viral etiology of certain human cancers: The Richard and Hinda Rosenthal Foundation Award Lecture. Cancer Res. 44: 2743–2749 (1984).
- 137. Robert-Gurroff, M., Nakao, Y., Notake, K., Ito, Y., Sliski, A., and Gallo, R. C. Natural antibodies to human retrovirus HTLV in a cluster of Japanese patients with adult T-cell leukemia. Science 215: 975-978 (1982).
- 138. Kalyanaraman, V. S., Sarngadharan, M. G., Nakao, Y., Ito, Y., Aoki, T., and Gallo, R. C. Natural antibodies to the structural core protein (p24) of the human T-cell leukemia (lymphoma) retrovirus found in the sera of leukemia patients in Japan. Proc. Natl. Acad. Sci. (U.S.) 79: 1653-1657 (1982).
- Essex, M., Todar, G., and Zur Hausen, H., eds. Viruses in Naturally Occurring Cancers. Cold Spring Harbor Press, New York, 1980.
- 140. Purtilo, D. T. Immune deficiency predisposing to Epstein-Barr virus-induced lymphoproliferative diseases: The X-linked lymphoproliferative syndrome as a model. Adv. Cancer. Res. 34: 279-312 (1981).
- McDougall, J. K., Crum L. P., Fenoglio, L. M., Goldstein, L. C., and Galloway, D. A. Herpesvirus-specific RNA and protein in carcinoma of the uterine cervix. Proc. Natl. Acad. Sci. (U.S.) 79: 3853–3857 (1982).
- 142. Popper, H., Gerber, Thung, S. N. The relationship of hepatocellular carcinoma to infection with hepatitis-B and related viruses in man and animals. Hepatology 2: 1S-9S (1982).
- Gisser, S. D. Papovavirus and squamous cell carcinoma. Hum. Pathol. 12: 190-193 (1981).
- Anonymous. Medical News. Papilloma virus and cervical dysplasia. J. Am. Med. Assoc. 245: 2483 (1981).
- 145. Murad, T., Contesso, G., and Mouriesse, H. Papillary tumors of large legiforous dusts. Cancon 48: 122, 123 (1981)
- of large lactiferous ducts. Cancer 48: 122-133 (1981).

 146. Papkoff, J., Lai, M. H.-T., Hunter, T., and Verma, I. M. Analysis of transforming gene products from Moloney murine sarcoma virus. Cell 27: 109-119 (1981).

 147. Ward, J. F. Some biochemical consequences of the spatial dis-
- Ward, J. F. Some biochemical consequences of the spatial distribution of ionizing-produced free radicals. Rad. Res. 86: 185– 195 (1981).
- Borek, C. Radiation oncogenesis in cell culture. Adv. Cancer Res. 37: 159-232 (1982).
- 149. Miller, R. C., Geard, C. R., Osmak, R. S., Rutledge-Freeman, M., Ong, A., Mason, H., Napholz, A., Perez, N., Harisiadis, L., and Borek, C. Modification of sister chromatid exchanges and radiation-induced transformation in rodent cells by the tumor promoter 12-O-tetradecanoylphorbol-13-acetate and two retinoids. Cancer Res. 41: 655-659 (1981).
- Ward, J. F. Molecular mechanisms of radiation-induced damage to nucleic acids. Adv. Rad. Biol. 5: 181–239 (1977).
- Forbes, P. D., Davies, R. E., and Urbach, F. Aging, environmental influences and photocarcinogenesis. J. Invest. Dermatol. 73: 134 (1979).
- 152. Patrick, M. H., and Radin, R. O. Photochemistry of DNA and polynucleotides: photoproducts. In Photochemistry and Photobiology of Nucleic Acids, Vol. II (S. Y. Yang, Ed.), Academic Press, 1976, pp. 35-95.

- Abo-Darub, J. M., Mackie, R., and Pitts, J. D. DNA repair deficiency in lymphocytes from patients with actinic keratoses. Bull. Cancer 65: 357-362 (1978).
- 154. Sutherland, B. M., Harber, L. C., and Kochevar, I. E. Pyrimidine dimer formation and repair in human skin. Cancer Res. 40: 3181-3185 (1980).
- Cooke, A., and Johnson, B. E. Dose response, wavelength dependence and rate of ultraviolet radiation-induced pyrimidine dimers in mouse skin. Biochem. Biophys. Acta 517: 24-30 (1978).
- 156. Advisory Committee on the Biological Effects of Ionizing Radiations. The Effects on Populations of Exposure to Low Levels of Ionizing Radiation. National Academy of Sciences, Washington, DC, 1972.
- Upton, A. C. Environmental standards for ionizing radiation: theoretical basis for dose-response curves. Environ. Health Perspect. 52: 31–39 (1983).
- Drake, J. W., and Baltz, R. H. The biochemistry of mutagenesis. Ann. Rev. Biochem. 45: 11-37 (1976).
- 159. Shooter, K. V. DNA phosphotriesters as indicators of cumulative carcinogen-induced damage. Nature 274: 612-614 (1978).
- Arlett, C. F., and Lehmann, A. R. Human disorders showing increased sensitivity to the induction of genetic damage. Ann. Rev. Genet. 12: 95-115 (1978).
- Hart, R. W., Hall, K. Y., and Daniel, F. B. DNA repair and mutagenesis in mammalian cells. Photochem. Photobiol. 28: 131– 155 (1978).
- Lindahl, T. DNA glycosylases, endonucleases for apurinic-apyrimidinic sites and base-excision repair. Prog. Nucl. Acid Res. 22: 135-192 (1979).
- Thibodeau, L., and Verly, W. G. Cellular localization of the apurinic/-apyrimidinic endodeoxyribonucleases in rat liver. Eur. J. Biochem. 107: 555-563 (1980).
- 164. Roberts, J. J. Cellular responses to carcinogen-induced DNA damage and the role of DNA repair. Brit. Med. Bull. 36: 25-31 (1980)
- 165. Ornstein, R. L., and Rein, R. Molecular models of induced DNA premutational damage and mutational pathways for the carcinogen 4-nitroquinoline 1-oxide and its metabolites. Chem.-Biol. Interact. 30: 87-103 (1980).
- 166. Pullman, B., and Pullman, A. Nucleophilicity of DNA. Relation to chemical carcinogenesis. In Carcinogens: Fundamental Mechanisms and Environmental Effects (B. Pullman, P. O. P. Ts'o, and H. Gelboin, Eds.), Reidel, Amsterdam, 1980, pp. 55-66.
- 167. Gelboin, H. Benzo(a)pyrene metabolism, activation, and carcinogenesis: role and regulation of mixed-function oxidases and related enzymes. Physiol. Rev. 60: 1107-1166 (1980).
 168. Friedberg, E. C., Bonura, T., Love, J. D., McMillan, S., Ra-
- 168. Friedberg, E. C., Bonura, T., Love, J. D., McMillan, S., Radany, E. H., and Schultz, R. A. The repair of DNA damage: Recent developments and new insights. J. Supramol. Struct. Cell Biochem. 16: 91-103 (1981).
- 169. Grunberger, D., and Weinstein, I. B. Biochemical effects of the modification of nucleic acids by certain polycyclic aromatic carcinogens. Prog. Nucl. Acid Res. Mol. Biol. 23: 105–149 (1979).
- Waring, M. J. DNA modification and cancer. Ann. Rev. Biochem. 50: 159-192 (1981).
- Singer, B., and Kroger, M. Participation of modified nucleotides in translation and transcription. Prog. Nucl. Acid. Res. Mol. Biol. 23: 151-194 (1979).
- 172. Lin, J.-K., Miller, J. A., and Miller, E. C. 2,3-Dihydro-2-(guan-7-yl)-3-hydroxyaflatoxin B1, a major acid hydrolysis product of aflatoxin B1-DNA or -ribosomal RNA adducts formed in hepatic microsome-mediated reactions and in rat liver in vivo. Cancer Res. 37: 4430–4438 (1977).
- 173. Kriek, E., and Westra, J. G. Metabolic activation of aromatic amines and amides and interactions with nucleic acids. In: Chemical Carcinogens and DNA (P. L. Grover, Ed.), CRC Press, Boca Raton, FL, 1979, pp. 1–28.
- 174. Beland, F. A., Beranek, D. T., Dooley, K. L., Heflich, R. H., and Kadlubar, F. F. Arylamine-DNA adducts in vitro and in vivo: their role in bacterial mutagenesis and urinary bladder carcinogenesis. Environ. Health Perspect., in press.
- 175. Grunberger, D., and Weinstein, I. B. Conformational changes in nucleic acids modified by chemical carcinogens. In: Chemical

- Carcinogens and DNA (P. L. Grover, Ed.), CRC Press, Boca Raton, FL, 1979, pp. 59-93.
- 176. Jeffrey, A. M., Kinoshita, T., Santella, R. M., Grunberger, D., Katz, L., and Weinstein, I. B. The chemistry of polycyclic aromatic hydrocarbon-DNA adducts. In: Carcinogens: Fundamental Mechanisms and Environmental Effects (B. Pullman, P. O. P. Ts'o, and H. Gelboin, Eds.), Reidel, Amsterdam; 1980, pp. 565-579.
- 177. Harris, C., Trump, B. F., Grafstrom, R., and Autrup, H. Differences in metabolism of chemical carcinogens in cultured human epithelial tissue and cells. In: Mechanisms of Chemical Carcinogenesis (C. Harris, P. Cerutti, and C. F. Fox, Eds.), A. Liss Co., New York, 1982, pp. 289–292.
- 178. Radman, M., Villani, G., Boiteux, S., Defais, M., Caillet-Fauquet, P., and Spadari, S. On the mechanism and genetic control of mutagenesis induced by carcinogenic mutagens. In: Origins of Human Cancer: Mechanisms of Carcinogenesis, Vol. 4 (J. D. Watson and H. Hiatt, Eds.), Cold Spring Harbor Press, 1977.
- 179. Magee, P. N. The relationship between mutagenesis, carcinogenesis and teratogenesis. In: Progress in Genetic Toxicology (D. Scott, B. A. Bridges, and F. H. Sorbels, Eds.), Elsevier/ North Holland Biomedical Press, Amsterdam, 1977, pp. 15–27.
- Gerchman, L. L., and Ludlum, D. B. The properties of O6methylguanine in templates for RNA polymerase. Biochem. Biophys. Acta. 308: 310-316 (1973).
- Eisenstadt, E., Warren, A. J., Porter, J., Atkins, D., and Miller, J. H. Carcinogenic epoxides of benzo(a)pyrene and cyclopenta(cd)pyrene induce base substitutions via specific transversions. Proc. Natl. Acad. Sci. (U.S.) 79: 1945-1949 (1982).
- 182. Swenson, D. H., and Kadlubar, F. F. Properties of chemical mutagens and chemical carcinogens in relation to their mechanisms of action. In: Microbial Testers: Probing Carcinogenesis (I. C. Felkner, Ed.), Dekker, New York, 1981, pp. 3-33.
- Razin, A., and Friedman, J. DNA methylation and its possible biological roles. Progr. Nucl. Acid Res. Mol. Biol. 25: 33-52 (1981).
- 184. Brand, K. G., Buoen, L. C., Johnson, K. H., and Brand, I. Etiologic factors, stages, and the role of the foreign body in foreign body tumorigenesis: a review. Cancer Res. 35: 279-286 (1975).
- 185. Miller, K., Weintraub, Z., and Kagan, E. The effect of asbestos on macrophages. In: The In Vitro Effects of Mineral Dusts (R. C. Brown, M. Chamberlain, R. Davies, and I. P. Gormley, Eds.), Academic Press, New York, 1980, pp. 305-312.
- Trosko, J. E., and Chang, C. C. Genes, pollutants and human diseases. Quart. Rev. Biophys. 11: 603-627 (1978).
- 187. Newmark, P. Cancer genes—processed genes—jumping genes. Nature 296: 393-394 (1982).
- 188. Grover, P. L. (Ed.) Chemical Carcinogens and DNA, Vols. I and II, CRC Press, Boca Raton, FL, 1979.
- 189. Neidle, S. Carcinogens and DNA. Nature 283: 135 (1980).
- 190. Cohen, J. S. DNA: is the backbone boring? Trends Biochem. Sci. 5: 58-60 (1980).
- Lipetz, P. D., Galsky, A. G., and Stephens, R. E. Relationship of DNA tertiary and quaternary structure to carcinogenic processes. Adv. Cancer Res. 36: 165-210 (1982).
- Weinstein, I. B. Current concepts and controversies in chemical carcinogenesis. J. Supramol. Struct. Cell Biochem. 17: 99-120 (1981).
- Poirier, M. C. Antibodies to carcinogen-DNA adducts. J. Natl. Cancer Inst. 67: 515-519 (1981).
- 194. Randerrath, K., Reddy, M. V., and Gupta, R. C. ³²P-labeling test for DNA damage. Proc. Natl. Acad. Sci. (U.S.) 78: 6126–6129 (1981).
- Terzaghi, M., and Little, J. B. X-Radiation-induced transformation in a C3H mouse embryo-derived cell line. Cancer Res. 36: 1367-1374 (1976).
- Farber, E. Chemical carcinogenesis: a current biological perspective. Carcinogenesis 5: 1-5 (1984).
- 197. Farber, E. Sequential aspects of chemical carcinogenesis. In: Cancer A Comprehensive Treatise Etiology: Chemical and Physical Carcinogenesis. Vol. 1, 2nd Ed. (F. F. Becker, Ed.), Plenum Press, New York, 1982, pp. 485-506.

- Farber, E. Chemical carcinogenesis. New Engl. J. Med. 305: 1379–1389 (1981).
- 199. Maher, V. M., and McCormick, J. J. DNA repair and carcinogenesis. In: Chemical Carcinogens and DNA. Vol. I (P. L. Grover, Ed.), CRC Press, Boca Raton, FL, 1979, pp. 133-158.
- Cairns, J. The origin of human cancers. Nature 289: 353-357 (1981).
- 201. Larsen, K. H., Brash, D., Cleaver, J. E., Hart, R. W., Maher, V. M., Painter, R. B., and Sega, G. A. DNA repair assays as tests for environmental mutagens. A report of the U.S. EPA Gene-Tox program. Mutat. Res. 93: 287-318 (1982).
- Regan, J. D., Trosko, J., and Carrier, W. L. Evidence of excision of ultraviolet-induced pyrimidine dimers from the DNA of human cells in vitro. Biophys. J. 8: 319-325 (1968).
- Cleaver, J. E. Defective repair replication of DNA in xeroderma pigmentosum. Nature 218: 652–656 (1968).
- Lieberman, M. W., and Dipple, A. Removal of bound carcinogen during DNA repair in nondividing human lymphocytes. Cancer Res. 32: 1855–1860 (1972).
- Ahmed, F. E., and Setlow, R. B. Excision repair in mammalian cells. In: DNA Repair Mechanisms (P. C. Hanawalt, E. C. Friedberg, and C. F. Fox, Eds.), Academic Press, New York, 1978, pp. 333-336.
- 206. Grossman, L. Enzymes involved in the repair of damaged DNA. Arch. Biochem. Biophys. 211: 511-522 (1981).
- Ikenaga, M. Excision repair of DNA damage produced by 4nitroquinoline-1-oxide in cultured mammalian cells: Its relation to carcinogenesis. Gann Monogr. Cancer Res. 27: 21-32 (1982).
- Takebe, H., Ishiyaki, K., and Yagi, T. Genetic aspects of DNA repair deficiency. Gann Monogr. Cancer Res. 27: 13-19 (1982).
- Sekiguchi, M., Hayakawa, H., Makino, F., Tanaka, K., and Okada, Y. A human enzyme that liberates uracil from DNA. Biochem. Biophys. Res. Commun. 73: 293-299 (1976).
- Livneh, Z., Elad, D., and Sperling, J. Enzymatic insertion of purine bases into depurinated DNA in vitro. Proc. Natl. Acad. Sci. (U.S.) 76: 1089-1093 (1979).
- 211. Linn, S., Kuhnlein, U., and Deutsch, A. Enzymes from human fibroblasts for the repair of AP DNA. In: DNA Repair Mechanisms (P. C. Hanawalt, E. C. Friedberg, and C. F. Fox, Eds.), Academic Press, New York, 1978, pp. 199-203.
- 212. Deutsch, W. A. and Linn, S. DNA binding activity from cultured human fibroblasts that is specific for partially depurinated DNA and that inserts purines into apurinic sites. Proc. Natl. Acad. Sci. (U. S.) 76: 141-144 (1979).
- Verly, W. G., and Paquette, Y. An endonuclease for depurinated DNA in rat liver. Can. J. Biochem. 51: 1003-1009 (1973).
- 214. Craddock, V. M., and Henderson, A. R. The activity of 3-methyladenine DNA glycosylase in animal tissues in relation to carcinogenesis. Carcinogenesis 3: 747-750 (1982).
- 215. Margison, G. P. Chronic or acute administration of various dialkylnitrosamines enhances the removal of O6-methylguanine from rat liver DNA in vivo. Chem.-Biol. Interact. 38: 189-201 (1982).
- Laval, J., Pierre, J., and Laval, F. Release of 7-methylguanine residues from alkylated DNA by extracts of *Micrococcus luteus* and *Escherichia coli*. Proc. Natl. Acad. Sci. (U. S.) 78: 852-855 (1981).
- 217. Singer, B., and Brent, T. P. Human lymphoblasts contain DNA glycosylase activity excising N-3 and N-7 methyl and ethyl purines but not O6-alkylguanines or 1-alkyladenines. Proc. Natl. Acad. Sci. (U. S.) 78: 856-860 (1981).
- 218. Margison, G. P., and Pegg, A. E. Enzymatic release of 7-methylguanine from methylated DNA by rodent liver extracts. Proc. Natl. Acad. Sci. (U. S.) 78: 861-865 (1981).
- 219. Setlow, R. B. DNA repair pathways. In: DNA Repair and Mutagenesis in Eukaryotes (W. M. Generoso, M. D. Shelby, and F. J. de Serres, Eds.), Plenum Press, New York; 1980, pp. 45-54
- 220. Bogden, J. M., Eastman, A., and Bresnick, E. A system in mouse liver for the repair of O6-methylguanine lesions in methylated DNA. Nucl. Acid. Res. 9: 3089-3104 (1981).
- 221. Pegg, A. E., Scicchitano, D., and Dolan, M. E. Comparison of the rates of repair of O⁶-alkylguanines in DNA by rat liver and

- bacterial O⁶-alkylguanine-DNA alkyltransferase. Cancer Res. 44: 3806–3811 (1984).
- 222. Lindahl, T., Rydberg, B., Hjebnigren, T., Olsson, M., and Jacobson, A. Cellular defense mechanisms against alkylation of DNA. In: Lemontt J, Molecular and Cellular Mechanisms of Mutagenesis J. Lemontt and W. M. Generoso, Eds.), Plenum Press, New York, pp. 89-102.
- Lett, J. T., Caldwell, I., Dean, C. J., and Alexander, P. Rejoining of X-ray induced breaks in the DNA of leukaemia cells. Nature 214: 790-792 (1967).
- Corry, P. M., and Cole, A. Double strand rejoining in mammalian DNA. Nature 245: 100-101 (1973).
- 225. Cole, A., Shonka, F., Corry, P., and Cooper, W. G. CHO cell repair of single-strand and double-strand DNA breaks induced by gamma and alpha radiations. In: Molecular Mechanisms for Repair of DNA (P. C. Hanawalt and R. B. Setlow, Eds.), Plenum Press, New York, 1975, pp. 665-676.
- 226. Steiner, M. E., and Woods, W. G. Normal formation and repair of gamma radiation-induced single and double strand DNA breaks in Down syndrome fibroblasts. Mutat. Res. 95: 515-523 (1982).
- 227. Woods, W. G., Lopez, M., and Kalvonjian, S. L. Normal repair of gamma radiation-induced single-strand and double-strand DNA breaks in retinoblastoma fibroblasts. Biochim. Biophys. Acta 698: 40–48 (1982).
- 228. Nordenskjold, M., and Jernstrom, B. Induction and repair of DNA strand breaks in cultured human fibroblasts exposed to various phenols and dihydrodiols of benzo(a)pyrene. Chem.-Biol. Interact. 41: 155-168 (1982).
- 229. Kelner, A. Effect of visible light on the recovery of Streptomyces griseus conida from ultraviolet irradiation injury. Proc. Natl. Acad. Sci. (U. S.) 35: 73-79 (1949).
- Dulbecco, R. Experiments on photoreactivation of bacteriophages inactivated with ultraviolet radiation. J. Bacteriol. 59: 329-347 (1950).
- 231. Sutherland, B. M. Photoreactivating enzyme from human leukocytes. Nature 248: 109-112 (1974).
- Sutherland, B. M., Runge, P., and Sutherland J. C. DNA photoreactivating enzyme from placental mammals. Origin and characteristics. Biochemistry 13: 4710–4715 (1974).
- Meneghini, R., and Hanawalt, P. T4-endonuclease V-sensitive sites in DNA from ultraviolet-irradiated human cells. Biochim. Biophys. Act. 425: 428-437 (1976).
- 234. Meneghini, R., and Mench, C. F. M. Pyrimidine dimers in DNA strands of mammalian cells synthesized after UV-irradiation. In: DNA Repair Mechanisms (P. C. Hanawalt, E. C. Friedberg, and C. F. Fox, Eds.), Academic Press, New York, 1978, pp. 493-497.
- D'Ambrosio, S. M., Whetstone, J. W., Slazinski, L., and Lowney, E. Photorepair of pyrimidine dimers in human skin in vivo. Photochem. Photobiol. 34: 461–464 (1981).
- 236. D'Ambrosio, S. M., and Setlow, R. B. On the presence of UV-endonuclease sensitive sites in daughter DNA of UV-irradiated mammalian cells. In: DNA Repair Mechanisms (P. C. Hanawalt, E. C. Friedberg, and C. F. Fox, Eds.), Academic Press, New York, 1978, pp. 499-503.
 237. Bowden, G. T., Giesselbach, B., and Fusenig, N. E. Post-rep-
- Bowden, G. T., Giesselbach, B., and Fusenig, N. E. Post-replication repair of DNA in ultraviolet light-irradiated normal and malignantly transformed mouse epidermal cell cultures. Cancer Res. 38: 2709–2718 (1978).
- Waters, R. Repair of DNA in replicated and unreplicated portions of the human genome. J. Mol. Biol. 127: 117-127 (1979).
- Fornace, A., Jr. Recombination of parent and daughter strand DNA after UV-irradiation in mammalian cells. Nature 304: 552– 554 (1983).
- 240. Fujiwara, Y., and Tatsumi, M. Replicative bypass repair of ultraviolet damage to DNA of mammalian cells: caffeine sensitive and caffeine resistant mechanisms. Mutat. Res. 37: 91-110 (1976)
- Higgins, N. P., Kato, K., and Strauss, B. A model for replication repair in mammalian cells. J. Mol. Biol. 101: 417–425 (1976).
- 242. Sirover, M. A. Induction of the DNA repair enzyme uracil-DNA glycosylase in stimulated human lymphocytes. Cancer Res. 39:

- 2090-2095 (1979).
- D'Ambrosio, S. M. and Setlow, R. B. Enhancement of postreplication repair in Chinese hamster cells. Proc. Natl. Acad. Sci. (U.S.) 73: 2396–2400 (1976).
- 244. Painter, R. B. Does ultraviolet light enhance post-replication repair in mammalian cells? Nature 275: 243-245 (1978).
- 245. Sarasin, A. R., and Hanawalt, P. C. Carcinogens enhance survival of UV-irridated simian virus 40 in treated monkey kidney cells: induction of a recovery pathway? Proc. Natl. Acad. Sci. (U.S.) 75: 346-350 (1978).
- 246. Montesano, R., Bresil, H., and Margison, G. P. Increased excision of O⁶-methylguanine from rat liver DNA after chronic administration of dimethylnitrosamine. Cancer Res. 39: 1798–1802 (1979).
- Lindahl, T. DNA repair enzymes. Ann. Rev. Biochem. 51: 61– 87 (1982).
- 248. Mehta, J. R., Ludlum, D. B., Renard, A., and Verly, W. G. Repair of O₆-ethylguanine in DNA by a chromatin fraction from rat liver: transfer of the ethyl group to an acceptor protein. Proc. Natl. Acad. Sci. (U.S.) 78: 6766–6770 (1981).
- Hart, R. W., and Turturro, A. Evolution and longevity-assurance processes. Naturwissenschaften 68: 552-557 (1981).
- Wilkins, R. J., and Hart, R. W. Preferential DNA repair in human cells. Nature 247: 35–36 (1974).
- 251. Oleson, F. B., Mitchell, B. L., Dipple, A., and Lieberman, M. W. Distribution of DNA damage in chromatin and its relation to repair in human cells treated with 7-bromomethylbenz(a)anthracene. Nucl. Acid Res. 7: 1343-1361 (1979).
- 252. Cerutti, P. Persistence of carcinogen-DNA adducts in cultured mammalian cells. In: Mechanism of Chemical Carcinogenesis (C. C. Harris and P. A. Cerutti, Eds.), A. R. Liss, New York, 1982, pp. 419–427.
- 253. Sheikh, Y. M., Joyce, N. J., Daniel, F. B., Oravec, C. T., Cazer, F. D., Raber, J., Mhaskar, D., Witiak, D. T., Hart, R. W., and D'Ambrosio, S. M. Strain differences in organ selective DMBA-induced carcinogenicity: Comparative binding of dimethyl-benz(a)anthracene and its 2-fluoro analogue in Sprague-Dawley and Long-Evans rats. In: Polynuclear Aromatic Hydrocarbons (A. J. Dennis and W. M. Cooke, Eds.), Battelle Press, Columbus, 1981, pp. 625–639.
- 254. Cleaver, J. E., Thomas, G. H., Trosko, J. E., and Lett, J. T. Excision repair (dimer excision, strand breakage and repair replication) in primary cultures of eukaryotic (bovine) cells. Exptl. Cell Res. 74: 67–80 (1972).
- 255. Brown, H. S., Jeffrey, A. M., and Weinstein, I. B. Formation of DNA adducts in 10T1/2 mouse embryo fibroblasts incubated with benzo(a)pyrene or dihydrodiol oxide derivatives. Cancer Res. 39: 1673–1677 (1979).
- Cerutti, P., Shinohara, K., and Remsen, J. Repair of DNA damage induced by ionizing radiation and benzo(a)pyrene in mammalian cells. J. Toxicol. Environ. Health 2: 1375–1386 (1977).
- 257. Eastman, A., Mossman, B. T., and Bresnick, E. Formation and removal of benzo(a)pyrene adducts of DNA in hamster tracheal epithelial cells. Cancer Res. 41: 2605–2610 (1981).
- 258. Shinohara, K., and Cerutti, P. A. Excision repair of benzo-(a)pyrenedeoxyguanosine adducts in baby hamster kidney 21/ C13 cells and in secondary mouse embryo fibroblasts C57BL/6J. Proc. Natl. Acad. Sci. (U.S.) 74: 979-983 (1977).
- 259. Feldman, G., Remsen, J., Shinohara, K., and Cerutti, P. Excisability and persistence of benzo(a)pyrene DNA adducts in epithelioid human lung cells. Nature 274: 796–798 (1978).
- Dipple, A., and Roberts, J. J. Excision of 7-bromomethylbenz(a)anthracene-DNA adducts in replicating mammalian cells. Biochemistry 16: 1499–1503 (1977).
- 261. Ikenaga, M., Ishii, Y., Tada, M., Kakunaga, T., Takebe, H., and Kondo, S. Excision-repair of 4-nitroquinoline-1-oxide damage responsible for killing, mutation, and cancer. In: Molecular Mechanisms for Repair of DNA, Part B (P. C. Hanawalt and R. B. Setlow, Eds.), Plenum Press, New York, 1975, pp. 763–771.
- 262. Hart, R. W., and Setlow, R. B. Correlation between deoxyribonucleic acid excision-repair and life-span in a number of mammalian species. Proc. Natl. Acad. Sci. (U.S.) 71: 2169-2173

- (1974).
- 263. Sacher, G. A., and Hart, R. W. Longevity, aging and comparative cellular and molecular biology of the house mouse, *Mus musculus*, and the white-footed mouse, *Peromyscus leucopus*. In: Genetic Effects on Aging (D. Bergsma and D. Harrison, Eds.), Alan R. Liss, New York, Vol. 14, 1978, pp. 71-96.
- 264. Turturro, A., and Hart, R. W. DNA repair mechanisms in aging. In: Comparative Biology of Major Age-Related Diseases: Current Status and Research Frontiers (D. G. Sciapelli and G. Migaki, Eds.), A. R. Liss, 1984, pp. 19-45.
- 265. Cairns, J. Summary. In: Mechanism of Chemical Carcinogenesis (C. C. Harris and P. A. Cerutti, Eds.), AR Liss, New York, 1982, pp. 559–562.
- 266. Brash, D., and Hart, R. W. DNA damage and repair in vivo. J. Environ. Pathol. Toxicol. 2: 79-114 (1978).
- Walford, R. L. Multigene families, histocompatibility systems, tranformation, meiosis, stem cells, and DNA repair. Mech. Aging Dev. 9: 9-26 (1979).
- Janss, D. H., and Ben, T. L. Age-related modification of 7,12dimethylbenz(a)anthracene binding to rat mammary gland DNA. J. Natl. Cancer Inst. 60: 173-177 (1978).
- Sugimura, T. Poly(adenosine diphosphate ribose). Prog. Nucleic Acid. Res. Mol. Biol. 13: 127-151 (1973).
- Sugimura, T., and Miwa, M. Poly(ADP-ribose) and cancer research. Carcinogenesis 4: 1503–1506 (1983).
- 271. Oikawa, A., Tohda, H., Kanai, M., Miwa, M., and Sugimura, T. Inhibitors of poly(adenosine diphosphate ribose) polymerase induce sister chromatid exchanges. Biochem. Biophys. Res. Commun. 90: 1147-1152 (1980).
- 272. Breimer, L., and Lindahl, T. A DNA glycosylase for *Escherichia coli* that releases free urea from a polydeoxyribonucleotide containing fragments of base residues. Nucl. Acid Res., 8: 6199–6211 (1980).
- Demple, B., and Linn, S. DNA-glycosylases and UV repair. Nature, 287: 203–208 (1980).
- 274. Lloyd, R. S., and Hanawalt, P. C. Expression of the denV gene of bacteriophage T4 cloned in *Escherichia coli*. Proc. Natl. Acad. Sci. (U.S.) 78: 2796–2800 (1981).
- 275. Gombar, C. T., Katz, E. J., Magee, P. N., and Sirover, M. A. Induction of the DNA repair enzymes uracil DNA glycosylase and 3-methyladenine DNA glycosylase in regenerating rat liver. Carcinogesis 2: 595–599 (1981).
- 276. Krokan, H., and Wittwer, C. U. Uracil DNA-glycosylase from HeLa cells: general properties, substrate, specificity and effect of uracil analogs. Nucl. Acid Res. 9: 2599-2613 (1981).
- 277. Karran, P., and Lindahl, T. Hypoxanthine in deoxyribonucleic acid: generation by heat-induced hydrolysis of adenine residues and release in free form by a deoxyribonucleic acid glycosylase from calf thymus. Biochemistry 19: 6005–6011 (1980).
- 278. Karran, P., Lindahl, T., Ofsteng, I., Evenson, G. B., and Seeberg, E. Escherichia coli mutants deficient in 3-methyladenine-DNA glycosylase. J. Mol. Biol. 140: 101-127 (1980).
- Witkin, E. M. Ultraviolet mutagenesis and inducible DNA repair in Escherichia coli. Bacteriol. Rev. 40: 869-907 (1976).
- 280. Stara, J. F., Mukerjee, D., McGaughy, R., Durkin, P., and Dourson, M. L. The current use of studies on promoters and cocarcinogens in quantitative risk assessment. Environ. Health Perspect. 50: 359-368 (1983).
- Wani, A. A. DNA damage due to the spontaneous lability of nucleophilic agents. In: Biological Mechanisms in Aging Conference Proceedings, June 1980, (R. T. Schimke, Ed.), NIH Publication No. 81-2194, 1981, pp. 211-225.
- 282. Schendel, P. F. Inducible repair systems and their implications for toxicology. CRC Crit. Rev. Toxicol., 8: 311-362 (1981).
- Farber, E., and Cameron, R. The sequential analysis of cancer development. Adv. Cancer Res. 31: 125–226 (1980).
- 284. Loeb, L. A., Sirover, M. A., Weymouth, L. A., Dube, D. K., Seal, G., Agarwal, S. S., and Katz, E. Infidelity of DNA synthesis as related to mutagenesis and carcinogenesis. J. Toxicol. Environ. Health 2: 1297-1304 (1977).
- Burnet, F. M. Intrinsic mutagenesis: a genetic basis of ageing. Pathology 6: 1-11 (1974).
- 286. Guernsey, D. L., Ong, A., and Borek, C. Thyroid hormone mod-

- ulation of X-ray-induced in vitro neoplastic transformation. Nature 288: 591–592 (1980).
- Loeb, L. A., Weymouth, L. A., Kunkel, T. A., Gopinathan, K. P., Beckman, R. A., and Dube, D. K. On the fidelity of DNA replication. Cold Spring Harbor Symp. Quant. Biol. 43: 921-927 (1979).
- Sirover, M. A., and Loeb, L. A. Metal-induced infidelity during DNA synthesis. Proc. Natl. Acad. Sci. (U.S.) 73: 2331–2335 (1976).
- Loeb, L. A., Silber, J. R., and Fry, M. Infidelity of DNA replication in aging. In: Biological Mechanisms in Aging Conference Proceedings, June 1980 (R. T. Schimke, Ed.), NIH Publication No. 81–2194, 1981, pp. 270–278.
- 290. Kroger, M., and Singer, B. Ambiguity and transcriptional errors as a result of methylation of N-1 of purines and N-3 of pyrimidines. Biochemistry 18: 3493-3500 (1979).
- Mehta, J. R. and Ludlum, D. B. Synthesis and properties of O⁶-methyldeoxyguanylic acid and its copolymers with deoxycytidylic acid. Biochem. Biophys. Acta. 521: 770-778 (1978).
- Dube, D. K., and Loeb, L. A. Manganese as a mutagenic agent during in vitro DNA synthesis. Biochem. Biophys. Res. Commun. 67: 1041-1046 (1975).
- 293. Saffhill, R. The effect of ionizing radiation and chemical methylation upon the activity and accuracy of *E. coli* DNA polymerase I. Biochem. Biophys. Res. Commun. 61: 802–808 (1974).
- Shearman, C. W., and Loeb, L. A. Effects of depurination on the fidelity of DNA synthesis. J. Mol. Biol. 128: 197-218 (1979).
- Battula, N., and Loeb, L. A. The infidelity of avian myeloblastosis virus deoxyribonucleic acid polymerase in polynucleotide replication. J. Biol. Chem. 249: 4086–4093 (1974).
- 296. Kunkel, T. A., and Loeb, L. A. On the fidelity of DNA replication. Effect of divalent metal ion activators and deoxyribonucleoside triphosphate pools on in vitro mutagenesis. J. Biol. Chem. 254: 5718–5725 (1979).
- 297. Peterson, A. R., Landolph, J. R., Peterson, H., and Heidelberger, C. Mutagenesis of Chinese hamster cells is facilitated by thymidine and deoxycytidine. Nature 276: 508-510 (1978).
- 298. Meuth, M., Heureaux-Huard, N. L., and Trudel, M. Characterization of a mutator gene in Chinese hamster ovary cells. Proc. Natl. Acad. Sci. (U.S.) 76: 6505-6509 (1979).
- Weinberg, G., Ullman, B., and Martin, D. W., Jr. Mutator phenotypes in mammalian cell mutants with distinct biochemical defects and abnormal deoxyribonucleoside triphosphate pools. Proc. Natl. Acad. Sci. (U.S.) 78: 2447-2451 (1981).
- Topal, M. D., and Baker, M. S. DNA precursor pool: A significant target for N-methyl-N-nitrosourea in C3H/10T1/2 clone 8 cells. Proc. Natl. Acad. Sci. (U.S.) 79: 2211-2215 (1982).
- 301. Topal, M.D., Hutchinson, C. A. III, and Baker, M. S. DNA precursors in chemical mutagenesis: a novel application of DNA sequencing. Nature 298: 863-865 (1982).
- 302. Lindahl, T., and Nyberg, B. Rate of depurination of native deoxyribonucleic acid. Biochemistry 11: 3610–3618 (1972).
- 303. Brooks, A. L., Benjamin, S. A., James, R. K., and McClellan, R. O. Interaction of ¹⁴⁴Ce and partial hepatectomy in the production of liver neoplasms in the Chinese hamster. Radiat. Res. 91: 573–588 (1982).
- 304. Mehta, R. G., and Moon, R. C. Inhibition of DNA synthesis by retinyl acetate during chemically induced mammary carcinogenesis. Cancer Res. 40: 1109-1111 (1980).
- 305. Narisawa, T., Reddy, B. S., Wong, C.-Q, and Weisburger, J. H. Effect of vitamin A deficiency on rat colon carcinogenesis by N-methyl-N'-nitro-N-guanidine. Cancer Res. 36: 1379–1383 (1976).
- 306. Stott, W. T., Reitz, R. H., Schumann, A. M., and Watanabe, P. G. Genetic and nongenetic events in neoplasia. Food Cosmet. Toxicol. 19: 567–576 (1981).
- 307. Argyris, T. S. Epidermal tumor promotion by regeneration. In: Carcinogenesis: A Comprehensive Survey, Vol. 7, (E. Hecker, N. E. Fusenig, W. Kunz, F. Marks, and H. W. Thielmann, Eds.), Raven Press, New York, 1982, pp. 43–48.
- 308. Marks, F. Epidermal growth control mechanism, hyperplasia, and tumor promotion in the skin. Cancer Res. 36: 2636-2643 (1976).

- IARC. Chemicals, Industrial Processes and Industries Associated with Cancer in Humans. IARC Monographs, Supplement 4, IARC, Lyon, 1982.
- 310. Stott, W. T., and Watanabe, P. G. Differentiation of genetic versus epigenetic mechanisms of toxicity and its application to risk assessment. Drug Metabol. Rev. 13: 853-873 (1982).
- 311. Diamond, L., O'Brien, T. G., and Baird, W. M. Tumor promoters and the mechanism of tumor promotion. Adv. Cancer Res. 32: 1-74 (1980).
- 312. Berenblum, I. Sequential aspects of chemical carcinogenesis: skin. In: Cancer: A Comprehensive Treatise. Etiology: Chemical and Physical Carcinogenesis. Vol 1, 2nd Ed. (F. F. Becker, Ed.), Plenum Press, New York, 1982, pp. 451–484.
- 313. Verma, A. K., Conrad, E. A., and Boutwell, R. K. Differential effects of retinoic acid and 7, 8-benzoflavone on the induction of mouse skin tumors by the complete carcinogenesis process and by the initiation-promotion regimen. Cancer Res. 3519-3525 (1982).
- 314. Slaga, T. J. Overview of tumor promotion in animals. Environ. Health Perspect. 50: 3–14 (1983).
- 315. Littlefield, N. A., Farmer, J. H., Gaylor, D. W., and Sheldon, W. G. Effects of dose and time in a long-term, low-dose carcinogenic study. J. Environ. Pathol. Toxicol. 3: 17-34 (1979).
- Littlefield, N. A., Greenman, D. L., Farmer, J. H., and Sheldon, W. G. Effects of continuous and discontinued exposure to 2-AAF on urinary bladder hyperplasia and neoplasia. J. Environ. Pathol. Toxicol. 3: 35-54 (1979).
- 317. Hicks, R. M. Effect of promoters on the incidence of bladder cancer in experimental animal models. Environ. Health. Perspect. 50: 37-49 (1983).
- Van Duuren, B. L. Tumor-promoting and co-carcinogenic agents in chemical carcinogenesis. In: Chemical Carcinogens (C. E. Searle, Ed.), American Chemical Society Monograph, No. 173, American Chemical Society, Washington, DC, 1976, pp. 737–762.
- Leclercq, G., and Heuson, J. C. Physiological and pharmacological effects of estrogens in breast cancer. Biochim. Biophys. Acta. 560: 427-455 (1979).
- 320. Yager, J. D., Jr., and Yager, R. Oral contraceptive steroids as promoters of hepatocarcinogenesis in female Sprague-Dawley rats. Cancer Res. 40: 3680–3685 (1980).
- Solanki, V., and Slaga, T. J. Specific binding of phorbol ester tumor promoters to intact primary epidermal cells from SEN-CAR mice. Proc. Natl. Acad. Sci. (U.S.) 78: 2549–2553 (1981).
- 322. Dunphy, W. G., Delclos, K. B., and Blumberg, P. M. Characterization of specific binding of (³H)phorbol 12,13-dibutyrate and (³H)phorbol 12-myristate 13-acetate to mouse brain. Cancer Res. 40: 3635–3641 (1980).
- 323. Castagna, M., Takai, Y., Kaibuchi, K., Sano, K., Kikkawa, U., and Nishizuka, Y. Direct activation of calcium-activated phospholipid-dependent protein kinase by tumor-promoting phorbol esters. J. Biol. Chem. 257: 7847–7851 (1982).
- 324. Poland, A., and Glover E. 2,3,7,8-Tetrachlorodibenzo-p-dioxin: Segregation of toxicity with the Ah locus. Mol. Pharmacol. 17:86–94 (1980).
- 325. Slaga, T. J., Fischer, S. M., Weeks, C. E., and Klein-Szanto, A. J. P. Cellular and biochemical mechanisms of mouse skin tumor promoters. Rev. Biochem. Toxicol. 3: 231-281 (1981).
- 326. Clark, J. H., and Peck, E. J., Jr. Female Sex Steroids: Receptors and Function. Springer/Verlag, New York, 1979.
- 327. Boutwell, R. K. Some biological aspects of skin carcinogenesis. Progr. Exptl. Tumor Res. 4: 207–250 (1964).
- 328. Slaga, T. J., Fischer, S. M., Nelson, K., and Gleason, G. L. Studies on the mechanism of skin tumor promotion: Evidence for several stages in promotion. Proc. Natl. Acad. Sci. (U.S.) 77: 3659-3663 (1980).
- 329. Weinstein, I. B., Mufson, R. A., Lee, L. S., Fisher, P. B., Laskin, J., Horowitz, A. D., and Ivanovic, V. Membrane and other biochemical effects of the phorbol esters and their relevance to tumor promotion. In: Carcinogens: Fundamental Mechanisms and Environmental Effects (B. Pullman, P. O. P. Ts'o, and H. Gelboin, Eds.), Reidel, Amsterdam, 543–563, 1980.
- 330. Goldstein, B. D., Witz, G., Amoruso, M., Stone, D. S., and Troll,

- W. Stimulation of human polymorphonuclear leukocyte superoxide anion radical production by tumor promoters. Cancer. Letters, 11: 257–262 (1981).
- Slaga, T. J., Klein-Szanto, A. S. P., Triplett, L. L., Yotti, L. P., and Trosko, J. E. Skin tumor-promoting activity of benzoyl peroxide, a widely used free radical-generating compound. Science 213: 1023–1025 (1981).
- 332. Borek, C., and Troll, W. Modifiers of free radicals inhibit in vitro the oncogenic actions of x-rays, bleomycin and the tumor promoter 12-0-tetradecanoylphorbol 13-acetate. Proc. Natl. Acad. Sci. (U.S.) 80: 1304–1307 (1983).
- 333. Birnboim, H. C. DNA strand breakage in human leukocytes exposed to a tumor promoter, phorbol myristate acetate. Science 215: 1247–1249 (1982).
- 334. Rovera, G., O'Brien, T. G., and Diamond, L. Tumor promoters inhibit spontaneous differentiation of Friend erythroleukemia cells in culture. Proc. Natl. Acad. Sci. (U.S.) 74: 2894–2898 (1977)
- 335. Miao, R. M., Fieldsteel, A. H., and Fodge, D. W. Opposing effects of tumour promoters on erythroid differentiation. Nature 274: 271 (1978).
- 336. Greenberger, J. S., Newberger, P. E., Karpas, A., and Moloney, W. C. Constitutive and inducible granulocyte-macrophage functions in mouse, rat and human myeloid leukemia-derived continuous tissue culture lines. Cancer Res. 38: 3340–3348 (1978).
- 337. Emerit, I., and Cerutti, P. The tumor promoter phorbol-12-myristate-13-acetate induces chromosome aberrations in human lymphocytes via indirect action. In: Mechanism of Chemical Carcinogenesis (C. C. Harris and P. A. Cerutti, Eds.), AR Liss, New York, 1982, pp. 495-498.
- 338. Nagasawa, H., and Little, J. B. Effect of tumor promoters, protease inhibitors, and repair processes on x-ray-induced sister chromatid exchanges in mouse cells. Proc. Natl. Acad. Sci. (U.S.) 76: 1943–1947 (1979).
- 339. Zur Hausen, H., O'Neill, F. J., Freese, U. K., and Hecker, E. Persisting oncogenic herpesvirus induced by the tumour promoter TPA. Nature 272: 373-375 (1978).
- 340. Harnden, D. G., and Taylor, A. M. R. Chromosomes and neoplasia. Adv. Human Genet. 9: 1-70 (1979).
- 341. Klein, G. Lymphoma development in mice and humans: Diversity of initiation is followed by convergent cytogenetic evolution. Proc. Natl. Acad. Sci. (U.S.), 76: 2442-2446 (1979).
- Rowley, J. D. Human oncogene locations and chromosome aberrations. Nature 301: 290–291 (1983).
- 343. Clarkson, B. D., Fried, J., Chou, T.-C., Strife, A., Ferguson, R., Sullivan, S., Kitahara, T., and Oyama, A. Duration of the dormant state in an established cell line of human hematopoietic cells. Cancer Res. 37: 4506–4522 (1977).
- 344. Clarkson, B., Strife, A., and DeHarven, E. Continuous culture of seven new cell lines (SK-L1 to 7) from patients with acute leukemia. Cancer Res. 20: 926-947 (1967).
- 345. Todo, A., Strife, A., Fried, J., and Clarkson, B. D. Proliferative kinectics of human hematopoietic cells during different growth phases in vitro. Cancer Res. 31: 1330-1340 (1971).
- 346. Saedler, H., and Starlinger, P. O⁶ Mutations in the galactose operon in *E. coli*. Mol. Gen. Genet. 100: 178-189 (1967).
- 347. Goth, R., and Rajewsky, M. F. Persistence of O⁶-ethylguanine in rat brain DNA: correlation with nervous system specific carcinogenesis by ethylnitrosourea. Proc. Natl. Acad. Sci. (U.S.) 71: 639-643 (1974).
- 348. Radman, M., and Kinsella, A. R. Chromosomal events in carcinogenic initiation and promotion: implications for carcinogenicity testing and cancer prevention strategies. In: IARC Sci. Publ. No. IARC, Lyon, 1980, pp. 75-90.
- 349. Karin, M., Haslinge, A., Holtgreve, H., Richards, R. I., Krauter, P., Westphal, H. W., and Beato M. Characterization of DNA sequences through which cadmium and glucocorticoid hormones induce human metallothionein-IIA gene. Nature 308: 513-519 (1984).
- Doll, R. An epidemiological perspective of the biology of cancer. Cancer Res. 38: 3573–3583 (1978).
- Nilsson, K., and Klein, G. Phenotypic and cytogenetic characteristics of human B-lymphoid cell lines and their relevance for

- the etiology of Burkitt's lymphoma. Adv. Cancer Res. 37: 319-380 (1982).
- 352. Van Duuren, B. L., Sivak, A. Segal, A., Seidman, I., and Katz, C. Dose-response studies with a pure tumor-promoting agent, phorbol myristate acetate. Cancer Res. 33: 2166-2172 (1973).
- 353. Melnick, R., Boorman, G., Haseman, J., Montali, R., and Huff, J. Urolithiasis and bladder carcinogenicity of melamine in rodents. Toxicol. Appl. Pharmacol. 72: 292-303 (1984).
- 354. Perera, F. P. The genotoxic epigenetic distinction: relevance to cancer policy. Environ. Res. 34: 175-191 (1984).
- Hankin, J. H., and Rawlings, V. Diet and breast cancer: a review. Am. J. Clinical Nutr. 31: 2005–2016 (1978).
- Schwarz, J. A., Viaje, A., Slaga, T. J., Yuspa, S. H., Hennings, H., and Lichti, U. Fluocinolone acetonide: a potent inhibitor of mouse skin tumor promotion and epidermal DNA synthesis. Chem.-Biol. Interact. 17: 331-347 (1977).
- 357. Graham, S. Epidemiology of retinoids and cancer. J. Natl. Cancer Inst. 1423-1428 (1984).
- 358. Hart, I. R., and Fidler, I. J. Cancer invasion and metastasis. Quart. Rev. Biol. 55: 121-142 (1980).
- Sandberg, A. A. The Chromosomes in Human Cancer and Leukemia. Elsevier/North-Holland, Amsterdam, 1980.
- 360. Hopper, K. E., Harrison, J., and Nelson, D. S. Partial characterization of anti-tumor effector macrophages in the peritoneal cavities of concomitantly immune mice and mice injected with macrophage-stimulating agents. J. Reticuloendothel. Soc. 26: 259-271 (1979).
- Farrar, W. L., and Elgert, K. D. Suppressor cell activity in tumor-bearing mice. II. Inhibition of DNA synthesis and DNA polymerases by TBH splenic suppressor cells. J. Immunol. 120: 1354–1361 (1978).
- Ratner, L., Nordlund, J. J., and Lengyel, P. Interferon as an inhibitor of cell growth: studies with mouse melanoma cells. Proc. Soc. Exptl. Biol. Med. 163: 267-272 (1980).
- 363. Gimbrone, M. A., Jr., Cotran, R. S., Leapman, S. B., and Folkman, J. Tumor growth and neovascularization: an experimental model using the rabbit cornea. J. Natl. Cancer Inst. 52: 413-427 (1974).
- 364. Mueller, G. C., Kajiwara, K., Kim, U. H., and Graham, J. Proposed coupling of chromatin replication, hormone action and cell differentiation. Cancer Res. 38: 4041–4045 (1978).
- Fidler, I. J., Gersten, D. M., and Hart, I. R. The biology of cancer invasion and metastasis. Adv. Cancer Res. 28: 149-250 (1978).
- Liotta, L. A., Tryggvason, K., Garbisa, S., Hart, I., Foltz, C. M., and Shafie, S. Metastatic potential correlates with enzymatic degradation of basement membrane collagen. Nature 284: 67-68 (1980).
- 367. Poole, A. R., Tiltman, K. J., Recklies, A. D., and Stoker, T. A. M. Differences in secretion of the proteinase cathespin B at the edges of human breast carcinomas and fibroadenomas. Nature 273: 545-547 (1978).
- 368. Van De Velde, C. J. H., Van Putten, L. M., and Zwaveling, A. A new metastasizing mammary carcinoma model in mice: Model characteristics and applications. Eur. J. Cancer 13: 555-565
- Warren, B. A. Platelet-tumor cell interactions: morphological studies. In: Platelets: A Multidisciplinary Approach (G. D. Gaetano and S. Garattini, Eds.), Raven Press, New York, 1978, pp. 427-445
- 370. Poste, G., and Nicolson, G. L. Arrest and metastasis of bloodborne tumor cells are modified by fusion of plasma membrane vesicles from highly metastatic cells. Proc. Natl. Acad. Sci. (U.S.) 77: 399-403 (1980).
- 371. Baldwin, R. W., and Price, M. R. Neoantigen expression in chemical carcinogenesis. In: Cancer A Comprehensive Treatise Etiology: Chemical and Physical Carcinogenesis, Vol. 1, 2nd Ed. (F. F. Becker, Ed.), Plenum Press, New York, 1982, pp. 507– 548.
- 372. Kripke, M. L. Antigenicity of murine mouse skin tumors induced by ultraviolet light. J. Natl. Cancer Inst. 53: 1333-1336 (1974).
- 373. Hewitt, H. B. The choice of animal tumors for experimental studies of cancer therapy. Adv. Cancer Res. 27: 149-200 (1978).

- 374. Embleton, M. J., and Middle, J. G. Immune responses to naturally occurring rat sarcomas. Brit. J. Cancer 43: 44-52 (1981).
- 375. McKhann, C. F. Tumor immunology: past, present and future. In: Accomplishments in Cancer Research 1981 (J. G. Fortner and J. E. Rhoads, Eds.), Lippincott, Philadelphia, 1982, pp. 125–137
- Herlyn, M., Steplewski, Z., Herlyn, D., and Koprowski, H. Colo-rectal carcinoma-specific antigen: detection by means of monoclonal antibodies. Proc. Natl. Acad. Sci. (U.S.) 76: 1438 (1979).
- 377. Levine, A. J. Transformation-associated tumor antigens. Adv. Cancer Res. 37: 75-109 (1982).
- Theofilopoulos, A. N., and Dixon, F. J. Immune complexes in human diseases: a review. Am. J. Pathol. 100: 531-594 (1980).
- 379. Kripke, M. L. Speculations on the role of ultraviolet radiation in the development of malignant melanoma. J. Natl. Cancer Inst. 63: 541-548 (1979).
- 380. Klein, G. Immune and non-immune control of neoplastic development: Contrasting effects of host and tumor evolution. Cancer 45: 2486-2499 (1980).
- Lee, Y.-T. N., Sparks, F. C., Eilber, F. R., and Morton, D. L. Delayed cutaneous hypersensitivity and peripheral lymphocyte counts in patients with advanced cancer. Cancer 35: 748-755 (1975).
- 382. Fidler, I. J., Gersten, D. M., and Kripke, M. L. Influence of immune status on the metastasis of three murine fibrosarcomas of different immunogenicities. Cancer Res. 39: 3816-3821 (1979).
- 383. Heppner, G. H. Tumor heterogeneity. Cancer Res. 44: 2259–2265 (1984).
- 384. Reif, A. E. Synergism in carcinogenesis. Cancer Res. 73: 25–36 (1984).
- 385. Berenbaum, M. C. Criteria for analyzing the interactions between biologically-active agents. Adv. Cancer Res. 35: 269-335 (1981).
- 386. Selikoff, I. J., Hammond, E. C., and Churg, J. Asbestos exposure, smoking and neoplasia. J. Am. Med. Assoc. 204: 106–112 (1968).
- 387. Lakowicz, J. R., Englund, F., and Hidmark, A. Particle-enhanced membrane uptake of a polynuclear aromatic hydrocarbons: a possible role in cocarcinogenesis. J. Natl. Cancer Inst. 61: 1155–1159 (1978).
- 388. Kandaswami, C., and O'Brien, P. J. Effects of asbestos on membrane transport and metabolism of benzo(a)pyrene. Biochem. Biophys. Res. Commun. 97: 794-801 (1980).
- 389. Mossman, B. T., Craighead, J. E., and MacPherson, B. V. Asbestos-induced epithelial changes in organ cultures of hamster trachea: inhibition by retinyl methyl ether. Science 207: 311-313 (1980).
- 390. Mossman, B. T., and Craighead, J. E. Mechanisms of asbestos carcinogenesis. Environ. Res. 25: 269-280 (1981).
- 391. Miller, E. C., Miller, J. A., and Brown, R. R. On the inhibitory action of certain polycyclic hydrocarbons on azo dye carcinogenesis. Cancer Res. 12: 282–283 (1952).
- 392. Conney, A. H., Miller, E. C., and Miller, J. A. The metabolism of methylated aminoazo dyes. V. Evidence for induction of enzyme synthesis in the rat by 3-methylcholanthrene. Cancer Res. 16: 450-460 (1956).
- 393. Boveri, T. The Origin of Malignant Tumors. Williams and Wilkins, Baltimore, 1929.
- 394. Burdette, W. J. Significance of mutation in relation to the origin of tumors: a review. Cancer Res. 15: 201-226 (1955).
- 395. Miller, J. A. Carcinogenesis by chemicals: an overview. Cancer Res. 30: 559-576 (1970).
- 396. Malling, H. V. Dimethylnitrosamine: formation of mutagenic compounds by interactions with mouse liver microsomes. Mutat. Res. 13: 425-429 (1971).
- 397. Ames, B. N., Durston, W. E., Yamasaki, E., and Lee, F. D. Carcinogens are mutagens: a simple test system combining liver homogenates for activation and bacteria for detection. Proc. Natl. Acad. Sci. (U.S.) 70: 2281–2285 (1973).
- Hollstein, J., McCann, J., Angelosanto, F., and Nicols, W. Short-term tests for carcinogens and mutagens. Mutat. Res. 65: 133-226 (1979).

- 399. Purchase, I. F. H. An appraisal of predictive tests for carcinogenicity. Mutat. Res. 99: 53-71 (1982).
- 400. Weisburger, J. H., and Williams, G. M. Basic science of poisons. In: Toxicology, 2nd ed. (J. Doull, C. D. Klaassen, and M. O. Amdur, Eds.), MacMillan, New York, 1970.
- Committee on Chemical Environmental Mutagens, National Research Council. Identifying and estimating the genetic import of chemical mutagens. National Academy Press, Washington, DC, 1983.
- Hartman, P. E. Mutagens: some possible health impacts beyond carcinogenesis. Environ. Mut. 5: 139-152.
- 403. Brusick, D. J., Simmon, V. F., Rosenkranz, H. S., Ray, V. A., and Stafford, R. S. An evaluation of the *Escherichia coli* WP425 and WP425 uvrA reverse mutations assay. Mutat. Res. 76: 169– 190 (1980).
- Generoso, W. M., Bishop, J. B., Gosslee, D. G., Newell, G. W., Sheu, C. J., and Von Halle, E. Heritable translocation test in mice. Mutat. Res. 76: 191–215 (1980).
- 405. Hsie, A. W., Casciano, D. A., Couch, D. B., Krahn, D. F., O'Neill, J. P., and Whitfield, B. L. The use of Chinese hamster ovary cells to quantify specific locus mutation and to determine mutagenicity of chemicals. A report of the Gene-Tox Program. Mutat. Res. 86: 193-214 (1981).
- 406. Russell, L. B., Selby, P. B., Von Halle, E., Sheridan, W., and Valcovic, L. The mouse specific-locus test with agents other than radiations: interpretation of data and recommendations for future work. Mutat. Res. 86: 329–354 (1981).
- 407. Russell, L. B., Selby, P. B., Von Halle, E., Sheridan, W., and Valcovic, L. Use of the mouse spot test in chemical mutagenesis: interpretation of past data and recommendations for future work. Mutat. Res. 86: 355–379 (1981).
- 408. Latt, S. A. Allen, J., Bloom, S. E., Carrano, A., Falke, E., Kram, D., Schneider, E., Schreck, R., Tice, R., Whitfield, B., and Wolff, S. Sister-chromatid exchanges: a report of the Gene-Tox Program. Mutat. Res. 87: 17-62 (1981).
- 409. Bradley, M. O., Bhuyan, B., Francis, M. C., Langenbach, R., Peterson, A., and Huberman, E. Mutagenesis by chemical agents in V79 Chinese hamster cells: a review and analysis of the literature. A report of the Gene-Tox Program. Mutat. Res. 87: 81-142 (1981).
- 410. Preston, R. J., Au, W., Bender, M. A., Brewen, J. G., Carrano, A. V., Heddle, J. A., McFee, A. F., Wolff, S., and Wassom, J. S. Mammalian in vivo and in vitro cytogenetic assays: A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 87: 143-188 (1981).
- 411. Leifer, Z., Kada, T., Mandel, M., Zeiger, E., Stafford, R., and Rosenkranz, H. S. An evaluation of tests using DNA repairdeficient bacteria for predicting genotoxicity and carcinogenicity. A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 87: 211–297 (1981).
- 412. Kafer, E., Scott, B. R., Dorn, G. L., and Stafford, R. Asper-gillus nidulans: systems and results of tests for chemical induction of mitotic segregation and mutation. I. Diploid and duplication assay systems. A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 98: 1–48 (1981).
- 413. Scott, B. R., Dorn, G. L., Kafer, E., and Stafford, R. Aspergillus nidulans: systems and results of tests for induction of mitotic segregation and mutation. II. Haploid assay systems and overall response of all systems. A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 98: 49-94 (1981).
- 414. Larsen (Mavournin), K. H., Brash, D., Cleaver, J. E., Hart, R. W., Maher, V. M., Painter, R. B., and Sega, G. A. DNA repair assays as tests for environmental mutagens. A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 98: 287-318 (1982).
- 415. Legator, M. S., Bueding, E., Batzinger, R., Conner, T. H., Eisenstadt, E., Farrow, M. G., Ficsor, G., Hsie, A., Seed, J., and Stafford, R. S. An evaluation of the host-mediated assay and body fluid analysis. A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 98: 319-374 (1982).
- Constantin, M. J., and Owens, E. T. Introduction and perspectives of plant genetic and cytogenetic assays. A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 99: 1-12 (1982).

- 417. Constantin, M. J., and Nilan, R. A. Chromosome aberration assays in barley (*Hordeum vulgare*). A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 99: 13-36 (1982).
- 418. Constantin, M. J., and Nilan, R. The chlorophyll-deficient mutant assay in barley (*Hordeum vulgare*). A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 99: 37-49 (1982).
- Redei, G. P. Mutagen assay with Arabidopsis. A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 99: 243–255 (1982).
- 420. Ma, T. H. Vicia cytogenetic tests for environmental mutagens. A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 99: 257-271 (1982).
- Grant, W. F. Chromosome aberration assays in Allium. A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 99: 273–291 (1982).
- 422. Ma, T.-H. Tradescantia cytogenetic tests (root-tip mitosis, pollen mitosis, pollen mother-cell meiosis). A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 99: 293-302 (1982).
- 423. Van't Hof, J., and Schairer, L. A. Tradescantia assay system for gaseous mutagens. A report of the U. S. EPA Gene-Tox Program. Mutat. Res. 99: 303-315 (1982).
- 424. Plewa, M. J. Specific-locus mutation assays in Zea mays. A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 99: 317-337 (1982).
- 425. Vig, B. K. Soybean (Glycine max (L.) merrill) as a short-term assay for study of environmental mutagens. A report of the U. S. EPA Gene-Tox Program. Mutat. Res. 99: 339-347 (1982).
- 426. Heidelberger, C., Freeman, A. E., Pienta, R. J., Sivak, A., Bertram, J. S., Casto, B. C., Dunkel, V. C., Francis, M. W., Kakunage, T., Little, J. B., and Schechtman, L. M. Cell transformation by chemical agents: a review and analysis of the literature. A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 114: 283–385 (1983).
- 427. Wyrobek, A. J., Gordon, L. A., Burkhart, J. G., Francis, M. W., Kapp, R. W. Jr., Letz, G., Malling, H. V., Topham, J. C., and Whorton, M. D. An evaluation of the mouse sperm morphology test and other sperm tests in nonhuman mammals. A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 115: 1-72 (1983).
- 428. Wyrobek, A. J., Gordon, L. A., Burkhart, J. G., Francis, M. W., Kapp, R. W. Jr., Letz, G., Malling, H. V., Topham, J. C., and Whorton, M. D. An evaluation of human sperm as indicators of chemically induced alterations of spermatogenic function. A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 115: 73–148 (1983).
- 429. Loprieno, N., Barale, R., Von Halle, E. S., and von Borstel, R. C. Testing of chemicals for mutagenic activity with *Schizosaccharomyces pombe*. A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 115: 215–223 (1983).
- Clive, D., McCuen, R., Spector, J. F. S., Piper, C., and Mavournin, K. H. Specific gene mutations in L5178Y cells in culture. A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 115: 225-251 (1983).
- 431. Mitchell, A. D., Casciano, D. A., Meltz, M. L., Robinson, D. E., San, R. H. C., Williams, G. M., and Von Halle, E. S. Unscheduled DNA synthesis tests. A report of the U. S. EPA Gene-Tox Program. Mutat. Res. 123: 363-410 (1983).
- 432. de Serres, F. J., and Ashby, J., eds. Evaluation of short-term tests for carcinogens: Report of the International Collaborative Program. Elsevier/North Holland, New York, 1981.
- Maron, D. M., and Ames, B. N. Revised methods for the Salmonella mutagenicity test. Mutat. Res. 113: 173-215 (1983).
- 434. Ames, B. N., McCann, J., and Yamasaki, E. Methods for detecting carcinogens and mutagens with the Salmonella/mammalian-microsome mutagenicity test. Mutat. Res. 99: 53-71 (1982).
- 435. Thilly, W. G., DeLuca, J. G., Furth, E. E., Hoppe, H. IV, Kaden, D. A., Krolewski, J., Liber, H. L., Skopek, T. R., Slapikoff, S. A., Tizard, R. J., and Penman, B. W. Gene-locus mutation assays in diploid human lymphoblast lines. In: Chemical Mutagens, Vol. 6 (F. J. de Serres and A. Hollaender, Eds.) Plenum Press, New York, 1980.
- Rowley, J. D. Chromosome abnormalities in cancer. Cancer Genet. Cytogenet. 2: 175–198 (1980).

- 437. Sankaranarayanan, K. Genetic effects of ionizing radiation. In: Multicellular Eukaryotes and the Assessment of Genetic Radiation Hazards in Man. Elsevier Biomedical Press, Amsterdam, 1982
- Yunnis, J. J. The chromosomal basis of human neoplasia. Science 221: 227–236 (1983).
- 439. Tayler, J. H., Woods, P. S., and Hughes, W. L. The organization and duplication of chromosomes as revealed by autoradiographic studies using tritium-labelled thymidine. Proc. Natl. Acad. Sci. (U.S.) 43: 122–127 (1957).
- Latt, S. H. Microfluorometric detection of DNA replication in human metaphase chromosomes. Proc. Natl. Acad. Sci. (U.S.) 70: 3395-3399 (1973).
- 441. Heddle, J. A., Hite, M., Kirkhart, B., Mavournin, K., Mac-Gregor, J. T., Newell, G. W., and Salamone, M. F. The induction of micronuclei as a measure of genotoxicity. A report of the U.S. EPA Gene-Tox Program. Mutat. Res. 123: 61-118 (1983).
- Brusick, D. Principles of Genetic Toxicology. Plenum Press, New York, 1980.
- 443. Williams, G. M. Carcinogen induced DNA repair in primary rat liver cell cultures: a possible screen for chemical carcinogens. Cancer Letters, 1: 231–236 (1976).
- 444. Lutz, W. K. In vivo covalent binding of organic chemicals to DNA as a quantitative indicator in the process of chemical carcinogenesis. Mutat. Res. 65: 289-356 (1979).
- 445. Slater, E. E., Anderson, M.D., and Rosenkranz, H. S. Rapid detection of mutagens and carcinogens. Cancer Res. 31: 970-973 (1971).
- Heidelberger, C. Chemical carcinogenesis. Ann. Rev. Biochem. 44: 79-121 (1975).
- Weinberg, R. A. Oncogenes of spontaneous and chemically induced tumors. Adv. Cancer. Res. 36: 149–163 (1982).
- 448. Bishop, J. M. Retroviruses and cancer genes. Adv. Cancer Res. 37: 1-28 (1982).
- Yotti, L. P., Chang, C. C., and Trosko, J. E. Elimination of metabolic cooperation in Chinese hamster cells by a tumor promotor. Science 206: 1089–1091 (1979).
- Berenblum, I. The cocarcinogenic action of croton resin. Cancer Res. 1: 44–48 (1941).
- 451. Hecker, E. Structure-activity relationships in diterpene esters irritant and cocarcinogenic to mouse skin. In: Mechanisms of Tumor Promotion and Cocarcinogenesis. Carcinogenesis, Vol. 2. (T. J. Slaga, A. Sivak, and R. K. Boutwell, Eds.), Raven Press, New York, 1978.
- 452. Slaga, T. J., Fischer, S. M., Triplett, L. L., and Nesnow, S. Comparison of complete carcinogenesis and tumor initiation and promotion in mouse skin: the induction of papillomas by tumor initiation—promotion of a reliable short-term assay. J. Am. Coll. Toxicol. 1: 83-99 (1982).
- Pereira, M. A. Mouse skin bioassay for chemical carcinogenesis.
 J. Am. Coll. Toxicol. 1: 47–82 (1982).
- 454. Shimkin, M. B. Induced pulmonary tumors in mice: II. Reaction of lungs of strain A mice to carcinogenic hydrocarbons. Arch. Pathol. 29: 239–255 (1940).
- Andervont, H. B., and Shimkin, M. B. Biologic testing of carcinogens: II. Pulmonary-tumor-induction technique. J. Natl. Cancer Inst. 1: 225-239 (1941).
- Shimkin, M. B., and Stoner, G. D. Lung tumors in mice: application to carcinogenesis bioassay. Adv. Cancer Res. 21: 1-58 (1975).
- Stoner, G. D., and Shimkin, M. B. Strain A mouse lung tumor bioassay. J. Am. Coll. Toxicol. 1: 145-169 (1982).
- 458. Maronpot, R. R., Witschi, H. P., Smith, L. H., and McCoy, J. L. Recent experience with the strain A mouse pulmonary tumor bioassay model. In: Short-Term Bioassays in the Analysis of Complex Environmental Mixtures III (M. D. Waters, S. S. Sandhu, J. Lewtas, L. Claxton, N. Chernoff, and S. Nesnow, Eds.), Plenum Press, New York, 1983.
- 459. Peraino, C., Fry, R. J. M., and Staffeldt, E. Reduction and enhancement by phenobarbital of hepatocarcinogenesis induced in the rat by 2-acetyl-aminofluorene. Cancer Res. 31: 1506-1512 (1971).
- 460. Pitot, H. C., Barsness, L., Goldsworthy, T., and Kitagawa, T.

- Biochemical characterization of stages of hepatocarcinogenesis after a single dose of diethylnitrosamine. Nature 271: 456–457 (1978).
- 461. Solt, E., and Farber, E. New principle for the analysis of chemical carcinogenesis. Nature 263: 701-703 (1976).
- 462. Pereira, M. A. Rat liver foci bioassay. J. Am. Coll. Toxicol. 1: 101-117 (1982).
- 463. Gabridge, M. G., and Legator, M. S. A host-mediated assay for the detection of mutagenic compounds. Proc. Soc. Exptl. Biol. Med. 130: 831-834 (1969).
- 464. Huberman, E., and Sachs, L. Cell-mediated mutagenesis of mammalian cells with chemical carcinogens. Int. J. Cancer 13: 326-333 (1974).
- Langenbach, R., and Oglesby, L. The use of intact cellular activation systems in genetic toxicology assays. In: Chemical Mutagens, Vol. 8 (F. J. deSerres, Ed.), Plenum Press, New York, 1982.
- Ashby, J. The unique role of rodents in the detection of possible human carcinogens and mutagens. Mutat. Res. 115: 177-213 (1983).
- Margolin, B. H., Kaplan, N., and Zeiger, E. Statistical analysis of the Ames Salmonella/microsome test. Proc. Natl. Acad. Sci. (U.S.) 78: 3779–3783 (1981).
- 468. Bernstein, L., Kaldor, J., McCann, J., and Pike M. C. An empirical approach to the statistical analysis of mutagenesis data from the Salmonellla test. Mutat. Res. 97: 267-281 (1982).
- Stead, A. G., Hasselblad, V., Creason, J. P., and Claxton, L. Modeling the Ames test. Mutat. Res. 85: 13–27 (1981).
- 470. Snee, R. D., and Irr, J. D. Design of a statistical method for the analysis of mutagenesis at the hypoxanthine-guanine phosphoribosyl transferase locus of cultured Chinese hamster ovary cells. Mutat. Res. 85: 77-93 (1981).
- Purchase, I. F. H. ICPEMC working paper 2/6: An appraisal of predictive tests for carcinogenicity. Mutat. Res. 99: 53–71 (1982).
- Heinze, J. H., and Poulsen, N. K. The optimal design of batteries of short-term tests for detecting carcinogens. Mutat. Res. 117: 259-269 (1983).
- 473. McCann, J., Choi, E., Yamasaki, E., and Ames, B. Detection of carcinogens as mutagens in the Salmonella/microsome test: assay of 300 chemicals. Proc. Natl. Acad. Sci. (U.S.) 72: 5135–5139 (1975).
- 474. Shelby, M. D., and Stasiewicz, S. Chemicals showing no evidence of carcinogenicity in long-term, two-species rodent studies: their use in short-term test evaluations. Environ. Mut. 6: 871–878 (1984).
- 475. Rinkus, S. J., and Legator, M. S. Chemical characterization of 465 known or suspected carcinogens and their correlation with mutagenic activity in the *Salmonella typhimurium* system. Cancer Res. 39: 3289-3318 (1979).
- 476. Bartsch, H., Malaveille, C., Camus, A. M., Martel-Planche, G., Brun, G., Hautefeuille, A., Sabadie, N., Barbin, A., Kuroki, T., Drevon, C., Piccoli, C., and Montesano, R. Validation and comparative studies on 180 chemicals with Salmonella typhimurium strains and V79 Chinese hamster cells in the presence of various metabolizing systems. Mutat. Res. 76: 1–50 (1980).
- 477. Sugimura, T., Sato, S., Nagao, M., Yahagi, T., Matsushima, T., Seino, Y., Takechi, M., and Kawachi, T. Overlapping of carcinogens and mutagens. In: Fundamentals in Cancer Prevention (P. N. Magee et al., Eds.), University of Tokyo Press, Tokyo, 1976.
- 478. Purchase, I. F. H., Longstaff, E., Ashby, J., Styles, J. A., Anderson, D., Lefevre, P. A., and Westwood, F. R. An evaluation of six short-term tests for detecting organic chemical carcinogens. Brit. J. Cancer 37: 873-959 (1978).
- 479. Ashby, J., de Serres, F. J., Draper, M. H., Ishidate, M., Matter, B. E., and Shelby, M. The two IPCS collaborative studies on short-term tests for genotoxicity and carcinogenicity. Mutat. Res. 109: 123–126 (1983).
- 480. Waters, M., Sandhu, S., Simmon, V., Mortelmans, K., Mitchell, A., Jorgenson, T., Jones, D., Valencia, R., and Garret, D. Study of pesticide genotoxicity. In: Genetic Toxicology and Agricultural Perspectives (Basic Life Sciences, Vol 21) (R. A. Fleck and A. Hollaender, Eds.), Plenum Press, New York, 1982.

- 481. Sobels, F. H. Editorial introduction to papers produced by the new committee 1 of ICPEMC on "The development and implementation of a scheme to analyze and interpret short-term genetic test battery results." Mutat. Res. 115: 175-176 (1983).
- 482. Brusick, D. Unified scoring system and activity definitions for results from *in vitro* and submammalian mutagenesis test batteries. In: Health Risk Analysis: Proceedings of the Third Life Sciences Symposium. (C. R. Richmond, P. J. Walsh, and E. D. Copenhaver, Eds.) The Franklin Institute Press, Philadelphia, 1981.
- 483. Squire, R. A. Ranking animal carcinogens: a proposed regulatory approach. Science 214: 877-880 (1981).
- 484. Weisburger, J. H., and Williams, G. A. Carcinogen testing: current problems and new approaches. Science 214: 401-407 (1981).
- Berenblum, I. Early studies of carcinogenesis. In: Carcinogenesis as a Biological Problem, North-Holland Press, Amsterdam, 1974, pp. 1–66.
- Pitot, H. C. Fundamentals of Oncology, 2nd Ed., Marcel Dekker, New York, 1981.
- 487. Weisburger, E. K. History of the bioassay program of the National Cancer Institute. Prog. Exp. Tumor Res. 26: 187–201 (1983).
- Chu, K. C., Cueto, C., and Ward, J. M. Factors in the evaluation of 200 National Cancer Institute carcinogen bioassays. J. Toxicol. Environ. Health 8: 251–280 (1981).
- 489. Griesemer, R. A., and Cueto, C. Toward a classification scheme for degrees of experimental evidence for the carcinogenicity of chemicals for animals. In: Molecular and Cellular Aspects of Carcinogen Screening Tests (R. Montesano, H. Bartsch, and L. Tomatis, Eds.), IARC Scientific Publ., No. 27, Lyon, France, 1980, pp. 259-281.
- 490. Pershagin, G., Nordbert, G., and Bjorklung, N. E. Carcinomas of the respiratory tract in hamsters given arsenic trioxide and/ or benzo(a)pyrene by the pulmonary route. Environ. Res. 34: 227-241 (1984).
- 491. IARC. Polynuclear Aromatic Compounds, Part 2, Carbon Blacks, Mineral Oil and Some Nitroarenes. IARC Sci. Publ. No. 33, Lyon, France, 1984, p. 19.
- 492. WHO. Principles for the Testing and Evaluation of Drugs for Carcinogenicity. WHO Technical Report Series No. 426, WHO, Geneva, 1969.
- 493. FDA. Advisory Committee on Protocols for Safety Evaluation, Panel on Carcinogenesis. Report on cancer testing in the safety of food additives and pesticides. Toxicol. Appl. Pharmacol. 20: 419–438 (1971).
- 494. NCI. Carcinogenesis Bioassay Program, Division of Cancer Control and Prevention, United States National Cancer Institute. Guidelines for Carcinogen Bioassay in Small Rodents. Bethesda, MD, 1974.
- NAS. Principles and Procedures for Evaluating the Toxicity of Household Substances. National Academy of Sciences, Washington, DC, 1977.
- 496. WHO. Environmental Health Criteria 6, Principles and Methods for Evaluating the Toxicity of Chemicals, Part I. WHO Geneva, 1978.
- Food Safety Council, Chronic Toxicity Testing. Proposed system for food safety assessment. Food Cosmet. Toxicol. 16 (Suppl 2): 97-108 (1978)
- Interagency Regulatory Liaison Group, Work Group on Risk Assessment. Scientific bases for identification of potential carcinogens and estimation of risks. J. Natl. Cancer Inst. 63: 242– 268 (1979).
- 499. IARC. Report 1: Basic requirements for long-term assays for carcinogenicity. In: Long-Term and Short-Term Screening Assays for Carcinogens: A Critical Appraisal. IARC Monograph Series, Supplement 2, Lyon, 1980, pp. 21–84.
- 500. Health and Environmental Studies Program (HESP), Oak Ridge National Laboratory. Scientific Rationale for the Selection of Toxicity Testing Methods: Human Health Assessment. ORNL/ EIS-151, EPA-560/1-80-001 (1980).
- 501. Committee on Carcinogenicity of Chemicals in Food, Consumer Products and the Environment. Guidelines of the Testing of Chemicals for Carcinogenicity. Report on Health and Social Sub-

- jects, #25. Department of Health and Social Security, London, 1982.
- 502. Sontag, J. M. Aspects in carcinogen bioassay. In: Origins of Human Cancer (H. Hiatt, J. Watson and J. Winsten, Eds.), Book C, Cold Spring Harbor, 1977, pp. 1327-1338.
- 503. Board of Scientific Counselors, National Toxicology Program.
 Report of the NTP Ad Hoc Panel on Chemical Carcinogenesis
 Testing and Evaluation. DHHS, August 17, 1984.
 504. Zwickey, R. E., and Davis, K. J. Carcinogenicity screening. In:
- 504. Zwickey, R. E., and Davis, K. J. Carcinogenicity screening. In: Appraisal of the Safety of Chemicals in Foods, Drugs, and Cosmetics. FDA, DHEW. Washington DC, 1959, pp. 79–82.
- 505. Shimkin, M., and Triolo, V. A. History of chemical carcinogenesis: some prospective remarks. Progr. Exptl. Tumor Res. 11: 1–20 (1969).
- 506. Shimkin, M. B. Report of the discussion group No. 1: species and strain selection. In: Carcinogenesis Testing of Chemicals (L. Goldberg, Ed.), CRC Press, Cleveland, 1974, p. 15.
- 507. Shubik, P. Use of the Syrian golden hamster in chronic toxicity testing. Progr. Exptl. Tumor Res. 16: 176–184 (1972).
- 508. Novell, M. J., and Wolff, G. L. (Eds.). NCTR/NCI symposium on the use of imbred and outbred animals in toxicological testing. J. Toxicol. Environ. Health 5: 1–169 (1979).
- 509. Festing, M. F. W. Inbred Strains and Factorial Experimental Design in Toxicological Screening. Utrecht, Holland: ICLA Symposium (1979).
- 510. Huff, J. Carcinogenesis bioassay results from the National Toxicology Program. Environ. Health Perspect. 45: 185–198 (1982).
- 511. Ward, J. M., Griesemer, R. A., and Weisburger, E. K. The mouse liver tumor as an endpoint in carcinogenesis tests. Toxicol. Appl. Pharmacol. 51: 389-397 (1979).
- 512. Newberne, P. M. Assessment of the hepatocarcinogenic potential of chemicals: response of the liver. In: Toxicology of the Liver (G. Plaa and W. R. Hewitt, Eds.), Raven Press, New York, 1982.
- Vesselinovitch, S. D. Liver tumor induction. Toxicol. Pathol. 10: 110-118 (1982).
- 514. von Wittenau, M. S., and Estes, P. C. The redundancy of mouse carcinogenicity bioassays. Fund. Appl. Toxicol. 3: 361-369 (1983)
- 515. Peto, R., Pike, M., Day, N., Gray, R., Lee, P., Parish, S., Peto, J., Richard, S, and Wahrendorf, J. Guideline for simple, sensitive, significant tests for carcinogenic effects in long-term animal experiments. In: Long-term and Short-term Screening Assays for Carcinogens. A Critical Appraisal (IARC Monograph Series Suppl. 2), IARC, Lyon, 1980, pp. 311-426.
- 516. FDA. Good Laboratory Practice for Nonclinical Laboratory Studies, Title 21, CFR, Part 58, p. 203, revised as of April 1982.
- 517. Munro, I. C. Considerations in chronic toxicity testing: the chemical, the dose, the design. J. Environ. Pathol. Toxicol. 1: 183–197 (1977).
- 518. Kraft, P. L., and Bieber, M. A. The use of dietary fats in animal studies. Paper presented at Spring Symposium, Mid-Atlaqntic Chapter of the Society of Toxicology, Wilmington, DE, May 21, 1983.
- Gehring, P. J., and Blau, G. E. Mechanisms of carcinogenesis: dose-response. J. Environ. Pathol. Toxicol. 1: 163-179 (1977).
- 520. Gehring, P. J., Wantanabe, P. G., and Park, C. N. Resolution of dose-response toxicity data for chemicals requiring metabolic activation: example—vinyl chloride. Toxicol. Appl. Pharmacol. 44: 581-591 (1978).
- 521. Ramsey, J. C., and Gehring, P. J. The integration and applications of pharmacokinetic principles in realistic estimates of risk. In: Health Risk Analysis, Proceedings of the Third Life Sciences Symposium (C. R. Richmond, P. J. Walsh, and E. D. Copenhaver, Eds.), Franklin Institute Press, Philadelphia, 1981.
- 522. Tsuchiya, T., and Levy, G. Relationship between dose and plateau level of drugs eliminated by parallel first-order and capacity-limited kinetics. J. Pharm. Sci. 61: 541–544 (1972).
- 523. O'Flaherty, E. J. Toxicants and Drugs: Kinetics and Dynamics. Wiley and Sons, New York, 1981.
- 524. Gibaldi, M., and Pervier, D. Pharmacokinetics, 2nd Ed., M. Dekker, New York, 1982.

- Dayton, P. G., and Sanders, J. E. Dose-dependent pharmacokinetics: emphasis on phase 1 metabolism. Drug Metabol. Rev. 14: 347–405 (1983).
- 526. Ward. J. M. Background data and variations in tumor rates of control rats and mice. Prog. Exp. Tumor Res. 26: 241–258 (1983).
- 527. Solleveld, H. S., Haseman, J., and McConnell, E. E. The natural history of body weight gain, survival and neoplasia in the Fischer 344 rat. J. Natl. Cancer Inst. 72: 929-940 (1984).
- 528. Ward, J. M., and Reznik, G. Refinements of rodent pathology and the pathologists' contribution to evaluation of carcinogenesis bioassays. Progr. Exptl. Tumor Res. 26: 266-291 (1983).
- 529. Fears, T. R., and Douglas, J. F. Suggested procedures for reducing the pathology workload in a carcinogen bioassay program. Environ. Pathol. Toxicol. 1: 125-137 (1977).
- FDA. Report of the Chronic Toxicity and Carcinogenicity Panel. United States Food and Drug Administration, December 19, 1977
- National Cancer Advisory Board. Report of the Subcommittee on Environmental Carcinogenesis J. Natl. Cancer Inst. 58: 461– 465 (1977).
- 532. Special Problems with Toxicology Protocols. Proc. Toxicology Forum, February 1978, p. 86.
- 533. Berenblum, I. Historical perspective. In: Carcinogenesis, Vol. 2, Mechanisms of Tumor Promotion and Cocarcinogenesis (T. L. Slaga, A. Sivak, and R. K. Boutwell, Eds.), Raven Press, New York, 1978.
- 534. Ward, J. M., Goodman, D. G., Griesemer, R. A., Hardisty, J. F., Schueler, R. L., Squire, R. A., and Strandberg, J. D. Quality assurance for pathology in rodent carcinogenesis tests. J. Environ. Pathol. Toxicol. 2: 371-378 (1978).
- 535. International Expert Advisory Committee to the Nutrition Foundation. The Relevance of Mouse Liver Hepatoma to Human Carcinogenic Risk. Nutrition Foundation, Washington, DC, 1983
- 536. Gart, J. J., Chu, K. C., and Tarone, R. E. Statistical issues in interpretation of chronic bioassay tests for carcinogenicity. J. Natl. Cancer Inst. 62: 957–974 (1979).
- 537. Armitage, P. Tests for linear trends in proportions and frequencies. Biometrics 11: 375–386 (1955).
- Cox, D. R. The regression analysis of binary sequences (with discussion). J. Roy. Statist. Soc. B20: 215-242 (1958).
- 539. Breslow, N. A generalized Kruskal-Wallis test for comparing K samples subject to unequal patterns of censorship. Biometrika 57: 579-594 (1970).
- Cox, D. R. Regression models and life tables (with discussion).
 J. Roy. Statist. Soc. B34: 187–220 (1972).
- 541. Haseman, J. K. A re-examination of false positive rates for carcinogenesis studies. Fund. Appl. Toxicol. 3: 334-339 (1983).
- 542. Fears, T. R., Tarone, R. E., and Chu, K. C. False-positive and false-negative rates for carcinogenicity screens. Cancer Res. 37: 1941–1945 (1977).
- 543. Task Force of the Past Presidents of the Society of Toxicology. Animal data in hazard evaluation. Paths and pitfalls. Fund. Appl. Toxicol. 2: 101–107 (1982).
- 544. Dempster, A. P., Selwyn, M. D., and Weeks, B. J. Combining historical and randomized controls for assessing trends in proportions. J. Am. Statist. Assoc. 78: 221-227 (1983).
- 545. Hoel, D. G. Conditional two-sample tests with historical controls. In: Contributions to Statistics: Essays in Honor of Norman L. Johnson. (P. K. Sen, Ed.) North Holland Publishing Co., 1983.
- 546. Tarone, R. E. The use of historical control information in testing for a trend in proportion. Biometrics 38: 215–220 (1982).
- 547. MacMahon, B., and Pugh, T. F. Epidemiology: Principles and Methods. Little, Brown and Co., Boston, 1970.
- 548. Rothman, K. J. Causation and causal inference. In: Cancer Epidemiology and Prevention (D. Schottenfeld and J. F. Fraumeni, Jr., Eds.), W. B. Saunders, 1982, pp. 15–22.
- Doll, R., and Peto, R. The causes of cancer. J. Natl. Cancer. Inst. 66: 1191–1308 (1981).
- Fraumeni, J. F., Jr. Epidemiologic approaches to cancer etiology. Ann. Rev. Public Health 3: 85-100 (1982).
- 551. MacLure, R. M., and MacMahon, B. An epidemiologic perspec-

- tive of environmental carcinogenesis. Epidemiol. Rev. 2: 19-48 (1980).
- 552. Tomatis, L., Breslow, N. E., and Bartsch, H. Experimental studies in the assessment of cancer risk. In: Cancer Epidemiology and Prevention (D. Schottenfeld and J. F. Fraumeni, Jr., Eds.) W. B. Saunders, Philadelphia, 1982, pp. 44-73.
- 553. Day, N. E., and Brown, C. C. Multistage models and primary prevention of cancer. J. Natl. Cancer Inst. 64: 977-989 (1980).
- 554. Hutchison, G. B. The epidemiologic method. In: Cancer Epidemiology and Prevention (D. Schottenfeld and J. F. Fraumeni, Jr., Eds.), W. B. Saunders, 1982, pp. 3-14.
- Lilienfeld, A., Pederson, E., and Dowd, J. E. Cancer Epidemiology: Methods of Study. Johns Hopkins Press, Baltimore, 1967
- $556.\;$ Doll, R. The epidemiology of cancer. Cancer 45: 2475–2485 (1980).
- 557. Austin, P. F., and Roe, K. M. The decreasing incidence of endometrial cancer: public health implications. Am. J. Publ. Health 72: 65-68 (1982).
- 558. Cole, P. Introduction. The analysis of case-control studies: In: Statistical Methods in Cancer Research: Vol. 1 (N. E. Breslow and N. E. Day, Eds.), IARC, Lyon, 1980, pp. 14–40.
- 559. Perera, F. P., and Weinstein, I. B. Molecular epidemiology and carcinogen DNA adduct detection: new approaches to studies of human cancer causation. J. Chron. Dis. 35: 581-600 (1982).
- 560. Hoover, R. N., and Strasser, P. H. Artificial sweetners and human bladder cancer. Lancet ii 837–840 (1980).
- 561. U.S. Environmental Protection Agency. Methodology for Assessing Occupational Exposure to Toxic Chemicals. Contract 68–01–6271, Task 10. Draft 6/18/82, appendices 4/7 and 4/7/82, Washington, DC, 1982.
- 562. U.S. Environmental Protection Agency. Methodology for Assessing Exposures from Disposal of Toxic Substances. Contract 68-01-6271, Task 11, Draft 5/28/82, Washington, DC, 1982.
- 563. U.S. Environmental Protection Agency. Methodology for Assessing Exposures to Toxic Chemicals in the Ambient Environment. Contract 68-01-6271, Task 13, Draft 5/28/82, 4 Vols., Washington, DC, 1982.
- 564. Task Force on Environmental Cancer and Heart and Lung Disease. Summary of the Workshop on Exposure to Environmental Agents, Their Metabolism and Mechanisms of Toxicity, Research Needs. Project Groups on Exposure and Metabolic Mechanisms. Report to Congress, Washington, DC, 1981.
- 565. Kang, H. K., and Infante, P. F. Preliminary risk assessment for asbestos. Occupational Safety and Health Administration Docket No. H-033.
- Hogan, M. D., and Hoel, D. G. Estimated cancer risk associated with occupational asbestos exposure. Risk Anal. 1: 67–76 (1981).
- 567. White, M. C., Infante, P. F., and Chu, K. C. A quantitative estimate of leukemia mortality associated with occupational exposure to benzene. Risk Anal. 2: 195-204 (1982).
- 568. OSHA. Occupational exposure to inorganic arsenic; Supplemental statement of reasons for final rule Fed. Reg. 48: 1864–1903 (1983).
- 569. Jones, M. L., and Saito, E. C. Supporting document: Occupational exposure to Toxaphene, assumptions, use patterns and calculations. Office of Pesticide Programs, U. S. Environmental Protection Agency, Washington, DC, 1982.
- Office of Pesticide Programs. Toxaphene: Decision Document.
 U.S. Environmental Protection Agency, Washington, DC, 1982.
- 571. Severn, D. J. Use of exposure data for risk assessment. Paper presented at Symposium on Determination and Assessment of Pesticide Exposure, Hershey, PA, October 29, 1980.
- 572. Jensen, J. K. The assumptions used for exposure assessment. Paper presented at Symposium on Determination and Assessment of Pesticides Exposures, Hershey, PA, October 30, 1980.
- 573. Severn, D. J. Exposure assessment for agricultural chemicals. In: Genetic Toxicology: An Agricultural Perspective. Plenum Press, New York, 1982, pp. 235-242.
- 574. Paynter, O. E., Cummings, J. G., and Rogoff, M. H. United States Pesticide Tolerance System. Office of Pesticide Programs, U.S. Environmental Protection Agency, Washington, DC, 1983.
- 575. Arthur D. Little, Inc. A study of indirect food additive migration. Unpublished data obtained under U.S. FDA Contract 223–

- 77-2360, Washington, DC.
- 576. U.S. Food and Drug Administration. Procedures for estimating exposure to indirect food additives. Unpublished guidelines, Washington, DC 1982.
- 577. Beloian, A. Use of a food consumption model to estimate human contaminant intake. Environ. Monitoring Assessment 2: 115–127 (1982).
- 578. Robert Williams Technical and Economic Service, Inc. Packaging in 1980 materials and markets. Unpublished data obtained under U.S. FDA Contract 22177-0195, Washington, DC (1980).
- 579. Schwartz, P. S., Schroeder, L. W., and McKay, T. J. Indirect additives—exposure estimates—II. Unpublished report, U.S. Food and Drug Administration, Washington, DC, 1980.
 580. C. H. Kline and Co., Inc. Plastic packaging 1979. Unpublished
- C. H. Kline and Co., Inc. Plastic packaging 1979. Unpublished report, U.S. Food and Drug Administration, Washington, DC, 1979.
- Arthur D. Little, Inc. Food contact polymers and polymer additives used in packaging. Unpublished data obtained under U.S. FDA Contract 223-777-2360, Mod. No. 2, Washington, DC, 1979.
- 582. Abrams, I. J. Access to menu census VI data from July 1, 1977 –June 30, 1978. Unpublished data obtained under U.S. FDA Contract 223–77–2046, Market Research Corporation of America, Chicago, 1978.
- 583. Committee on GRAS List Survey—Phase III, Food and Nutrition Board, NAS-NRC. The 1977 survey of industry on the use of food additives. Food and Drug Administration, Washington, DC, 1979.
- 584. Committee on GRAS List Survey—Phase III, Food and Nutrition Board, NAS-NRC. Estimating distribution of daily intakes of certain GRAS Substances. Food and Drug Administration, Washington, DC, 1976.
- 585. Subcommittee on Review of the GRAS List (Phase II), Food Protection Committee, NAS-NRC. A comprehensive survey of industry on the use of food chemicals Generally Recognized as Safe (GRAS). NTIS Report PB-221-949, Springfield, VA, 1973.
- Callahan, M. Planning and exposure assessment. Prepublication draft, Office of Toxic Substances, U. S. Environmental Protection Agency, Washington, DC, 1982.
- 587. Versar Inc. Methods for assessing exposure to chemical substances. Prepared for the Office of Toxic Substances, US-EPA, Washington, DC, 1981.
- 588. Office of Technology Assessment. Assessment of technologies for determining carcinogenic risks from the environment. Congress of the United States, Pub. No. OTA-H-138, Washington, DC, 1981.
- 589. American Industrial Health Council. Chronic health hazards: Carcinogenesis, mutagenesis, teratogenesis, a framework for sound science in Federal decision making. Scarsdale, New York, 1981.
- Davis, D. L., and Gusman, S. Exposure assessment: new frontier, old problems. Toxic Substances J. 4: 3 (1982).
- Davis, D. L., and Gusman, S. Exposure assessment introduction. Toxic Substances J. 4: 4-11 (1982).
- 592. Wallace, L. A. Recent progress in developing and using personal monitors to measure human exposure to air pollutants. Environ. International 5: 73–75 (1981).
- 593. Wallace, L. A., and Ott, W. Personal monitors: A state-of-art survey. J. Air. Poll. Control Assoc. 32: 601-610 (1982).
- 594. Environmental Monitoring Systems Laboratory. Interim report of field activities pertaining to the measurement of carbon monoxide exposure of residents of Washington, D.C., and Denver, Colorado. Research Triangle Park, NC, 1983.
- 595. Ott, W., Blacker, S., and Akland, G. Research plan for population exposure monitoring methodology: Vehicular air pollutants. U.S. Environmental Protection Agency, Washington, DC, 1981.
- 596. Flachsbart, P. G. Field survey procedures for measuring carbon monoxide exposures of commuters in the Washington Metropolitan Area. Report under Cooperative Agreement No. CR-810344-01-0, U.S. Environmental Protection Agency, Environmental Monitoring Systems Laboratory, Research Triangle Park, NC, 1982.
- 597. Flachsbart, P. G., and Ott, W. R. Field surveys of carbon mon-

- oxide in commercial settings using personal exposure monitors. Draft report, U.S. Environmental Protection Agency, Office of Research and Development, July 1981.
- 598. U.S. Environmental Monitoring Systems Laboratory. Carbon monoxide concentrations in four U.S. cities during the winter of 1981. Research Triangle Park, NC, draft report, 1983.
- 599. Ott, W. R., and Willits, N. H. CO exposures of occupants of motor vehicles: Modeling the dynamic response of the vehicle. SIMS Technical Report No. 48, Stanford University, Department of Statistics, Stanford, CA.
- 600. Akland, G. G. CO exposures in Washington, D.C. and Denver, Colorado. Draft report, U.S. Environmental Protection Agency, Environmental Monitoring Systems Laboratory, Research Triangle Park, NC, 1983.
- Holland, D. M. Carbon monoxide levels in microenvironment types of four U.S. cities. Environ. International (submitted).
- 602. Schiermeier, F. A. Air monitoring milestones: RAPS field measurements are in. Environ. Sci. Technol. 12: 644-651 (1978).
- 603. Strothmann, J. A., and Schiermeier, F. A. Documentation of the regional air pollution study (RAPS) and related investigations in the St. Louis air quality region. U.S. Environmental Protection Agency, Pub. No. EPA-600/4-0761, Washington, DC, 1979
- 604. Schiermeir, F. A., Wilson, W. E., Poolee, F., Ching, J. K. S., and Clarke, J. F. Sulfur transport and transformation in the environment (STATE): a major EPA research program. Bull. Am. Meteor. Soc. 60: 1303-1312 (1979).
- 605. Possiel, N. C., Wilson, W. E., Pooler, F., Ching, T. K. S., and Clarke, J. F. Recent EPA urban and regional scale oxidant field programs in the northeastern U.S. Paper presented at Annual Meeting of the Air Pollution Control Association, New Orleans, LA, June 20-25, 1982.
- 606. Glass, G. E., Strachan, W., Willford, W., Armstrong, F. A., and Kaiser, K. L. Organic contaminants. In: The Waters of Lake Huron and Lake Superior, Vol. III (Part B). Lake Superior. Report to the International Joint Commission by the Upper Great Lakes Reference Group, Windsor, Ontario, 1977, EPA 600-15-77/042, pp. 417-429 and 499-502.
- 607. Poldoski, J. E., and Glass, G. E. Methodological considerations in Western Lake Superior Water-Sediment exchange studies of some trace elements. In: National Bureau of Standards Special Publication 422, Washington, DC, 1976, pp. 1073–1088.
- Publication 422, Washington, DC, 1976, pp. 1073-1088.

 608. Veith, G. D., Kuehl, D. W., Puglisi, F. A., Glass, G. E., and Eaton, J. G. Residues of PCBs and DDT in the Western Lake Superior ecosystem. Arch. Environ. Contam. Toxicol. 5: 487-499 (1977).
- 609. Welch, K. J., Kuehl, D. W., Leonard, E. L., Vieth, G. D., and Schoenthal, N. D. Background hydrocarbon residues in fishes from the Great Lakes and eastern Montana. Bull. Environ. Contam. Toxicol. 26: 724–728 (1981).
- 610. McNaught, D. C., Kuehl, D. W., and Leonard, E. N. Proceedings of the Symposium on atmospheric inputs of pollutants to the Great Lakes. Great Lakes Res. 8: 239–375 (1982).
- the Great Lakes. Great Lakes Res. 8: 239-375 (1982).
 611. Callaway, R. J., Vieth, G. D., and Schoenthal, N. D. Preliminary analysis of the dispersion of sewage sludge discharged from vessels to New York Bight waters. Draft report, Environmental Protection Agency, Corvallis, OR, 1982.
- 612. Callaway, R. J. Flushing study of South Beach Marina, Oregon. J. Waterway, Port, Coastal and Ocean Division, ASCE, 107 (WWZ), Proc. Paper 16265; 1981, pp. 47-58.
 613. Swartz, R. C., and Lee, H., II. Biological processes affecting
- 613. Swartz, R. C., and Lee, H., II. Biological processes affecting the distribution of pollutants in marine biodegradation and migration. In: Contaminants and Sediments, Vol 2. Ann Arbor Science Publishers, Ann Arbor, 1980 pp. 533–553.
- 614. Lee, H., II, and Swartz, R. C. Biological processes affecting the distribution of pollutants in marine sediments. Part II. Biodeposition and bioturbation. In: Contaminants and Sediments, Vol 2. Ann Arbor Science Publishers, Ann Arbor, 1980, pp. 555-606.
- 615. Gossett, R. W., et al. DDT, PCB, and benzo(a)pyrene levels in white croaker (*Genyonemus lineatus*) from Southern California. Marine Poll. Bull., in press.
- 616. Puffer, H. W., et al. Consumption rates of potentially hazardous marine fish catch in the Metropolitan Los Angeles area. Final

- report on EPA Grant no. 807–120010, Environmental Research Laboratory, U.S. Environmental Protection Agency, Corvallis, OR, 1981.
- 617. Veith, G. D., et al. Fish, wildlife and estuaries. Pestic. Monit. J., 13: 1-11 (1979).
- Keeley, J. W. Guest editorial: New directions in international groundwater research. Groundwater 20: 138-141 (1982).
- 619. Wilson, J. T., Piwoni, M. D., and Dunlap, W. J. Transport and fate of organic pollutants in the subsurface environment. Prepublication draft, Groundwater Research Branch, Robert S. Kerr Environmental Research Laboratory, U.S. Environmental Protection Agency, Ada, OK, 1982.
- Fausett, R. S., and Pressler, C. L. Kerosene heaters: Project status report and staff recommendations. US-CPSC, Washington, DC, 1983.
- Eberle, S. Indoor air quality: Fuel-fired appliances; Briefing materials on unvented gas space heaters. US-CPSC, Washington, DC, 1983.
- National Academy of Sciences Committee on Indoor Pollutants. Indoor Pollutants. National Academy Press, Washington, DC, 1981
- 623. Ott, W. R. Human activity patterns: A review of the literature for estimation of exposures to air pollution. U. S. Environmental Protection Agency, Office of Research and Development, Draft Report (1982).
- 624. Spengler, J. D., Letz, R., Ozkaynak, H., and Soczek, M. L. Feasibility of predicting personal or population exposures utilizing ambient air quality models and human activity data. Final report for Project 1D6390NASA, Strategies and Standards Division, U.S. Environmental Protection Agency, Research Triangle Park, N.C.
- 625. Wallace, L., Zweidinger, R., Erickson, M., Cooper, S., Whitaker, D., and Pellizzari, E. Monitoring individual exposure-measurements of volatile organic compounds in breathing zone air, drinking water and exhaled breath. Environ. Intern. 8: 269–282 (1982).
- 626. Wallace, L., Pellizzari, E., and Hartwell, T., Rosenweig, M., Sparacino, C., and Zelon, H. Individual human exposure to volatile organic compounds encountered during normal daily activities. Work Plan, Office of Research and Development, U.S. Environmental Protection Agency, Washington, DC, 1982.
- 627. Wallace, L. A., Pellizzari, E., Hartwell, T., Rosenzweig, M., Erickson, M., Sparacino, C., and Zelon, H. Personal breathing zone air, drinking water, food and exhaled breath. Environ. Res. (submitted).
- 628. Pellizzari, E. D., Erickson, M. D., Sparacino, C. M., Hartwell, T. D., Zelon, H., Rosenzweig, M., Leininger, C., Blau, P., and Giguere, M. Total exposure assessment methodology (TEAM) study, Phase II, Part I. Formulation of an exposure and body burden monitoring program. Work Plan, Office of Research and Development, U.S. Environmental Protection Agency, Washington, DC, 1982.
- 629. Pellizzari, E. D., Erickson, M. D., Giguere, M. T., Hartwell, T. D., Handy, R. W., Williams, S. R., Sparacino, C. M., Zelon, H., and Waddell, R. W. Study on toxic chemicals in environmental and human samples, Part II. Protocols for environmental and human sampling and analysis. Work Plan, Office of Research and Development, U.S. Environmental Protection Agency, Washington, DC, 1982.
- 630. Handy, R.W., and Pellizzari, E. D. Total exposure assessment methodology (TEAM) study, Phase II, Part III. Quality Assurance Project Plan Draft Work Plan. Office of Research and Development, U.S. Environmental Protection Agency, Washington, DC, 1982.
- 631. Kutz, F. W., Murphy, R. S., and Strassman, S. C. Survey of pesticide residues and their metabolites in urine from the general population. In: Pentachlorophenol (K. R. Rao, Ed.), Plenum Press, New York, 1978, pp. 363–369.
- 632. National Center for Health Statistics. Plan and operation of the Second National Health and Nutrition Examination Survey, 1976–1980. Vital and Health Statistics, Series 1, No. 15, PHS Pub. No. 81–1317, Public Health Service, U.S. Government Printing Office, Washington, DC, 1981.

- 633. Murphy, R. S., Kutz, F. W., and Strassman, S. C. Selected pesticide residues or metabolites in blood and urine specimens from a general population survey. Proceedings of the Conference on Research Needs for Evaluation of Health Effects on Toxic Chemical Waste Dumps, National Institute of Environmental Health Sciences, Research Triangle Park, NC, October 27–28, 1981.
- 634. Kutz, F. W., Yobs, A. R., Strassman, S. C., and Viar, J. F. Jr. Pesticides in people. Pestic. Monit. J. 11: 61-63 (1977).
- 635. Lucas, R. M., Iannocchione, V. G., and Melroy, D. K. Polychlorinated biphenyls in human adipose tissue and mother's milk. Final report under contract number 6801–5848, U.S. Environmental Protection Agency, Washington, DC, 1982.
- mental Protection Agency, Washington, DC, 1982.
 636. Berlin, A., Yodaiken, R. E., and Henman, B. A. (Eds.). Assessments of Toxic Agents in the Workplace: Roles of Ambient and Biological Monitoring. Nijhoff, Netherlands, 1984.
- 637. Lauwerys, R. R. (Ed.). Industrial Chemical Exposure: Guidelines for Biological Monitoring. Biomedical Press, Foster City, California, 1983.
- 638. Aitio, A., Riihimaki, V., and Vainio, H. (Eds.). Biological Monitoring and Surveillance of Workers Exposed to Chemicals. Hemisphere Press, Washington, DC, 1983.
- 639. Miller, S. A monitoring report. Environ. Sci. Technol. 17: 343A–346A (1983).
- 640. U.S. Environmental Protection Agency. EPA Environmental Modeling Catalogue. Draft report, prepared under Contract No. 68-07-4723, Washington, DC, 1982.
- 641. U.S. Environmental Protection Agency. EPA Environmental Data Base and Model Index, Index Summary. EPA Information Clearinghouse, Office of Planning and Management, Washington, DC, 1981.
- 642. U.S. Environmental Protection Agency. Models Survey, Questionnaire. EPA Form 3700-1A, EPA Information Clearinghouse, Office of Planning and Management, Washington, DC, 1980.
- 643. Whitten, G. Z., Hugh, H., and Kilus, J. P. The carbon bond mechanism for photochemical smog. Environ. Sci. Technol. 14: 690-700 (1980).
- 644. Eschenroeder, A. Dynamic modeling for assessment of exposure. Toxic Substances J. 4: 38-54 (1982).
- 645. Goodin, W. R. Advanced air quality modeling. Engin. Bull. 58: 21–36 (1981).
- 646. Shreffler, J. H., and Schere, K. L. Evaluation of four urbanscale photochemical air quality simulation models. Draft report, Meteorology and Assessment Division, Environmental Sciences Research Laboratory, U. S. Environmental Protection Agency, Research Triangle Park, NC, 1982.
- 647. Schere, K. L., and Shreffler, J. H. Final evaluation of urbanscale photochemical air quality simulation models. Meteorology and Assessment Division, Environmental Sciences Research Laboratory, U.S. Environmental Protection Agency, Research Triangle Park, NC, 1982.
- 648. Hendry, D. G., and Kenley, R. A. Atmospheric reaction products of organic compounds. SRI International, EPA Contract No. 68-01-5123, Report No. EPA-560/12-79-001, U. S. Environmental Protection Agency, Washington, DC (1979).
- 649. Lamb, R. G. A regional scale (100km) model photochemical air polution, Part I: Theoretical formulation. Draft report, Meteorology and Assessment Division, Environmental Sciences Research Laboratory, U.S. Environmental Protection Agency, Research Triangle Park, NC, 1982.
- 650. Leavery, T. F., et al. EPA complex terrain model development. Final Report, EPA Contract No. 68-02-3421, Report No. EPA-600/3-82-036, U.S. Environmental Protection Agency, Washington, DC (1982).
- National Academy of Science, National Research Council. Indoor Pollutants. U.S. Environmental Protection Agency, Pub. No. EPA-600/6-82-001, NTIS Pub. No. PB82180563, Washington, DC (1982).
- 652. Paul, J. F., and Patterson, R. L. Hydrodynamic simulation of movement of larval fishes in Western Lake Erie and their vulnerability to power plant entrainment. In: Proceeding of the 1977 Winter Simulation Conference, Gaithersburg, MD, Dec. 5-7, 1977 (H. J. Highland, et al., Eds.), National Bureau of Stan-

- dards, 1979.
- 653. Bierman, V., Jr., and Dolan, D. M. Modeling of phytoplanktonnutrient dynamics in Saginaw Bay, Lake Huron. J. Great Lakes Res. 7: 409-439 (1981).
- 654. Bierman, V. J., and Swain, W. R. Mass balance modeling of DDT dynamics in Lakes Michigan and Superior. Environ. Sci. Technol. 16: 572-578 (1982).
- 655. Paul, J. F., Kasprzyk, R., and Lick, W. Turbidity in the Western Basin of Lake Erie. J. Geophys. Res., 87(C8): 5779-5784 (1982).
- 656. Dolan, D. M., and Bierman, V. J., Jr. Mass balance modeling of heavy metal in Saginaw Bay, Lake Huron. J. Great Lakes Res. 8: 676-694 (1982).
- 657. Ingfle, S. E., Kenisten, J. A., and Schults, D. W. REDEQL-EPAK, Aqueous Chemical Equilibrium Computer Program. Report No. EPA-600/3-80-049, Environmental Research Laboratory, U.S. Environmental Protection Agency, Corvallis, OR, 1980.
- 658. Burnes, L. A., Cline, D. M., and Lassiter, R. R. Exposure Analysis Modeling System (EXAMS): User manual and system documentation. Pub. No. EPA-600/2-82-023, U.S. Environmental Protection Agency, Washington, DC, 1982.
- 659. Callaway, R. J. Surface horizontal dispersion of pollutants in open coastal waters. Paper presented at International Symposium on Discharge of Sewage from Sea Outfalls, London, 2 Sept. 1974, HM 6661 Dd 196552 600 6/74 McC3309, Her Majesty's Stationary Office, McCorguodale Printers Ltd, London, 1974.
- 660. Lick, W., Paul, J., and Sheug, Y. P. The dispersion of contaminants in the near-shore region. Modeling Biochemical Process in Aquatic Ecosystems. Ann Arbor Science Publishers, Ann Arbor, 1976, pp. 93-111.
- 661. Teeter, A. M., and Baumgartner, D. J. Prediction of initial mixing for municipal ocean discharges. Environmental Research Laboratory, U.S. Environmental Protection Agency, Corvallis, OR. 1979.
- 662. Overton, W. S. A model for terrestrial exposure assessment. Draft annual report, Environmental Research Laboratory, U.S. Environmental Protection Agency, Corvallis, OR, 1982.
- 663. INTERA Environmental Consultants, Inc. Hydrologic modeling of aldicarb transport to ground water on Long Island. Prepared for South Carolina Pesticide Epidemiology Study Center, Medical University of South Carolina, Houston, Texas, 1980.
- 664. Wilson, J. T., Enfield, C. G., Dunlop, W. J., Cosby, R. L., Foster, D. A., and Baskin, L. B. Transport and fate of selected organic pollutants in sandy soil. J. Environ. Qual. 10: 501-506 (1981).
- 665. Offutt, C. K., and Severn, D. J. Assessment and prediction of pesticides in groundwater. Workshop on Pesticides in Soil and Groundwater, Univ. of California, Davis, CA, June 15-16, 1982.
- 666. Enfield, C. G., Carsel, R. F., Cohen, S. Z., Phanm, T., and Walters, D. M. Approximating pollutant transport to groundwater. Ground Water 20: 711-722 (1982).
- 667. U.S. Environmental Protection Agency. Environmental fate summaries for 26 pesticides. Memo from Stuart Cohen to Peter McGrath, Dec. 1979, Office of Pesticides Programs, U.S. Environmental Protection Agency, Washington, DC.
 668. Fugav, M. Assessment of total exposure to an air pollutant.
- 668. Fugav, M. Assessment of total exposure to an air pollutant. Proceedings of the International Conference on Environmental Sensing and Assessment, Washington, DC, September 1975.
 669. Moschandreas, D. J., and Stark, J. W. The GEOMET indoor-
- 669. Moschandreas, D. J., and Stark, J. W. The GEOMET indooroutdoor air pollution model. Report GEOMET EF-628. U.S. Environmental Protection Agency 600/7-78/106, Washington, DC, 1978.
- 670. Wallace, L. I. Briefing papers on ORD activities in modeling, microenvironments and activity patterns, July, 1982, Office of Research and Development, U.S. Environmental Protection Agency, Washington, DC, 1982.
- 671. Falls, A. H., and Seinfeld, J. H. Continued development of a kinetic mechanism for photochemical smog. Environ. Sci. Technol. 12: 1398-1406 (1978).
- 672. Perez, K. T., et al. The importance of physical and biotic scaling to the experimental simulation of a coastal marine ecosystem. Helgolander Wiss. Meeresunters 30: 144–162 (1977).
- 673. U.S. Environmental Protection Agency. Experimental marine

- microcosm test protocol. An assessment of the ecological effects, fate and transport of chemicals in a site-specific marine ecosystem. Draft paper, Environmental Research Laboratory, Narragansett, RI, 1982.
- 674. Perez, K. T., and Dywer, R. L. An experimental examination of ecosystem linearization. Amer. Naturalist 21: 305-323 (1983).
- 675. Perez, K. T., et al. Environmental assessment of a phthalate ester, di-(2-ethylhexyl)phthalate (DEHP), derived from a marine microcosm. In: Aquatic Toxicology and Hazard Assessment: Sixth Symposium, Special Technical Pub. 802, American Society for Testing and Materials, Philadelphia, 1983.
- 676. Metcalf, R. L., et al. Design and evaluation of a terrestrial model ecosystem. Final report of EPA Cooperative Agreement R803249, Pub. No. 600/3-79-004, Environmental Research Laboratory, U.S. Environmental Protection Agency, Corvallis, OR, 1977.
- 677. Gile, J. D., and Gillett, J. W. Fate of selected fungicides in terrestrial laboratory ecosystem. J. Agr. Food Chem. 27: 1159–1164 (1979).
- 678. Gile, J. D., and Gillett, J. W. Fate of ¹⁴C Dieldrin in a simulated terrestrial ecosystem. Arch. Environ. Contam. Toxicol. 8: 107–124 (1979).
- 679. Gile, J. D., Collins, J. C., and Gillett, J. W. Fate of selected herbicides in a terrestrial laboratory microcosm. Environ. Sci. Technol. 14: 1124-1128 (1980).
- 680. Gile, J. D., Collins, J. C., and Gillett, J. W. Fate and impact of wood preservatives in a terrestrial microcosm. J. Agr. Food Chem. 30: 295-301 (1982).
- 681. Goodman, E. D., et al. Ecosystem responses to alternative pesticides in the terrestrial environment: A system approach. Final report of EPA Coop. Agreement R805624, Michigan State University, East Lansing, MI, 1982.
- 682. Gile, J. D. 2,4-D—Its distribution and effects in a ryegrass ecosystem. Draft report, Environmental Research Laboratory, U.S. Environmental Protection Agency, Corvallis, OR, 1982.
- 683. Gile, J. D., and Gillett, J. W. Transport and fate of organophosphate insecticides in a laboratory model ecosystem. J. Agr. Food Chem. 29: 616-621 (1981).
- 684. Gile, J. D. Research brief, biological effects and interactions of pesticides in a soil-plant-water microcosm. Unpublished, Environmental Research Laboratory, U.S. Environmental Protection Agency, Corvallis, OR, 1982.
- 685. Eaton, J. G. Recent developments in the use of laboratory bioassays to determine 'safe' levels of toxicants for fish. Bioassay Techniques and Environmental Chemistry. Ann Arbor Science Publishers. Ann Arbor, 1983.
- 686. Brungs, W. A., McCormick, J. H., Nieheisel, T. W., Spehar, R. L., Stephen, C. E., and Stokes, G. N. Effects of pollution on freshwater fish. J. Water Poll. Cont. Fed. 49: 1415-1493 (1977).
- 687. McKin, J. M., Eaton, J. G., and Holcombe, G. W. Metal toxicity to embryos and larvae of eight species of fresh water fish—II: copper. Bull. Environ. Contam. Toxicol. 19: 608-616 (1978).
- 688. Kopperman, H. L., Kuehl, D. W., and Glass, G. E. Chlorinated compounds found in waste-treatment effluents and their capacity to bioaccumulate. In: Proceedings of the Conference on the Environmental Impact of Water Chlorination, Oak Ridge, Tennessee, Oct. 22–24, 1975 (R. L. Jolley, Ed.), U.S. Government Printing Office, Pub. No. 1976669979/196, Washington, DC, 1975.
- 689. Wilson, J. T., and Nooran, M. J., and McNabb, J. F. Biodegradation of contaminants in the subsurface environment. Draft report, Robert S. Kerr Environmental Research Laboratory, Ada, OK, 1982.
- 690. Girman, J. R. Pollutant emission rates from indoor combustion appliances and cigarette smoke. Environ. Int. 8: 213–221 (1982).
- 691. Duan, N. Microenvironment types: A model for human exposure to air pollution, SIMS Technical Report No. 47, Department of Statistics, Stanford University, Stanford, CA, 1981.
- 692. Traynor, G. W. The effects of ventilation on residential air pollution due to emissions from a gas fired range. Lawrence Berkeley Laboratory, Pub. No. LBL-12563, Berkeley, CA, 1981.
- 693. Moschandreas, D. J. Indoor air pollution in the residential en-

- vironment. U.S. Environmental Protection Agency, Pub. No. EPA600/7-78-229a, Washington, DC, 1978.
- 694. Oser, B. L., and Hall, R. L. Criteria employed by the expert panel of FEMA for the GRAS evaluation of flavoring substances. Food Cosmet. Toxicol. 15: 457 (1977).
- 695. Lyman, W. S., Reehl, W. F., and Rosenblatt, D. H. (Eds.). Handbook of Chemical Property Estimation Methods Environmental Behavior of Organic Compounds. McGraw-Hill, New York, 1982.
- 696. Veith, G. D. Structure-activity research at the Environmental Research Laboratory—Duluth. Unpublished, Environmental Research Laboratory, U.S. Environmental Protection Agency, Duluth, MN, 1983.
- 697. Chiou, C. T., Schmedding, D. W., and Block, J. H. Correlation of water solubility with octanol-water partition coefficient. J. Pharmacol. Sci. 70: 1176-1177 (1981).
- 698. Chiou, C. T., and Schmedding, D. W. Partitioning of organic compounds in octanol-water systems. Environ. Sci. Technol. 16: 4-10 (1982).
- 699. Veith, G. D., Austin, N. M., and Morris, R. T. A rapid method for estimating log P for organic chemicals. Water Res. 13: 43– 47 (1979).
- 700. Veith, G. D., DeFoe, D. L., and Berg, B. V. Measuring and estimating the bioconcentration factor of chemicals in fish. J. Fish Res. Board Can. 36: 1040-1048 (1979).
- Veith, G. D., Call, D. J., and Brooke, L. T. Structure-toxicity relationships for the fathead minnow. I. Narcotic industrial chemicals. Unpublished, Environmental Research Laboratory, U.S. Environmental Protection Agency, Duluth, MN, 1982.
- 702. Enslein, K., Lander, T. R., Tomb, M. E., and Craig, P. N. A predictive model for estimating rat LD₅₀ values. A monograph prepared by Health Designs, Inc., Rochester, NY, 1983.
- 703. Versar, Inc. Draft Exposure Assessment for Formaldehyde. Prepared for Office of Toxic Substances, U.S. Environmental Protection Agency, Washington, DC, 1983.
- 704. TSPC Solvents Work Group #2. Draft Executive Summary for TSPC solvents exposure assessment. U.S. Environmental Protection Agency, Washington, DC, 1982.
- TSPC Solvents Work Group #2. Draft exposure assessment for TSPC solvents, U.S. Environmental Protection Agency, Washington, DC, 1982.
- U.S. Environmental Protection Agency. Permit Compliance System (PCS), Washington, DC, 1982.
- National Air Data Bank. AEROS Manual Series, Vol. 3, Summary and Retrieval. U.S. Environmental Protection Agency, Publ. No. EPA 450/2-76-0096, Washington, DC, 1981.
- 708. National Climatic Center, Index of Original Surface Weather Records (Hourly Synoptic and Autographic), 52 Vols., National Climatic Center, Asheville, NC.
- U.S. Geological Survey. Catalog of information on water data. Office of Water Data Coordination, Washington, DC, U.S. Environmental Protegion Agency. Organic Chemical Manufacturing, Pub. No. EPA-450/3-80-0238, Washington, DC, 1980.
- 710. Hall, L. H. The OTS Graphical Exposure Modeling System (GEMS). Prepublication draft, Office of Toxic Substances, U.S. Environmental Protection Agency, Washington, DC, 1982.
- Anderson, G. D. Modeling of Human Exposure to Airborne Toxic Materials. Paper presented at the American Chemical Society Meeting, Sept. 1982, Kansas City, MO, 1982.
- 712. Johnson, T., and Paul, R. A. The NAAQS Exposure Model (NEM) Applied to Carbon Monoxide. Draft, prepared for Strategies and Air Standards Division, Office of Air Quality Planning and Standards Division, Office of Air Quality Planning and Standards, U.S. Environmental Protection Agency, Research Triangle Park, NC.
- 713. Ott, W. R. Exposure estimates based on computer generated activity patterns. Paper No. 81-57.6, presented at the 74th Annual Meeting of the Air Pollution Control Association, Philadelphia, PA, June 21-26, 1981.
- 714. Cheng, C. K., Thomas, J., Ott, W., Mage, D., and Wallace, L. Sensitivity analysis of the enhanced Simulation of Human Air Pollution Exposure (SHAPE) Model. Project Report for Contract No. 68-01-6595, General Software Corporation, U.S. En-

- vironmental Protection Agency, Washington, DC, 1983.
- U.S. Environmental Protection Agency. Mandatory quality assurance program, administrative Book I. Internal File on Administrative Correspondence, Office of Research and Development, Washington, DC, 1982.
- U.S. Environmental Protection Agency. Mandatory Quality Assurance Program, QA Guidelines, Book II. Internal File on Correspondence, Office of Research and Development, Washington, DC. 1982.
- 717. Engineering-Science, Inc. CMA/EPA five-plant study. Prepared for Chemical Manufacturers Association, Washington, DC (1982).
- 718. U.S. Environmental Protection Agency. Environmental Monitoring at Love Canal, Vol. 1, Pub. No. EPA-600/4-82-038a, Washington, DC, 1982.
- 719. Mount, D. I. Present Approaches to Toxicity Testing—A Perspective. In: Aquatic Toxicology and Hazard Evaluation. Special Technical Publ. 634, American Society for Testing and Materials, Philadelphia, PA, 1977, pp. 5–14.
- 720. Brungs, W. A., and Mount, D. I. Introduction to a discussion of the use of aquatic toxicity tests for evaluation of effects of toxic substances. In: Aquatic Toxicology and Hazard Evaluation Special Technical Publ., Philadelphia, PA, 1978, pp. 15–26.
- 721. Mount, D. I. Margins of safety for aquatic communities. Aquatic Toxicology and Hazard Assessment: Fourth Conference. Special Technical Publ. 737, American Society for Testing and Materials, 1981, pp. 5–9.
- 722. Kross, R. D., and Lewis, S. C. Environmental analysis—Are we becoming too sensitive? Environ. Forum 1: 15-19 (1983).
- Food Safety Council. Proposed system for safety assessment.
 Food Safety Council, Washington, DC, 1980.
- 724. Interagency Regulatory Liaison Group. Scientific bases for identification of potential carcinogens and estimation of risks. Report written by the workgroup on risk assessment. Fed. Reg. 44: 39858–39879 (1979).
- 725. Hoel, D. G., Gaylor, D. W., Kirschstein, R. L., Saffiotti, U., and Schneiderman, M. A. Estimation of risks of irreversible delayed toxicity. J. Toxicol. Environ. Health 1: 133-151 (1975).
- 726. Gaylor, D. W., and Shapiro, R. E. Extrapolation and risk estimation for carcinogenesis. In: Advances in Modern Toxicology, Vol. 1, New Concepts in Safety Evaluation (part 2), (M. A. Mehlman, R. E. Shapiro, and H. Blumenthal, Eds.), NY: John Wiley and Sons, New York, 1979, pp. 65–87.
- 727. Gaylor, D. W., and Kodell, R. L. Linear interpolation algorithm for low-dose risk assessment of toxic substances. J. Environ. Pathol. Toxicol. 4: 305-312 (1980).
- 728. Van Ryzin, J. Quantitative risk assessment. J. Occup. Med. 22: 321–326 (1980).
- 729. Krewski, D., and Van Ryzin, J. Dose response models for quantal response toxicity data. In: Statistics and Related Topics (D. Csorgo, R. Dawson, and E. Saleh, Eds.), North Holland, Amsterdam, 1981, pp. 201–231.
- Mantel, N., and Bryan, W. R. "Safety" testing of carcinogenic agents. J. Natl. Cancer Inst. 27: 455–470 (1961).
- Mantel, N., Bohidar, N. R., Brown, C. C., Ciminera, J. L., and Tukey, J. W. An improved "Mantel-Bryan" procedure for "safety testing" of carcinogens. Cancer Res. 35: 865–872 (1975).
- 732. Crump, K. S. Response to open query: Theoretical problems in the modified Mantel-Bryan procedure. Biometrics 33: 752-757 (1977).
- 733. Armitage, P., and Doll, R. Stochastic models for carcinogenesis. In: Proceedings of the Fourth Berkeley Symposium on Mathematical Statistics and Probability (L. Lecam and J. Neyman, Eds.), University of California Press, Berkeley, CA, 1961, pp. 19–38.
- 734. Crump, K. S., Hoel, D. G., Langley, C. H., and Peto, R. Fundamental carcinogenic processes and their implications for low dose risk assessment. Cancer Res. 36: 2973–2979 (1976).
- Rai, K., and Van Ryzin, J. A. A generalized multihit doseresponse model for low-dose extrapolation. Biometrics 37: 341– 352 (1981).
- 736. Haseman, J. K., Hoel, D. G., and Jennrich, R. I. Some practical problems arising from the use of the gamma multihit model for

- risk estimation. J. Toxicol. Environ. Health 8: 379-386 (1976).
- 737. Whittemore, A., and Altschuler, B. Lung cancer incidence in cigarette smokers: further analysis of Doll and Hill's data for British physicians. Biometrics 32: 805-816 (1976).
- 738. Hartley, H. O., and Sielken, R. L. Estimation of "safe doses" in carcinogenic experiments. Biometrics 33: 1-30 (1977).
- 739. Krewski, D., Crump, K. S., Farmer, J., Gaylor, D. W., Howe, R., Portier, C., Salsburg, D., Sielken, R. L., and Van Ryzin, J. A comparison of statistical methods for low dose extrapolation utilizing time-to-tumor data. Fund. Appl. Toxicol. 3: 140-160 (1983).
- 740. ED_{01} Task Force of the Society of Toxicology. Re-examination of the ED_{01} study. Fund. Appl. Toxicol. 1: 28–128 (1981).
- Hoel, D. G. Statistical measures of risk. Drug Metab. Rev. 13: 829–838 (1982).
- Hoel, D. G. Incorporation of background in dose-response models. Fed. Proc. 39: 73-75 (1980).
- 743. Anderson, M. W., Hoel, D. G., and Kaplan, N. L. A general scheme for the incorporation of pharmacokinetics in low-dose risk estimation for chemical carcinogenesis: example—vinyl chloride. Toxicol. Appl. Pharmacol. 55: 154-161 (1980).
- 744. Hoel, D. G., Kaplan, N. L., and Anderson, M. W. The implication of nonlinear kinetics on risk estimation in carcinogenesis. Science 219: 1032–1037 (1983).
- 745. State of California, Department of Human Services. Carcinogen Identification Policy: A statement of science as a basis of policy. Section 2: Methods for estimating cancer risks from exposures to carcinogens (1982).
- 746. Weinstein, I. B. Letter: carcinogen policy at EPA. Science 219: 794-795 (1983).
- 747. Consultative Panel on Health Hazards of Chemical Pesticides. Pest Control (Volume 1): An Assessment of Present and Alter-

- native Technologies, National Academy of Sciences, Washington, DC, 1975.
- 748. Hoel, D. G. Low-dose and species-to-species extrapolation for chemically induced carcinogenesis. In: Cold Spring Harbor Laboratory, Report No. 1: Assessing Chemical Mutagens: The Risk to Humans. Banbury, New York, 1979, pp. 135–145.
- 749. Crouch, E., and Wilson, R. Interspecies comparison of carcinogenic potency. J. Toxicol. Environ. Health 5: 1095-1118 (1978).
- 750. Bernstein, L., Gold, L. S., Ames, B. N., Pike, M. C., and Hoel, D. G. Some tautologous aspects of the comparison of carcinogenic potency in rats and mice. Fund. Appl. Toxicol. 5: 79–86 (1985).
- Whittemore, A. S. The age distribution of human cancer for exposures of varying intensity. Am. J. Epidemiol. 106: 418-432 (1977).
- 752. Hoel, D. G. Comment: carcinogenic risk. Risk Analysis 1: 63-64 (1981).
- 753. Calabrese, E. J. The role of exposure data in standard setting. Toxic Substances J. 4: 12–22 (1982).
- 754. U.S. Environmental Protection Agency. Handbook for Performing Exposure Assessments. Draft, Office of Health and Environmental Assessment, Office of Research and Development, Washington, DC, 1983.
- 755. Aravamudan, K., Bonazountas, M., and Eschenroeder, A. Environmental partitioning model for risk assessment of chemicals: Part 2. Narrative description of the methodology. EPA Contract No. 68-01-3857, Task Order 17, 2 Vol., U.S. Environmental Protection Agency, Washington, DC, 1980.
- tection Agency, Washington, DC, 1980.
 756. Aravamudan, K. Bonazountas, M., and Eschenroeder, A. Environmental Partitioning model for risk assessment of chemicals: Part 2. User Workbook. Contract No. 68-013857, Task Order 17, U.S. Environmental Protection Agency, Washington, DC, 1980.